

Methylmercury in Aquatic Environments

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Contributor: Zivan Gojkovic , Aleksandra Skrobonja , Christiane Funk , Ines Garbayo , Carlos Vélchez

Methylmercury (MeHg) is the most important and the most abundant organic Hg pollutant in the aquatic ecosystem that can affect human health through biomagnification. It is the most toxic organic Hg form, which occurs naturally and by human-induced contamination in water and is further biomagnified in the aquatic food web. MeHg is the only Hg form that accumulates in living organisms and is able to cross the blood–brain barrier, presenting an enormous health risk. Anthropogenic activity increases eutrophication of coastal waters worldwide, which promotes algae blooms. Microalgae, as primary producers, are especially sensitive to MeHg exposure in water and are an important entrance point for MeHg into the aquatic food web. MeHg assimilated by microalgae is further transferred to fish, wildlife and, eventually, humans as final consumers. MeHg biomagnifies and bioaccumulates in living organisms and has serious negative health effects on humans, especially newborns and children.

methylmercury

mercury cycling

microalgae

biomagnification

1. Introduction

Mercury is a naturally occurring element that is highly poisonous to humans and wildlife ^[1]. Exposure to Hg has a number of potential negative effects on human health, including cognitive deficits in children due to fetal exposure, increases in fatal and nonfatal heart attacks and increases in premature death in adults ^[1]. Anthropogenic activities largely contribute to the mobilization of Hg from long-term geologic storage into the biosphere, where Hg cycles between the air, soil and water. While inorganic Hg does not accumulate in living organisms, microbes convert a portion of Hg to methylmercury (MeHg), which is then subjected to further biomagnification in the aquatic food web ^{[1][2]} and causes health risks to humans, especially among populations that frequently consume fish ^[3]. MeHg is bound to fish proteins that form muscle tissue and cannot be removed by cooking. Even the consumption of farmed fish can lead to MeHg exposure because of the presence of MeHg in feed ^[3]. To prevent MeHg accumulation, it is important to understand how MeHg enters at the base of the aquatic food web, where the natural feed for fish is located ^[4]. MeHg is a highly toxic organic Hg form present in natural water ecosystems; it is easily taken up by microalgae and significantly concentrates in the aquatic food web ^[5]. Once accumulated inside the microalgae cell, it is further transferred to grazers, fish larvae, fish, aquatic birds and, eventually, humans as final consumers.

Eutrophication is another phenomenon that can enhance the transfer of MeHg, not only to microalgae, but also to higher trophic levels ^[6]. Eutrophication in coastal areas is a globally expanding problem that can effect an increase in MeHg production and its accumulation in food webs ^[6]. This phenomenon also leads to greater net primary production, which increases organic matter (OM) concentration and decreases light penetration, thereby changing redox conditions in the water column and sediments ^[6].

2. MeHg Interaction with Organic Matter (OM), S and Se Organic Compounds

A key aspect in MeHg bioaccumulation and toxicity is the degree to which MeHg interacts with complexing agents in solution (OH^- , Cl^- , organic sulfur compounds—especially thiols (ligands containing sulfhydryl group R-SH), OM, artificial chelating agents such as EDTA salts) [4][7]. OM that is naturally present in aqueous systems promotes Hg (II) methylation mainly because it contains metabolic substrate molecules essential for heterotrophic microbes and strong ligands that bind Hg(II) [8]. In the case of highly enclosed terrestrial water basins, such as the Baltic Sea, hypoxic and anoxic zones are caused by an increased biological oxygen demand (BOD) due to the excess of nutrients and OM runoffs where Hg binds to allochthonous (terrestrially discharged) OM in the surface layers of the water column and sinks to deeper waters, where it can be released in the microbial remineralization process of OM [9][10][11]. In natural waters where OM and reduced S species are absent, MeHg forms inorganic complexes, MeHgCl and MeHgOH, with chloride and hydroxide ions [12]. The chemical speciation of MeHg prior to cell exposure controls its bioavailability and is a key factor determining the uptake rate and accumulation in microalgae [13][14][15]. MeHg forms stronger complexes with organic thiols, which also have higher stability constants than complexes with OH^- and Cl^- groups [16].

The bioconcentration step of MeHg from water to the base of the food web is crucial for MeHg concentrations found in aquatic organisms higher up in the food web [5][12][17]. Uptake by microalgae is the first and largest step of MeHg bioaccumulation in aquatic food webs, which is influenced by many factors that drive seasonal changes in water MeHg concentrations [8]. OM is an important parameter of MeHg production and uptake by phytoplankton. Measurements from the Bothnian Sea in the Northern Baltic, taken as one of the few studied models of biogeochemical cycling of MeHg in natural aquatic environments, showed that MeHg concentrations can vary by up to an order of magnitude between years (from 103 ± 12 fM in September 2014 to 18 ± 9 fM in August 2016), which underlines the interannual variability in water column MeHg concentrations [8]. Both Hg(II) and MeHg preferentially bind to DOM over inorganic particles, which affects Hg bioavailability [8]. DOM can also decrease light attenuation in water, which decreases MeHg photodegradation. The association between MeHg and dissolved organic carbon (DOC) suggests that labile DOC is the important factor for the remineralization rate and Hg(II) methylation potential [8]. DOM promotes methylation of Hg(II) by (i) stimulating microbial activity and methylation processes, (ii) providing methyl groups for methylation, and (iii) enhancing the solubility of HgS(s) in mineral form [18]. Increased DOM inputs from terrestrial runoffs may lead to higher inputs of Hg and MeHg and increase MeHg formation in the aquatic ecosystem [18].

Hg methylation mainly takes place within 24 h after entering aquatic ecosystems, and to a greater extent under nonequilibrium conditions, before inorganic Hg becomes complexed with substances present in DOM [18]. It is often difficult to characterize DOM, and because ~50% of DOM is organic carbon, it is usually measured and expressed as DOC [14]. Natural levels of DOC vary in different aquatic environments: 4.0 ± 0.02 mg/L in the Baltic Sea; 3.78 ± 1.42 mg/L in Lake Titicaca (Bolivia), with peaks of up to 8 mg/L in some regions; 1.57–17.6 mg/L in the Sacramento River Delta (Sacramento, CA, USA) [14][19][20][21]. Humic matter forms part of allochthonous DOM in seawater [8]. In the case of the Northern Baltic, humic matter concentration decreases from the Bothnian Bay to the Bothnian Sea

and further remains constant around 10 µg/L [8]. Humic content reduces microbial MeHg degradation by decreasing its bioavailability and decreases photodegradation of MeHg by increasing light attenuation [8]. Humic substances can also bind Hg (II) and correlate more strongly than DOC with Hg(II) concentration in water streams. Binding to humic substances makes Hg(II) less available to microbial reduction and methylation [22]. Humic substances are not easily remineralized; thus, an increased proportion of humic matter can decrease the activity of microorganisms that act by chemical reduction and methylation of Hg (II), which further decreases the rates of Hg transformation reactions [8]. In the Northern Baltic Sea, most Hg(II) was available for methylation by forming stable complexes with humic substances or by converting Hg(II) to elemental Hg [8]. The stability constants for Hg–ligand complexes drive metal internalization, thus determining rates of further chemical conversion of Hg species inside the cell. In this sense, it was demonstrated that a Hg–ligand (Hg–L) complex in the culture medium reacts with a biotic ligand (R) at the microalgae cell surface, forming a new complex (Hg–R) prior to metal internalization, and the rate for the formation of complexes at the cell surface is determined by the relative thermodynamic stability constants for Hg–L and Hg–R [13].

Similar to thiols (R–SH), organic selenium also has an affinity towards MeHg and can modify its toxicity by complexation [2]. Selenium belongs to the same group in the periodic table as S and exerts similar chemical affinities, so it can act as a S analogue in amino acids (selenomethionine—SeMet and selenocysteine—SeCys) as well as other organic compounds [23]. Some microalgae strains can take up Se(VI) salts from the medium, biotransform it to organic Se compounds, such as SeMet, which is less toxic to the cell than Se(VI), and accumulate it in the biomass [24][25]. A biologically important Se amino acid—SeMet [23]—significantly inhibits the uptake of MeHg by diatoms and mussels [26]. Detoxification of MeHg in seabirds and marine mammals involves its demethylation by reactive oxygen species (ROS) and the subsequent formation of high-molecular-weight Hg–Se–protein compounds, which are then degraded in lysosomes, forming insoluble Hg–Se compounds [27]. It was demonstrated that fish and wildlife living in environments with elevated Se levels exhibit lower MeHg accumulation [27]. There are a few proposed mechanisms for the protective role of Se against Hg toxicity, such as competition for binding sites or the formation of Hg–Se complexes. It was suggested that the increased Hg(II) uptake in the presence of SeMet is due to the formation of Hg–Se complexes that can be transported across the membrane at a faster rate [26].

3. Biogeochemical Cycling of MeHg and Its Presence in Global Aquatic Ecosystem

Human exposure to MeHg comes predominately from diets containing Hg-contaminated fish and seafood [28][29]. Understanding the cycling of Hg in aquatic systems is essential to assess the environmental risks to human health. Hg cycling differs in freshwater and ocean ecosystems. Hg can be released into the atmosphere both by natural (e.g., volcano eruptions) and anthropogenic sources: mining and burning of fossil fuels [30][31]. However, the more toxic organic form, MeHg, is the dominant species found in rice paddies and fish [32][33][34]. Hg speciation in nature has three different forms: elemental Hg(0), Hg(II) bound to particulate OM and cationic inorganic Hg(II), mainly as

HgCl₂. Elemental Hg(0) travels around the globe, and cationic Hg is transported at intermediate distances, while particulate is not transported very far [35].

MeHg contamination is also associated with long-term emissions from fossil fuel combustion throughout the industrialized world [36]. The burning of fossil fuels, coal, the extraction of gold, smelting and chemical production increase atmospheric Hg input in terrestrial and aquatic ecosystems [37]. The burning of coal is the most abundant anthropogenic source of Hg emissions in the USA [36]. Hg leaching from fossil fuel combustion can contribute to a significant increase in Hg concentration in rivers that enter the sea [28]. Subsequently, Hg(II) reduces to elemental Hg(0), which is usually volatilized to the atmosphere, while a small portion of Hg(II) is converted to MeHg, which is the most toxic form of Hg and acts primarily as a neurotoxin in humans and wildlife [29]. It is known that Hg emissions are ubiquitous, and MeHg as a contaminant bioaccumulates to a great extent in the aquatic food web [35]. It was estimated that bioconcentration factors of MeHg are 10⁶–10⁸ fold from initial water MeHg concentrations, resulting in MeHg levels that are of global toxicological concern [30][35].

Biogeochemical cycling of Hg includes emissions of anthropogenic Hg, transport through the atmosphere, deposition into ocean, subsequent transformation into MeHg [38], and its incorporation into aquatic food webs, which depends on its absorption or modification by microorganisms (**Figure 1a**). Microorganisms perform four types of Hg transformation: reduction of Hg(II) to Hg(0), degradation of organic Hg compounds, methylation of Hg(II) and the oxidation of Hg(0) to Hg(II) [29]. Methylation of Hg occurs in wetlands and lakes as a biological process performed by a range of known heterotrophic microbes (iron- and sulfur-reducing bacteria, methanogens and nitrite oxidizers) [8]. The bulk of MeHg in natural ecosystems originates from methylation of atmospherically deposited Hg by sulfur-reducing bacteria within aquatic sediments [30][36]. Reducing anaerobic conditions within aquatic sediments that support the growth and Hg methylation activity of sulfur-reducing bacteria are in redox potential (Eh) and pH ranges of $0.4 \leq Eh \leq 0.0$ V and $5 \leq pH \leq 10$ [36]. MeHg is predominately formed in hypoxic and anoxic environments from its inorganic form, Hg(II), via biological reactions, and the methylation process has been mostly linked to the presence of sulfate- and iron-reducing bacteria [39]. However, more recently, methanogens and other microbes have been found to play important roles in the MeHg formation process [40][41][42][43][44].

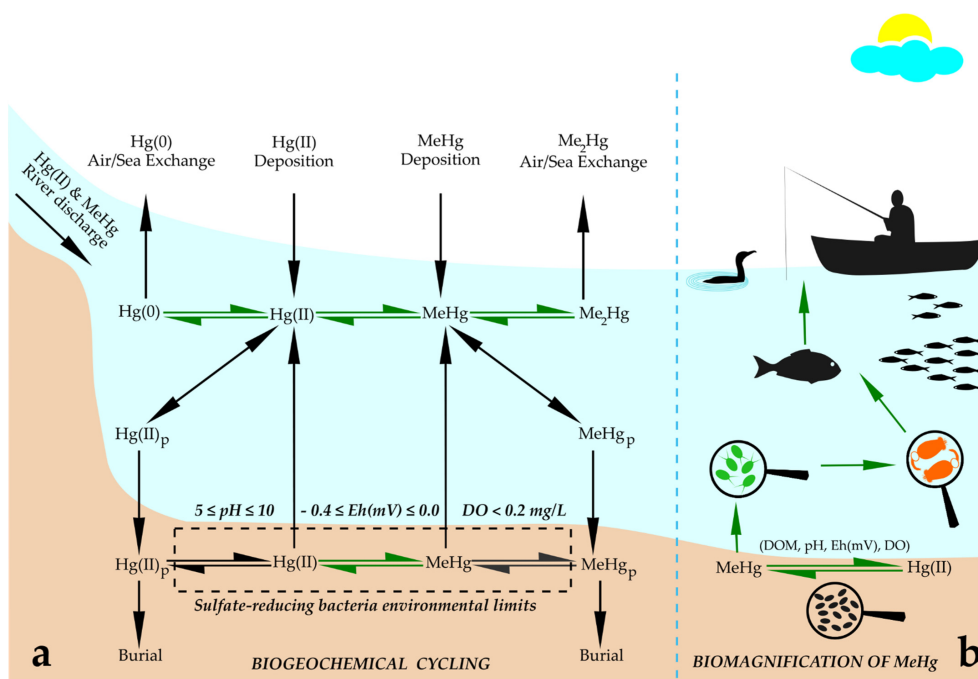


Figure 1. Schematic representation of (a) basic processes in biogeochemical cycling of Hg in coastal areas. Abbreviations: Hg(0) , elemental mercury; Hg(II) , divalent mercury; MeHg , methylmercury; Me_2Hg , dimethylmercury; MeHg_p , methylmercury bound to particulate organic matter; Hg(II)_p , divalent mercury bound to particulate organic matter. Green arrows indicate biologically mediated processes; environmental conditions for sulfate-reducing bacteria, where Hg methylation occurs, are $5 \leq \text{pH} \leq 10$; $-0.4 \leq \text{Eh(mV)} \leq 0.0$; $\text{DO} < 0.2 \text{ mg/L}$. (b) Biomagnification of MeHg through the aquatic food web: MeHg enters the food web at the very bottom via bacterial methylation activity, then it is partially taken up by microalgae; from that point on, it biomagnifies through rotifers that graze on microalgae, and further via smaller fish that prey on rotifers, then predator fish that prey on smaller fish, and finally transfers to aquatic birds and humans that consume contaminated fish. Abbreviations: Hg(II) , divalent mercury; MeHg , methylmercury; DOM, dissolved organic matter; Eh(mV), redox potential; DO, dissolved oxygen. Adapted from [6][30][35][36][45].

Microalgae play a direct seasonal role in Hg chemical species circulation in aquatic environments [8]. As already explained above, MeHg enters the food web via microalgae and further bioaccumulates in aquatic organisms, which can finally result in biomagnification to concentrations with adverse effects on reproduction and neural development in fish and mammals [46]. The biggest MeHg increase in water and microalgae biomass in the Baltic Sea was found in summer and fall, with 60–70% increase in the intracellular MeHg content of algae biomass compared to a yearly average [6]. It was suggested that this occurs due to the remineralization of dead algae biomass from the spring and summer blooms, which results in higher direct methylation and uptake by microalgae [6]. It was found that the maximal impact of eutrophication on the intracellular MeHg content coincides with the highest zooplankton grazing on algae biomass [6]. Extreme MeHg levels in natural water bodies were reported for the Olt River (Romania) and its three reservoirs, which are heavily contaminated with Hg from a nearby chlor-alkali plant [47]. Total Hg concentrations in the Olt River were up to 88 ng/L but decreased by 10 times inside the reservoir (to 8 ng/L) [47]. MeHg concentrations increase from the entrance to the reservoir to the central part to 0.7 ng/L,

confirming high methylation within the eutrophic reservoir [47]. Based on the vertical water column profiles of total Hg and MeHg, it was suggested that Hg methylation mainly occurs deep in the water column, close to the bottom, at the sediment–water interface. MeHg concentrations and bioconcentration factors of phytoplankton confirm that the highest biomagnification of MeHg takes place in the primary producers [47]. High MeHg concentrations (>15 ng/L) in the surface water of some artificial wetlands and its accumulation in saturated sediments can affect the invertebrate community to levels that are potentially hazardous for insectivorous biota such as birds [48].

Similar to freshwater systems, Hg(II) can be deposited to the ocean by dry and wet deposition, and elemental Hg(0) by dry deposition, where wet deposition is defined as removal from the atmosphere through precipitation, while dry deposition is considered to be Hg settling or uptake without precipitation [29][30]. The total Hg deposition (wet and dry) to oceans was estimated to be 10 Mmol/year [29]. A significant load of Hg is introduced into the sea by rivers, with a high amount of Hg contributed by river sediment [28]. The load of Hg introduced with suspended and trailing river debris leads to Hg enrichment of the marine coastal zone [28]. The Hg(II) deposited to the ocean can be reduced to Hg(0), adsorbed onto particles or methylated [30]. The general trend is that oxygen-depleted areas are spreading in coastal and offshore waters worldwide [9][49]. Seawater can be considered as normoxic, with a normal dissolved oxygen (DO) concentration of 5–8 mg/L; hypoxic (DO < 0.2 mg/L); or anoxic, with the presence of hydrogen sulfide and a DO < 0.1 mg/L [6][50]. One of the main processes responsible for a DO decrease is O₂ consumption by bacteria during the decomposition and mineralization of OM [50]. Excessive eutrophication negatively affects the oxygenation of seawater as the enrichment of water with nutrients such as N, P and DOC in the surface layer results in increased phytoplankton growth and biomass production, which causes algae blooms and reduces DO and water transparency [50]. Total Hg content in marine ecosystems is in the picomolar (pM) range and depends on the position, size and exposure to anthropogenic activity of the aquatic ecosystem (Table 1). MeHg accounts for 3–35% of total Hg levels in the oceans, and its concentration in water is usually in femtomolar range (fM) [8][30][46].

Table 1. Concentrations of total Hg and MeHg in different aquatic ecosystems according to the literature. Data are presented in the original units (ng/L, pM and fM) that were used by the respective authors.

Location	Total Hg ¹ pM or ng/L	MeHg pM, fM or ng/L	References
-	-	-	-
Southern Baltic Sea	1.5 ± 0.7 pM	23 ± 13 fM	[9]
Northern Baltic Sea	1.0 ± 0.3 pM	37 ± 15 fM; 21 ± 9 fM	[6][9]
Bothnian Bay (Baltic)	1.24 ± 0.3 pM	80 ± 25 fM	[8]
Bothnian Sea (Baltic)	0.84 ± 0.24 pM	21 ± 9 fM	[8]
Bothnian Bay (Baltic)	11.5 ± 1.66 pM	116–236 fM	[18]
Atlantic Ocean (Southern Polar Front)	0.93 ± 0.69 ng/L	0.26 ± 0.12 ng/L	[51]

Location	Total Hg ¹ pM or ng/L	MeHg pM, fM or ng/L	References
-	-	-	-
Jiaozhou Bay (Yellow Sea)	8.46–27.3 ng/L	0.08–0.83 ng/L	[52]
Yellow Sea	6.7–27.5 pM	— ²	[53]
South China Sea	0.8–2.3 ng/L	0.05–0.22 ng/L	[54]
North Atlantic Ocean	2.4 pM	—	[55]
Pacific Ocean	1.2 pM	—	[56]
Mediterranean Sea	1.0 pM; 2.5 pM	—	[38][57]
Average in oceans	1.5 pM	—	[58]
Lake Titicaca (Bolivia)	—	0.01–0.18 ng/L	[21]
Olt River (Romania)	8–88 ng/L	0.7 ng/L	[47]
Râne River estuary (Baltic)	2.0–5.95 pM	306 fM	[8]
Wetlands in Rouge Park, Canada	1.45 ± 0.91 ng/L	0.59 ± 0.45 ng/L	[48]
Lakes in Oil Sands Region, Canada	0.4–5.3 ng/L	0.01–0.34 ng/L	[37]
Lake Victoria, Africa	3–15 ng/L	—	[59]
Average in surface water of lakes and rivers	—	0.003–1.03 ng/L	[39]

ertheless, new evidence has led to the hypothesis that MeHg can also be formed in oxic brackish coastal waters. It was demonstrated that the concentration of MeHg in experimental microcosms containing oxic surface waters from the pelagic zone of the Bothnian Sea significantly increased within 27 h after the addition of 250 pM Hg(II) and under normal DOC concentration (4.0 mg/L), where bacterial activity played a central role in MeHg production [18]. It was also found that enhanced MeHg production was not reflected by substantial changes in the bacterial community composition or gene pool [18].

It was found that the contribution of MeHg in total Hg in Canadian lakes is strongly and positively correlated with turbidity and total dissolved N and slightly, more weakly correlated with chlorophyll-a and DOC, which suggests that Hg methylation rates are higher in lakes with greater productivity and algae biomass, where increased oxygen use by decomposers in sediments receiving high deposition rates of labile OM and related anoxia in sediments would be expected to drive Hg methylation rates [37]. Although the basic parameters driving the biogeochemical cycling of Hg are known, in-lake influences on these parameters and Hg cycling are complex and highly variable and can mask climate- or industry-related changes over time [37].

References

1. Swain, E.B.; Jakus, P.M.; Rice, G.; Lupi, F.; Maxson, P.A.; Pacyna, J.M.; Penn, A.; Spiegel, S.J.; Veiga, M.M. Socioeconomic Consequences of Mercury Use and Pollution. *Ambio* 2007, 36, 45–61.
2. Le Faucheur, S.; Campbell, P.G.C.; Fortin, C.; Slaveykova, V.I. Interactions between Mercury and Phytoplankton: Speciation, Bioavailability, and Internal Handling. *Environ. Toxicol. Chem.* 2014, 33, 1211–1224.
3. Mergler, D.; Anderson, H.A.; Chan, L.H.M.; Mahaffey, K.R.; Murray, M.; Sakamoto, M.; Stern, A.H. Methylmercury Exposure and Health Effects in Humans: A Worldwide Concern. *Ambio* 2007, 36, 3–11.
4. Pickhardt, P.C.; Fisher, N.S. Accumulation of Inorganic and Methylmercury by Freshwater Phytoplankton in Two Contrasting Water Bodies. *Environ. Sci. Technol.* 2007, 41, 125–131.
5. Moyer, H.A.; Miles, C.J.; Philips, E.J.; Sargent, B.; Merritt, K.K. Kinetics and Uptake Mechanisms for Monomethylmercury between Freshwater Algae and Water. *Environ. Sci. Technol.* 2002, 36, 3550–3555.
6. Soerensen, A.L.; Scharup, A.T.; Gustafsson, E.; Gustafsson, B.G.; Undeman, E.; Björn, E. Eutrophication Increases Phytoplankton Methylmercury Concentrations in a Coastal Sea—A Baltic Sea Case Study. *Environ. Sci. Technol.* 2016, 50, 11787–11796.
7. Gorski, P.R.; Armstrong, D.E.; Hurley, J.P.; Krabbenhoft, D.P. Influence of Natural Dissolved Organic Carbon on the Bioavailability of Mercury to a Freshwater Alga. *Environ. Pollut.* 2008, 154, 116–123.
8. Soerensen, A.L.; Scharup, A.T.; Skrobonja, A.; Björn, E. Organic Matter Drives High Interannual Variability in Methylmercury Concentrations in a Subarctic Coastal Sea. *Environ. Pollut.* 2017, 229, 531–538.
9. Soerensen, A.L.; Scharup, A.T.; Skrobonja, A.; Bouchet, S.; Amouroux, D.; Liem-Nguyen, V.; Björn, E. Deciphering the Role of Water Column Redoxclines on Methylmercury Cycling Using Speciation Modeling and Observations from the Baltic Sea. *Glob. Biogeochem. Cycles* 2018, 32, 1498–1513.
10. Deutsch, B.; Alling, V.; Humborg, C.; Korth, F.; Mörtz, C.M. Tracing Inputs of Terrestrial High Molecular Weight Dissolved Organic Matter within the Baltic Sea Ecosystem. *Biogeosciences* 2012, 9, 4465–4475.
11. Ripszam, M.; Paczkowska, J.; Figueira, J.; Veenaas, C.; Haglund, P. Dissolved Organic Carbon Quality and Sorption of Organic Pollutants in the Baltic Sea in Light of Future Climate Change. *Environ. Sci. Technol.* 2015, 49, 1445–1452.
12. Miles, C.J.; Moyer, H.A.; Philips, E.J.; Sargent, B. Partitioning of Monomethylmercury between Freshwater Algae and Water. *Environ. Sci. Technol.* 2001, 35, 4277–4282.

13. Skrobonja, A.; Gojkovic, Z.; Soerensen, A.L.; Westlund, P.-O.; Funk, C.; Björn, E. Uptake Kinetics of Methylmercury in a Freshwater Alga Exposed to Methylmercury Complexes with Environmentally Relevant Thiols. *Environ. Sci. Technol.* 2019, 53, 13757–13766.
14. Luengen, A.C.; Fisher, N.S.; Bergamaschi, B.A. Dissolved Organic Matter Reduces Algal Accumulation of Methylmercury. *Environ. Toxicol. Chem.* 2012, 31, 1712–1719.
15. Ndu, U.; Mason, R.P.; Zhang, H.; Lin, S.; Visscher, P.T. Effect of Inorganic and Organic Ligands on the Bioavailability of Methylmercury as Determined by Using a Mer-Lux Bioreporter. *Appl. Environ. Microbiol.* 2012, 78, 7276–7282.
16. Berthon, G. Critical Evaluation of the Stability Constants of Metal Complexes of Amino Acids with Polar Side Chains (Technical Report). *Pure Appl. Chem.* 1995, 67, 1117–1240.
17. Mason, R.P.; Reinfelder, J.R.; Morel, F.M.M. Uptake, Toxicity, and Trophic Transfer of Mercury in a Coastal Diatom. *Environ. Sci. Technol.* 1996, 30, 1835–1845.
18. Rodríguez, J.; Andersson, A.; Björn, E.; Timonen, S.; Brugel, S.; Skrobonja, A.; Rowe, O. Inputs of Terrestrial Dissolved Organic Matter Enhance Bacterial Production and Methylmercury Formation in Oxidic Coastal Water. *Front. Microbiol.* 2022, 13, 809166.
19. Andersson, A.; Brugel, S.; Paczkowska, J.; Rowe, O.F.; Figueroa, D.; Kratzer, S.; Legrand, C. Influence of Allochthonous Dissolved Organic Matter on Pelagic Basal Production in a Northernly Estuary. *Estuar. Coast. Shelf Sci.* 2018, 204, 225–235.
20. Quiroga-Flores, R.; Guédron, S.; Achá, D. High Methylmercury Uptake by Green Algae in Lake Titicaca: Potential Implications for Remediation. *Ecotoxicol. Environ. Saf.* 2021, 207, 111256.
21. Achá, D.; Guédron, S.; Amouroux, D.; Point, D.; Lazzaro, X.; Fernandez, P.E.; Sarret, G. Algal Bloom Exacerbates Hydrogen Sulfide and Methylmercury Contamination in the Emblematic High-Altitude Lake Titicaca. *Geosciences* 2018, 8, 438.
22. Scharf, A.T.; Ndu, U.; Balcom, P.H.; Mason, R.P.; Sunderland, E.M. Contrasting Effects of Marine and Terrestrially Derived Dissolved Organic Matter on Mercury Speciation and Bioavailability in Seawater. *Environ. Sci. Technol.* 2015, 49, 5965–5972.
23. Gojkovic, Ž.; Garbayo, I.; Ariza, J.L.G.; Márová, I.; Vílchez, C. Selenium Bioaccumulation and Toxicity in Cultures of Green Microalgae. *Algal Res.* 2015, 7, 106–116.
24. Umyšová, D.; Vítová, M.; Doušková, I.; Bišová, K.; Hlavová, M.; Čížková, M.; Machát, J.; Doucha, J.; Zachleder, V. Bioaccumulation and Toxicity of Selenium Compounds in the Green Alga *Scenedesmus Quadricauda*. *BMC Plant Biol.* 2009, 9, 58.
25. Gojkovic, Ž.; Vílchez, C.; Torronteras, R.; Vigara, J.; Gómez-Jacinto, V.; Janzer, N.; Garbayo, I. Effect of Selenate on Viability and Selenomethionine Accumulation of *Chlorella sorokiniana* Grown in Batch Culture. *Sci. World J.* 2014, 2014, 13.

26. Wang, W.-X.; Wong, R.S.K.; Wang, J.; Yen, Y. Influences of Different Selenium Species on the Uptake and Assimilation of Hg(II) and Methylmercury by Diatoms and Green Mussels. *Aquat. Toxicol.* 2004, 68, 39–50.
27. Scheuhammer, A.M.; Meyer, M.W.; Sandheinrich, M.B.; Murray, M.W. Effects of Environmental Methylmercury on the Health of Wild Birds, Mammals, and Fish. *Ambio* 2007, 36, 12–18.
28. Gębka, K.; Bełdowska, M. Variation in the Content of Different Forms of Mercury in River Catchments of the Southern Baltic Sea—Case Study. *Pol. Hyperb. Res.* 2020, 72, 63–72.
29. Gworek, B.; Bemowska-Kalabun, O.; Kijeńska, M.; Wrzosek-Jakubowska, J. Mercury in Marine and Oceanic Waters—A Review. *Water Air Soil Pollut.* 2016, 227, 371.
30. Selin, N.E. Global Biogeochemical Cycling of Mercury: A Review. *Annu. Rev. Environ. Resour.* 2009, 34, 43–63.
31. Malm, O.; Pfeiffer, W.C.; Souza, C.M.M.; Reuther, R. Mercury Pollution Due to Gold Mining in the Madeira River Basin, Brazil. *AMBIO A J. Hum. Environ.* 1990, 19, 11–15.
32. Zhang, H.; Feng, X.; Larssen, T.; Qiu, G.; Vogt, R.D. In Inland China, Rice, Rather than Fish, Is the Major Pathway for Methylmercury Exposure. *Environ. Health Perspect.* 2010, 118, 1183–1188.
33. United States Environmental Protection Agency, US EPA. Biennial National Listing of Fish Advisories Fact Sheet EPA-823-F-09-007 September 2009. Available online: <https://www.epa.gov/sites/default/files/2019-05/documents/2008-biennial-nlfa.pdf> (accessed on 13 August 2022).
34. Jiang, G.-B.; Shi, J.-B.; Feng, X.-B. Mercury Pollution in China. An Overview of the Past and Current Sources of the Toxic Metal. *Environ. Sci. Technol.* 2006, 40, 3673–3678.
35. Krabbenhoft, D. Methylmercury Contamination of Aquatic Ecosystems: A Widespread Problem with Many Challenges for the Chemical Sciences; Norling, P., Wood-Black, F., Masciangioli, T., Eds.; Water and Sustainable Development: Opportunities for the Chemical Sciences—A Workshop Report to the Chemical Sciences Roundtable; National Academy of Sciences: Washington, DC, USA, 2004; ISBN 0-309-53173-X.
36. Sams, C.E. Methylmercury Contamination: Impacts on Aquatic Systems and Terrestrial Species, and Insights for Abatement. In *Advancing the Fundamental Sciences: Proceedings of the Forest Service National Earth Sciences Conference*, San Diego, CA, USA, 18–22 October 2004; Furniss, M., Clifton, C., Ronnenberg, K., Eds.; U.S. Department of Agriculture, Forest Service, Pacific Northwest Research Station: San Diego, CA, USA, 2007.
37. Emmerton, C.A.; Cooke, C.A.; Wentworth, G.R.; Graydon, J.A.; Ryjkov, A.; Dastoor, A. Total Mercury and Methylmercury in Lake Water of Canada's Oil Sands Region. *Environ. Sci. Technol.* 2018, 52, 10946–10955.

38. Cinnirella, S.; Bruno, D.E.; Pirrone, N.; Horvat, M.; Živković, I.; Evers, D.C.; Johnson, S.; Sunderland, E.M. Mercury Concentrations in Biota in the Mediterranean Sea, a Compilation of 40 Years of Surveys. *Sci. Data* 2019, 6, 205.
39. Bravo, A.G.; Peura, S.; Buck, M.; Ahmed, O.; Mateos-Rivera, A.; Ortega, S.H.; Schaefer, J.K.; Bouchet, S.; Tolu, J.; Björn, E.; et al. Methanogens and Iron-Reducing Bacteria: The Overlooked Members of Mercury-Methylating Microbial Communities in Boreal Lakes. *Appl. Environ. Microbiol.* 2018, 84, e01774-18.
40. Compeau, G.C.; Bartha, R. Sulfate-Reducing Bacteria: Principal Methylators of Mercury in Anoxic Estuarine Sediment. *Appl. Environ. Microbiol.* 1985, 50, 498–502.
41. Kerin, E.J.; Gilmour, C.C.; Roden, E.; Suzuki, M.T.; Coates, J.D.; Mason, R.P. Mercury Methylation by Dissimilatory Iron-Reducing Bacteria. *Appl. Environ. Microbiol.* 2006, 72, 7919–7921.
42. Gilmour, C.C.; Podar, M.; Bullock, A.L.; Graham, A.M.; Brown, S.D.; Somenahally, A.C.; Johs, A.; Hurt, R.A.; Bailey, K.L.; Elias, D.A. Mercury Methylation by Novel Microorganisms from New Environments. *Environ. Sci. Technol.* 2013, 47, 11810–11820.
43. Parks, J.M.; Johs, A.; Podar, M.; Bridou, R.; Hurt, R.A.J.; Smith, S.D.; Tomanicek, S.J.; Qian, Y.; Brown, S.D.; Brandt, C.C.; et al. The Genetic Basis for Bacterial Mercury Methylation. *Science* 2013, 339, 1332–1335.
44. Yu, R.-Q.; Reinfelder, J.R.; Hines, M.E.; Barkay, T. Mercury Methylation by the Methanogen *Methanospirillum hungatei*. *Appl. Environ. Microbiol.* 2013, 79, 6325–6330.
45. Engstrom, D.R. Fish Respond When the Mercury Rises. *Proc. Natl. Acad. Sci. USA* 2007, 104, 16394–16395.
46. Canário, J.; Vale, C.; Poissant, L.; Nogueira, M.; Pilote, M.; Branco, V. Mercury in Sediments and Vegetation in a Moderately Contaminated Salt Marsh (Tagus Estuary, Portugal). *J. Environ. Sci.* 2010, 22, 1151–1157.
47. Bravo, A.G.; Cosio, C.; Amouroux, D.; Zopfi, J.; Chevalley, P.-A.; Spangenberg, J.E.; Ungureanu, V.-G.; Dominik, J. Extremely Elevated Methyl Mercury Levels in Water, Sediment and Organisms in a Romanian Reservoir Affected by Release of Mercury from a Chlor-Alkali Plant. *Water Res.* 2014, 49, 391–405.
48. Sinclair, K.A.; Xie, Q.; Mitchell, C.P.J. Methylmercury in Water, Sediment, and Invertebrates in Created Wetlands of Rouge Park, Toronto, Canada. *Environ. Pollut.* 2012, 171, 207–215.
49. Breitburg, D.; Levin, L.A.; Oschlies, A.; Grégoire, M.; Chavez, F.P.; Conley, D.J.; Garçon, V.; Gilbert, D.; Gutiérrez, D.; Isensee, K. Declining Oxygen in the Global Ocean and Coastal Waters. *Science* 2018, 359, eaam7240.

50. Rak, D.; Walczowski, W.; Dzierzbicka-Głowacka, L.; Shchuka, S. Dissolved Oxygen Variability in the Southern Baltic Sea in 2013–2018. *Oceanologia* 2020, 62, 525–537.
51. Canário, J.; Santos-Echeandia, J.; Padeiro, A.; Amaro, E.; Strass, V.; Klaas, C.; Hoppema, M.; Ossebaar, S.; Koch, B.P.; Laglera, L.M. Mercury and Methylmercury in the Atlantic Sector of the Southern Ocean. *Deep Sea Res. Part II Top. Stud. Oceanogr.* 2017, 138, 52–62.
52. Mao, L.; Liu, X.; Wang, B.; Lin, C.; Xin, M.; Zhang, B.-T.; Wu, T.; He, M.; Ouyang, W. Occurrence and Risk Assessment of Total Mercury and Methylmercury in Surface Seawater and Sediments from the Jiaozhou Bay, Yellow Sea. *Sci. Total Environ.* 2020, 714, 136539.
53. Ci, Z.; Zhang, X.; Wang, Z.; Niu, Z. Phase Speciation of Mercury (Hg) in Coastal Water of the Yellow Sea, China. *Mar. Chem.* 2011, 126, 250–255.
54. Fu, X.; Feng, X.; Zhang, G.; Xu, W.; Li, X.; Yao, H.; Liang, P.; Li, J.; Sommar, J.; Yin, R.; et al. Mercury in the Marine Boundary Layer and Seawater of the South China Sea: Concentrations, Sea/Air Flux, and Implication for Land Outflow. *J. Geophys. Res. Atmos.* 2010, 115, 1–11.
55. Mason, R.P.; Rolfhus, K.R.; Fitzgerald, W.F. Mercury in the North Atlantic. *Mar. Chem.* 1998, 61, 37–53.
56. Laurier, F.J.G.; Mason, R.P.; Gill, G.A.; Whalin, L. Mercury Distributions in the North Pacific Ocean —20 Years of Observations. *Mar. Chem.* 2004, 90, 3–19.
57. Cossa, D.; Martin, J.-M.; Takayanagi, K.; Sanjuan, J. The Distribution and Cycling of Mercury Species in the Western Mediterranean. *Deep Sea Res. Part II Top. Stud. Oceanogr.* 1997, 44, 721–740.
58. Lamborg, C.H.; Fitzgerald, W.F.; Damman, A.W.H.; Benoit, J.M.; Balcom, P.H.; Engstrom, D.R. Modern and Historic Atmospheric Mercury Fluxes in Both Hemispheres: Global and Regional Mercury Cycling Implications. *Glob. Biogeochem. Cycles* 2002, 16, 51-1–51-11.
59. Ramlal, P.S.; Bugenyi, F.W.B.; Kling, G.W.; Nriagu, J.O.; Rudd, J.W.M.; Campbell, L.M. Mercury Concentrations in Water, Sediment, and Biota from Lake Victoria, East Africa. *J. Great Lakes Res.* 2003, 29, 283–291.

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