

# Electronic Noses in COPD

Subjects: Oncology

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Exhaled breath analysis is a non-invasive method to study lung diseases, and electronic noses have been extensively used in breath research.

Studies with electronic noses have proved that the pattern of exhaled volatile organic compounds is different in COPD.

More recent investigations have reported that electronic noses could potentially distinguish different endotypes (i.e., neutrophilic vs. eosinophilic) and are able to detect microorganisms in the airways responsible for exacerbations.

This entry reviews the published literature on electronic noses and COPD and help in identifying methodological, physiological, and disease-related factors which could affect the results.

Keywords: COPD ; electronicnose ; VOCs

## 1. Introduction

Chronic obstructive pulmonary disease (COPD) is a common disorder of the respiratory system which is characterised by a progressive airflow limitation caused by exposure to noxious particles, usually tobacco smoke, in susceptible individuals<sup>[1]</sup>. However, other factors, such as premature birth, frequent childhood infections, asthma, or passive smoking, could also contribute to COPD [1]. The disease may affect the large airways, respiratory bronchioles, and lung parenchyma, however the extent of the involvement of different lung regions may vary<sup>[2]</sup> (Figure 1).

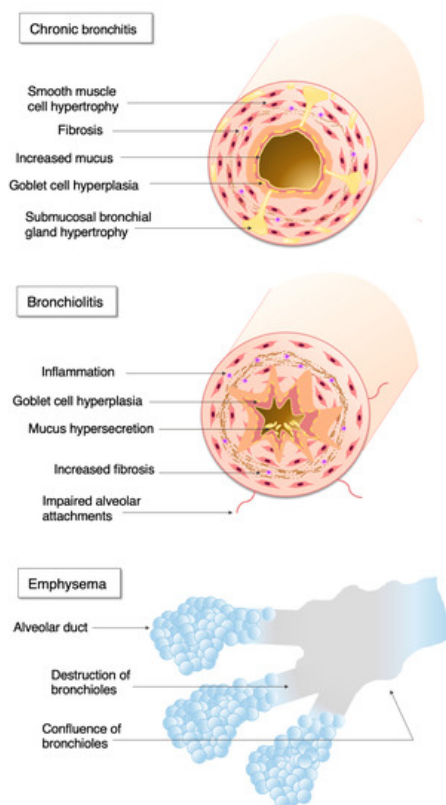


Figure 1. The pathophysiology of chronic obstructive pulmonary disease.

Large airway disease is characterised by mucus hypersecretion, ciliary and epithelial dysfunction, mucosal and submucosal inflammation, as well as enhanced bronchial blood flow. Patients may present with symptoms of chronic productive cough or chronic bronchitis. Most of these patients have small airway disease, which is characterised by airway inflammation, peribronchial fibrosis, and subsequent small airway narrowing. Parenchymal involvement is termed

emphysema, and it is characterised by progressive loss of the lung tissue, impaired oxygen intake, and carbon dioxide removal. People with small airway disease and emphysema often complain of progressive shortness of breath. Although widely recognised as a progressive disease, the activity of disease varies largely between patients. Around half of patients have a rapid ( $\geq 50$  mL/year loss) decline in forced expiratory volume in the first second (FEV<sub>1</sub>), a marker quantifying airway obstruction<sup>[3]</sup>, and around 30% are prone to acute exacerbations, major events leading to health deterioration and associated with high healthcare burden and mortality<sup>[4]</sup>.

COPD is diagnosed based on medical history, symptoms, and lung function showing fixed airflow obstruction. Although the diagnosis, especially the differential diagnosis from other lung diseases (i.e., asthma, bronchiectasis), is sometimes difficult, in most cases it can be made based on simple and cheap pulmonary function tests. It is important to have reliable biomarkers which could differentiate patients with eosinophilic airway inflammation and reflect on disease activity (i.e., predict lung function decline and future exacerbations). This is essential clinical information, as inhaled corticosteroids (ICS) seem to be more effective in patients with raised airway eosinophils <sup>[5]</sup>, as well as patients with a high exacerbation burden<sup>[6]</sup>. On the other hand, in some patients recurrent exacerbations are maintained by colonising bacteria and patients may benefit from prophylactic antibiotic treatment<sup>[7]</sup>. Hence, biomarkers reflecting on bacterial colonisation and specifying bacteria would have significant clinical value. Similar to stable disease, acute exacerbations are also heterogeneous and patients may benefit from tailored treatment depending on the inflammatory profile<sup>[8]</sup> and infectious cause<sup>[9]</sup>.

Exhaled breath analysis is a widely used technique for investigating airway diseases.<sup>[10]</sup> It is totally harmless, and therefore can be performed even in very frail patients and during acute breathlessness, such as in exacerbation. Therefore, it has a great yet not fully explored clinical potential to distinguish patients with different inflammatory endotypes and airway microbiology. One of the most important limiting factors is the lack of standardisation<sup>[11]</sup> and the effect of various endogenous (airway calibre, comorbidities, etc.) and exogenous factors (diet, smoking, pollution) which may limit their use. Traditionally, techniques assessing breath biomarkers are divided into methods investigating volatile and non-volatile particles<sup>[11]</sup> and the measurement of breath temperature<sup>[12]</sup>. In this review, we will focus on the measurement of volatile organic compounds (VOCs) using electronic noses in COPD.

## 2. Electronic Nose Studies in COPD

As described above, the composition of exhaled VOCs in could be altered due to several endogenous and exogenous factors. This chapter summarises the published evidence for case-control studies (Table 1). First of all, it has to be emphasised that the electronic nose signal in COPD seems to be stable, with a within-day reproducibility of 0.80 and an overall mean between-day reproducibility around 0.70<sup>[13][14][15]</sup>

Table 1. Clinical studies conducted on electronic noses in patients with COPD.

Comparator Group	Device	Number of Subjects	Classification Technique	Sensitivity (%)	Specificity (%)	Cross-Validation Value (%)	Remarks	Reference
Healthy	Cyrano 320	N = 37 COPD N = 13 H	LDA	83	76	79	COPD vs. H	[16]
		N = 74 ECOPD N = 19		72	67		ECOPD vs. COPD	
	Cyrano 320	ECOPD + P N = 50 COPD N = 30 H	LDA	88	75	ND	ECOPD + P vs. COPD	[17]
Infection		N = 50 COPD N = 30 H		91	75		ECOPD + P vs. ECOPD	
	Aeonose	N = 22 COPD + BI N = 21 COPD without BI		73	76		COPD + VI vs. COPD without VI	
		N = 18 COPD + VI N = 25 COPD without VI	ANN	83	72	ND	COPD + BI vs. COPD without BI	[18]

Comparator Group	Device	Number of Subjects	Classification Technique	Sensitivity (%)	Specificity (%)	Cross-Validation Value (%)	Remarks	Reference
References								
1.		N = 10 LC N = 10 COPD N = 10 H	LDA	ND	ND	88	LC vs. H	[20]
2.	Cyanose	N = 320	ROC analysis	80	48	ND	Diagnostic accuracy increased when combined with sputum	[20]
3.	Cancer	N = 31 COPD	principal components	ND	ND	ND	Diagnosis	[21]
4.		N = 49 COPD	Random	73	72	ND	LC	[21]
5.	Custom made colorimetric sensor	N = 15 forest method		73	72	ND	LC	[21]
6.		N = 88 COPD + S vs. H		ND	98.1	100	COPD + S vs. H	[22]
7.	Smoking Cyanose	N = 28 COPD + HAP N = 178	LDA + SVM	ND	97.5	100	COPD + HAP vs. H	[22]
8.		N = 31 COPD		ND	ND	78	COPD vs. H	[23]
9.	Asthma and lung cancer	N = 37 COPD N = 31	LDA	ND	ND	80	COPD vs. LC	[23]
10.		N = 20 A N = 30		ND	ND	87	A vs. H	[24]
11.	Smoking	N = 20 COPD N = 20 H		ND	ND	88	LC vs. H	[24]
12.		N = 40 COPD N = 60 A	LDA	91	90	83	COPD vs. reversible A (N = 39)	[24]
13.		N = 115		ND	ND	NS	COPD vs. non-S	[25]
14.	SpiroNose	N = 206	performed	ND	ND	NS	combined asthma and COPD clusters	[25]
15.								
16.								

- | Comparator Group  | Electronic Nose | Number of Subjects | Classification Technique | Sensitivity (%) | Specificity (%) | Cross-Validation Value (%) | Reference |
|---|-----------------|--------------------|--------------------------|-----------------|-----------------|----------------------------|-----------|
| 17. Shafiek, H.; Fiorentino, F.; Merino, J.L.; López, C.; Oliver, A.; Segura, J.; de Paulis, J.; Sibila, O.; Agustí, A.; Cosío, B.G.                        | Cyanose         | N = 15             | LDA                      | ND              | ND              | 82.1                       | [26]      |
| 18. Van Geffen, W.H.; Bruins, M.T.S.; Kersjens, H.A.  | Cyanose         | N = 13             | LDA                      | ND              | ND              | 82.1                       | [26]      |
| 19. Dragonieri, S.; Annema, J.T.; Vos, R.; van der Schee, M.P.; Spanevello, A.; Cariani, P.; Rocco, G.B.; K.F.; Sterk, P.J.                                 | OSA             | N = 20             | LDA                      | ND              | ND              | 82.1                       | [26]      |
| 20. Hubers, A.J.; Brinkman, P.; Boksem, R.J.; Rhodius, R.J.; Witte, B.I.; Zwinderman, A.H.; Heideman, D.A.; Duin, S.; Koning, R.; Steenbergen, R.D.; et al. | Custom          | N = 20             | PLS-DA                   | 44              | 93              | ND                         | [27]      |
| 21. Mazzone, P.J.; Hammel, J.; Fennell, R.; Na, J.; Czich, C.; Laskowski, D.; Mekhail, T.   | Custom          | N = 20             | PLS-DA                   | 44              | 93              | ND                         | [27]      |
| 22. Rodríguez-Aguilar, M.; Díaz de León-Martínez, L.; Gorocica-Rosete, P.; Padilla, R.P.; Thirión-Romero, I.; Ornelas-Ramírez, O.; Flores-Ramírez, R.       | Cyanose         | N = 23             | LDA                      | ND              | ND              | 82.1                       | [28]      |
| 23. De Vries, R.; Brinkman, P.; van der Schee, M.P.; Fens, N.; Dijkers, E.; Bootsma, S.K.; de Jongh, F.H.; Sterk, P.J.                                      | AAT             | N = 10             | LDA                      | ND              | ND              | 82.1                       | [28]      |
| 24. Fens, N.; Roldaan, A.C.; van der Schee, M.P.; Boksem, R.J.; Zwinderman, A.H.; de Jongh, F.H.; Sterk, P.J.   | Cyanose         | N = 117            | PLS-DA                   | 63              | 74              | ND                         | [29]      |
| 25. de Vries, R.; Dagelet, Y.W.F.; Spoor, P.; Snoey, E.; Jak, P.M.C.; Brinkman, P.; Dijkers, E.; Bootsma, S.K.; Elskamp, F.; de Jongh, F.H.C.; et al.       | Cyanose         | N = 117            | PLS-DA                   | 63              | 74              | ND                         | [29]      |
| 26. de Jongh, F.H.C.; et al.  | Cyanose         | N = 117            | PLS-DA                   | 63              | 74              | ND                         | [29]      |
| 27. Scarlata, S.; Pennazza, G.; Santonico, M.; Samanegro, S.; Rossi, B.; Banti, I.; Rivera, C.; Vernile, C.; De Vincenzi, A.; Antonelli, I.; et al.         | Cyanose         | N = 117            | PLS-DA                   | 63              | 74              | ND                         | [29]      |
| 28. Hattesohl, A.D.; Jörres, R.A.; Dressel, H.; Schmid, S.; Vogelmeier, C.; Greulich, T.; Noeske, S.; Bals, R.; Koczulla, A.R.                              | Cyanose         | N = 117            | PLS-DA                   | 63              | 74              | ND                         | [29]      |
| 29. de Jongh, F.H.C.; et al.  | Cyanose         | N = 117            | PLS-DA                   | 63              | 74              | ND                         | [29]      |
| 30. Antonelli, I.; et al.   | Cyanose         | N = 117            | PLS-DA                   | 63              | 74              | ND                         | [29]      |
| 31. de Jongh, F.H.C.; et al.  | Cyanose         | N = 117            | PLS-DA                   | 63              | 74              | ND                         | [29]      |
| 32. de Jongh, F.H.C.; et al.  | Cyanose         | N = 117            | PLS-DA                   | 63              | 74              | ND                         | [29]      |
| 33. de Jongh, F.H.C.; et al.  | Cyanose         | N = 117            | PLS-DA                   | 63              | 74              | ND                         | [29]      |
| 34. de Jongh, F.H.C.; et al.  | Cyanose         | N = 117            | PLS-DA                   | 63              | 74              | ND                         | [29]      |
| 35. de Jongh, F.H.C.; et al.  | Cyanose         | N = 117            | PLS-DA                   | 63              | 74              | ND                         | [29]      |

55, doi: 10.1186/1309-8009-21-2019, 2019. reversible or not) or degree of airways obstruction, because the externally validated discriminative accuracy remained almost the same [24]; these results suggest that COPD has a specific VOC pattern production, independent from the degree of airway obstruction. Regardless of smoking, COPD can be discriminated from OSA with an accuracy of 0.75-0.80 [26][27] and a sensitivity and specificity of 0.75, while the presence of both diseases in the same patient (i.e., overlap syndrome) cannot be clearly distinguished by COPD [26]. Likewise, COPD can be discriminated from lung cancer [19][23][31]. In all these studies, the participants performed exhaled breath analysis apart from spirometry and observed some restrictions in eating, smoking, and taking medication before the test, limiting its applicability in clinical practice. A combination of a metal-oxide semiconductor e-nose with a spirometer (i.e., "SpiroNose", AMC, Amsterdam; Comon-Invent BV, Delft, The Netherlands) has represented a paramount step in the applicability of e-nose in clinical practice, allowing real-time analysis and eliminating the VOC collection and storage step. The study of De Vries and colleagues has demonstrated that SpiroNose is able to discriminate COPD Global Initiative for Obstructive Lung Disease (GOLD) stages II-IV from healthy controls, asthma, and lung cancer with a AUROC of 0.80, 0.81, and 0.88 [20], respectively, without the need for restrictions before the test.

Alpha-1 antitrypsin (AAT) deficiency is a relatively rare genetic cause for COPD. In a pilot study, an electronic nose was applied in the discrimination of 10 patients with AAT deficiency, 23 patients with COPD without AAT deficiency, and 10 healthy subjects. The authors concluded a good discriminative cross-validated accuracy based on LDA [28]. They also supplemented 11 AAT-deficient patients with human purified AAT and found a significant change in "breathprint". This change could be either due to the direct effect of AAT on the exhaled VOC pattern or may represent immunological alterations due to the augmentation therapy [28].

The "breathprint" was associated with the exercise capacity of COPD patients, expressed by the six minute walking distance and the disease-specific prognostic index BODE (Body mass index, Obstruction, Dyspnea, and Exercise), and was able to predict those patients with a steeper decline more accurately than GOLD classification with PLS-DA [32], helping clinicians tailor their interventions and follow up and also helping diagnose frail patients who could benefit from palliative care.

Although the technique is promising and is cheaper and easier to use than GC-MS, electronic noses are still more expensive than the current diagnostic spirometry and they warrant some expertise. In addition, due to the unspecific nature of the signals, they cannot easily be interpreted in clinical practice. Therefore, their role alone would be limited in diagnostic and differential diagnostic settings. However, their combination with traditional spirometry has merit in identifying endotypes and differentiating COPD from asthma with fixed airway obstruction [14][23][25]. Airway sampling using invasive techniques, such as bronchoscopy is not always feasible in COPD, and even sputum induction holds risks for patients with very severe COPD [33]. Although endotyping and monitoring airway inflammation hold essential clinical value [5], the currently used surrogates, such as blood eosinophils, only weakly correlate with their percentages in sputum [34]. In addition, it has recently been suggested that temporal variation, rather than the baseline values of blood eosinophilia, better predicted treatment response to inhaled corticosteroids in COPD [35]. The monitoring of airway inflammation via electronic nose holds clinical potential, and future studies should focus on this.