Small Cell Lung Carcinoma

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Small cell lung cancer (SCLC) is an aggressive malignancy characterized by rapid proliferation, early dissemination, acquired therapy resistance, and poor prognosis. Early diagnosis of SCLC is crucial since most patients present with advanced/metastatic disease, limiting the potential for curative treatment. While SCLC exhibits initial responsiveness to chemotherapy and radiotherapy, treatment resistance commonly emerges, leading to a five-year overall survival rate of up to 10%. New effective biomarkers, early detection, and advancements in therapeutic strategies are crucial for improving survival rates and reducing the impact of this devastating disease.

Keywords: small cell lung carcinoma (SCLC) ; biomarkers ; diagnosis ; therapeutic targets

1. Introduction

Lung cancer remains a significant global health concern, with staggering mortality rates. According to GLOBOCAN, it accounted for 2.1 million new cases and 1.8 million deaths in 2018, making it the leading cause of cancer-related deaths worldwide [1]. Lung cancer is categorized into two main histological types: non-small cell lung cancer (NSCLC) and small cell lung cancer (SCLC). NSCLC comprises approximately 85% of cases, while SCLC represents around 15% ^[2]. SCLC is an aggressive neoplasm characterized by rapid proliferation, early dissemination, metastases, acquired therapy resistance, and poor outcomes [3]. Each year, approximately 250,000 new cases of SCLC are reported, resulting in at least 200,000 deaths worldwide ^[1]. While historically more common in men, the prevalence of SCLC among women has risen due to global smoking trends. Exposure to tobacco carcinogens (polycyclic aromatic hydrocarbons and tobaccospecific nitrosamines) is considered a key risk factor for SCLC, as only 2% of all SCLC cases are among never-smokers ^[4]. Early diagnosis of SCLC is crucial as most patients present with metastatic disease, limiting the potential for curative treatment. While SCLC exhibits initial responsiveness to chemotherapy and radiotherapy, treatment resistance often emerges, leading to a five-year overall survival rate of only 10% ^[5]. Poor prognosis is associated with factors such as male gender, poor performance status, and age over 70 [5][6]. Diagnostic procedures for SCLC typically involve physical examination, performance status evaluation, laboratory tests, and imaging techniques, including contrast-enhanced CT scans of the chest and abdomen, brain MRI or CT, and optional FDG PET/CT for limited-stage disease. Pathological examination following bronchoscopy, lymph node biopsy, and metastatic lesion biopsy is essential for accurately classifying SCLC [5]. To combat the high mortality rates associated with lung cancer, smoking cessation, and prevention remain the most critical interventions in reducing lung cancer mortality [5][6].

Common clinical manifestations of SCLC at diagnosis include central tumor masses, mediastinal involvement, and extrathoracic spread in 75–80% of patients ^[6]. Symptoms may include cough, wheezing, dyspnea, hemoptysis, weight loss, pain, fatigue, and paraneoplastic syndromes. Metastasis frequently occurs in the brain, liver, adrenal glands, bone, and bone marrow, often resulting in neurological deficits and paraneoplastic syndromes ^[7].

With the addition of programmed cell death protein-1 (PD-1) and programmed death-ligand 1 (PD-L1) inhibitors to chemotherapy in the first line of extensive small cell lung cancer (SCLC), a step forward has been made in improving overall treatment outcomes for patients with SCLC ^[8]. In routine clinical practice, there are currently no available predictive biomarkers for immunotherapy response, and the use of programmed death-ligand 1 (PD-L1) and tumor mutational burden (TMB) testing is not recommended ^[5]. The need for biomarkers to predict treatment response in patients with SCLC is urgent. Potential biomarkers such as PD-L1 expression, high TMB (TMB-H), and microsatellite instability (MSI-H) need further investigation for applicability in SCLC. Effective biomarkers, early detection, and advancements in therapeutic strategies are crucial for improving survival rates and reducing the impact of this devastating disease.

2. Pathology of SCLC

SCLC belongs to the spectrum of neuroendocrine pulmonary neoplasms that share some common morphologic, ultrastructural, immunohistochemical, and molecular genomic characteristics ^{[9][10]}. Four major neuroendocrine pulmonary neoplasms are carcinoids (typical and atypical) and neuroendocrine carcinomas (SCLC and large cell neuroendocrine carcinomas; LCNEC). A typical carcinoid is a low-grade neoplasm, and atypical carcinoid is intermediate-grade, whereas both neuroendocrine carcinomas are, per definition, high-grade neoplasms. The current evidence suggests that carcinoids (typical and atypical) are closely related and etiologically different from SCLC and LCNEC ^{[9][10]}. Carcinoids are not precursor lesions of neuroendocrine carcinoids can be seen in patients with multiple endocrine neoplasis 1 (MEN1) syndrome (OMIM#131100), while somatic *MEN1* gene mutations are commonly observed in carcinoids ^{[9][10]}. Rare cases of histologic transformation of epidermal growth factor receptor (EGFR)—or anaplastic large kinase (ALK)-altered pulmonary adenocarcinomas have also been well-documented ^[11]. It is widely accepted that SCLC has the same endodermal origins as other major subtypes of lung carcinoma (e.g., adenocarcinoma or squamous cell carcinoma), arising from multipotent precursor cells ^{[9][10][12][13]}.

Morphologically, SCLC is composed of densely packed, small neoplastic cells with scanty cytoplasm and finely granular nuclear chromatin but without prominent nucleoli; nuclear molding and smudging are commonly present (**Figure 1**A,B). The cells are round or oval, although spindle cells (fusiform pattern of cancer cells) are frequently seen. Mitotic figures are numerous, while the tumor necrosis and crush artifacts may be extensive.



Figure 1. (**A**,**B**) Hematoxylin and Eosin (H&E) stain of a lung biopsy showing a small cell carcinoma with sheet-like diffuse growth pattern and basophilic appearance (A, magnification 10×); Image 1B reveals a prominent nuclear molding of neoplastic cells (magnification 20×).

SCLC expresses neuroendocrine markers, such as synaptophysin, chromogranin-A, and CD56/NCAM, which should be used as a panel ^{[14][15]}. CD56 is the most sensitive as it stains 90–100% of all SCLC, while synaptophysin and chromogranin-A can be negative in >50% of cases ^{[16][17][18]}. Neuron-specific enolase (NSE) is frequently positive in SCLC but is considered non-specific due to its widespread expression in non-neuroendocrine neoplasms (both pulmonary and extrapulmonary) ^[19]. Thyroid transcription factor 1 (TTF-1) is positive in ~80–90% of cases ^[10]. Other pulmonary biomarkers, including Napsin-A (positive in adenocarcinomas), p63, and p40 (positive in squamous cell carcinomas), are not immunoreactive in SCLC and can help in differential diagnosis, particularly on small biopsies. Other challenging cases (metastatic neuroendocrine tumors from other anatomic locations, e.g., mammary, gastrointestinal, or Merkel cell carcinoma from the skin) can be resolved using clinical history and other specific immunohistochemical biomarkers.

3. Genomic Features of SCLC

Recent research has focused on understanding the genetic basis of SCLC to identify new therapeutic targets and develop more effective treatments ^[20]. Genetic alterations contribute significantly to the development and progression of SCLC. Concomitant inactivation of two tumor suppressor genes, *TP53* and *RB1*, is found in most SCLC cases ^{[21][22]} and is found in up to 90% and 50–90% of SCLC cases, respectively. These molecular features are strikingly different from those seen in NSCLC, in which various oncogenic driver mutations/fusions prevail (e.g., *EGFR, KRAS, ALK, BRAF, RET, ROS1, MET, NTRK1-3, HER2/ERBB2*) ^{[22][23]}. Additionally, genetic alterations contributing to SCLC's development include amplifying the MYC family of oncogenes (*MYC, MYCL*, and *MYCN*), inactivation of the phosphatase and tensin homolog (*PTEN*) tumor suppressor gene, and mutations in the Notch signaling pathway. Genomic alterations of *MYC* family members are seen in SCLC and represent biomarkers of poor prognosis. In particular, *MYCN* alterations are related to SCLC cases with immunotherapy failure. The most important genes altered in SCLC in humans are summarized in **Table 1**.

Different studies have identified recurrent mutations in chromatin remodeling genes, such as *ARID1A*, *ARID1B*, and *SMARCA4*, which regulate gene expression. These mutations may contribute to the dysregulation of critical genes involved in cell proliferation and survival, leading to the development of SCLC, characterized by a high frequency of mutations in genes that regulate cell cycle and DNA damage response pathways, such as *TP53*, *RB1*, and *PTEN*. Additionally, SCLC often exhibits widespread chromosomal instability, with frequent amplifications and deletions of large genome regions. In addition to these genetic alterations, SCLC is characterized by a high frequency of copy number alterations, including amplification of MYC family members and deletion of the tumor suppressor gene cyclin-dependent kinase inhibitor 2A (*CDKN2A*) ^[24]. In addition, the changes in the stroma and immune microenvironment are additional factors involved in the pathogenesis of SCLC ^[25].

Overall, the genetic landscape of SCLC is complex and heterogeneous, with multiple genetic alterations contributing to its aggressive phenotype. Understanding the underlying genetic mechanisms of SCLC is crucial for developing effective targeted therapies and personalized treatment strategies for patients with this aggressive cancer. SCLC mutational characteristics reveal a clear causal connection with smoking. Direct scientific evidence confirms that carcinogens from tobacco are responsible for initiating SCLC ^[26].

Genomic profiling in patients with SCLC has not revealed mutationally defined subtypes of SCLC. However, due to the lack of larger studies, this may be a consequence of the insufficient number of tumor samples included in analyses. Therefore, there is a substantial need for clinical trials that include the analyses of tumor tissue to identify vital genomic triggers. However, there is an accentuated difficulty in tumor material collection. Ethnicity or smoking status did not affect the consistency of mutational differences; however, the prevalence of oncogenic triggers is considered higher in never-smokers with SCLC compared to tobacco users ^[27].

In addition, genetically modified mice have provided critical genetic lessons and contributed to the knowledge of molecular mechanisms of SCLC etiopathogenesis, metastasis, and response to treatment. It has been shown that tumors in mice show genetic alterations and histological features like those in humans. Ferone et al. provided a comprehensive review of lung cancers and lessons from mouse studies, showing an enormous contribution of animal studies in pulmooncology ^[28].

Table 1. Most important genes altered in SCLC (mostly according to memorial Sloan Kettering-integrated mutation profiling of actionable cancer targets—MSK-IMPACT sequencing of SCLC tumors)—data adopted from Cheng et al. ^[29], Rudin et al. ^[23], and Liu et al. ^[30].

Gene	Aliases	Gene Location on Human Chromosome and Number of Amino Acids	Gene Alteration in SCLC	Known Function and Features	Frequency of Mutation in SCLC (% in Various Cohorts)	Refs.
TP53	Tumor protein 53; p53; Phosphoprotein P53; Antigen NY-CO-13; Transformation- Related Protein 53; BCC7, LFS1, TRP53, tumor protein BMFS5	Chromosome 17 at position 17p13.1.; 375 amino acids	Inactivating mutation; deletion	Nuclear phosphoprotein involved in the regulation of cell proliferation; tumor suppressor; transcription regulation	77–89	Chang et al. [<u>31]</u> Rudin et al. [<u>23]</u>
RB1	RB1, pRb, RB, retinoblastoma 1, OSRC, PPP1R130, p105-Rb, pp110, Retinoblastoma protein, RB transcriptional corepressor 1, p110- RB1	Chromosome 13 at position 13q14.1- q14.2.; 928 amino acids	Inactivating mutation; deletion; loss or inactivation of both copies of the gene	Tumor suppressor protein that is dysfunctional in several major cancers. Prevents excessive cell growth by inhibiting cell cycle progression -key regulator of the G1/S transition of the cell cycle	50–90	George et al. [21] Febres- Aldana et al. [32]

Gene	Aliases	Gene Location on Human Chromosome and Number of Amino Acids	Gene Alteration in SCLC	Known Function and Features	Frequency of Mutation in SCLC (% in Various Cohorts)	Refs.
KMT2D	KMT2D, ALR, KABUK1, MLL2, MLL4, lysine methyltransferase 2D, histone-lysine methyltransferase 2D, TNRC21, AAD10, KMS, CAGL114	Chromosome 12 at position 12q13.12.; 5316 amino acids	Inactivating mutation; deletion; gene fusion; truncating nonsense/frameshift/splice site mutations	Key regulator of transcriptional enhancer function; major enhancer regulator in mammalian cells, including regulation of development, differentiation, metabolism, and tumor suppression.	5–13	Wu et al. ^[33] Simbolo et al. ^[34] Augert et al. ^[35]
CREBBP	AW558298, CBP, CBP/p300, KAT3A, p300/CBP, RSTS, CREB binding protein, RSTS1, MKHK1	Chromosome 16 at position 16p13.3. 2414 amino acids.	Inactivating mutation, deletion	Crucial role in transcriptional regulation and chromatin remodeling. Interacts with various transcription factors and coactivators, influencing the expression of target genes involved in cell growth, differentiation, and development.	4–10	Carazo et al. ^[36] Jia et al. ^[37]
PTEN	PTEN, 10q23del, BZS, CWS1, DEC, GLM2, MHAM, MMAC1, PTEN1, TEP1, phosphatase and tensin homolog, Phosphatase and tensin homolog, PTENbeta	Chromosome 10 at position 10q23.3. 403 amino acids	inactivating mutations, deletions, or loss of expression	Tumor suppressor involved in the regulation of the <i>PI3K/AKT/mTOR</i> pathway, which plays a critical role in cell survival and proliferation. PTEN's protein phosphatase activity may be involved in the regulation of the Cell cycle, preventing cells from growing and dividing too rapidly.	3-10	Sivakumar et al. ^[38] Zhang et al. ^[39]
FAT1	CDHF7, CDHR8, FAT, ME5, hFat1, FAT atypical cadherin 1	Chromosome 4 at position 4q35.2. 4410 amino acids	Inactivation mutation; deletion	Cell-cell adhesion, migration and communication, regulation of tissue growth, cell polarity, and migration; tumor suppressor gene	2–10	JiaXin et al. ^[40] Pop-Bica et al. ^[41]
PIK3CA	PIK3CA, CLOVE, CWS5, MCAP, MCM, MCMTC, PI3K, p110- alpha, PI3K-alpha, phosphatidylinositol- 4,5-bisphosphate 3- kinase catalytic subunit alpha, CLAPO, CCM4	Chromosome 3 at position 3q26.3.; 1068 amino acids	Activating mutation; mutations in specific regions	The <i>PIK3CA</i> gene for synthesis of the catalytic subunit alpha of the enzyme phosphatidylinositol 3-kinase, having crucial role in cell growth, proliferation, and survival	1-7	Hung et al. ^[42] Pop-Bica et al. ^[41]
NOTCH1	NOTCH1, Notch1, 9930111A19Rik, Mis6, N1, Tan1, lin- 12, AOS5, AOVD1, hN1	Chromosome 9 at position 9q34.3. 2527 amino acids	Inactivating mutation	Tumor suppressor; involved in cell signalling processes	1–6	Li et al. ^[43] Roper et al. ^[44] Herbreteau et al. ^[45]

Gene	Aliases	Gene Location on Human Chromosome and Number of Amino Acids	Gene Alteration in SCLC	Known Function and Features	Frequency of Mutation in SCLC (% in Various Cohorts)	Refs.
NF1	NFNS, VRNF, WSS, neurofibromin 1	Chromosome 17 at position 17q11.2. 2818 amino acids	Inactivating mutation, deletion	Tumor suppressor. Neurofibromin 1 plays a role in regulating cell growth and proliferation by negatively regulating the activity of Ras, associated with uncontrolled cell growth.	3–4	Ross et al. ^[45] Shimizu et al. ^[47]
APC	BTPS2, DP2, DP2.5, DP3, GS, PPP1R46, adenomatous polyposis coli, WNT signaling pathway regulator	Chromosome 5 at position 5q22.2. 2843 amino acids	Inactivating mutation, deletion	Crucial role in regulating the Wnt signaling pathway and controlling cell proliferation, growth, differentiation, and migration.	3-4	Jin et al. ^[48] Grote et al. ^[49]
EGFR	ERBB, ERBB1, HER1, NISBD2, PIG61, mENA, epidermal growth factor receptor, erbB-1, ERRP	Chromosome 7 at position 7p12.1. 1210 amino acids	Activating mutation	Oncogene; a receptor tyrosine kinase that plays a critical role in cell growth, proliferation, and survival; involved in <i>RAS</i> signaling pathway.	3-4	Ding et al. ^[50] Hao et al. ^[51]
KRAS	C-K-RAS, CFC2, K- RAS2A, K-RAS2B, K- RAS4A, K-RAS4B, KI-RAS, KRAS1, KRAS2, NS, NS3, RALD, RASK2, K- ras, KRAS proto- oncogene, GTPase, c-Ki-ras2, OES, c-Ki- ras, K-Ras 2, K-Ras, Kirsten Rat Sarcoma virus	Chromosome 12 at position 12p12.1. 189 amino acids	Activating mutation	A GTPase involved in cell signalingpathways that regulate cell growth and proliferation (<i>RASIMAPK</i>). <i>KRAS</i> mutations can lead to the constitutive activation of the KRAS protein, resulting in dysregulated cell signaling and increased cell proliferation.	1-3	Otegui et al. (52) Li et al. ⁽⁵³⁾
<i>NOTCH</i> 3	CADASIL, CASIL, IMF2, LMNS, CADASIL1, notch 3, notch receptor 3	Chromosome 19 at position 19p13.2. 2345 amino acids	Inactivating mutation, deletion	Involved in cell signaling pathways. Notch signaling plays a critical role in cellular processes, such as cell fate determination, differentiation, and development.	<3	Herbreteau et al. ^[45] Du et al. ^[54]

4. Biomarkers in SCLC

In contrast to NSCLC, the discovery of therapeutic targets in SCLC has not been easy, partly because driver mutations are in first-line loss of function or untargetable, e.g., MYC family members ^[23]. The recent division of SCLC into molecular subtypes based on the expression of transcription factors has provided an essential step in searching for new therapeutic targets for the disease. This classification system identifies four distinct subtypes of SCLC: achaete-scute homolog 1 (ASCL1), neurogenic differentiation factor 1 (NEUROD1), yes-associated protein 1 (YAP1), and POU class 2 homeobox 3 (POU2F3) ^[57].

Ngwh blood-barsed biomarkers for the early detection of the several the several detection of the showing promising results. "Liquito fations biomarkers such as tumor-dende for the stress of the str Human cells (CTC), and circulating tumo_{En} to the to be promising tools in cancer monitoring creation with the second se cells express different tumor-speatific NHRARErs, including Delta-like protein 3 (DLL-3), which maying associated with a worse prognosis in patients with S_{Crids}^{Crids} . However, whether these biomarkers listed in **Table 2** with S_{Crids}^{Crids} . in the population, especially in cancer with aggressive biologic behavior such as SCLC, remains unknown. To date, it ARIDIA B120, BAF250, Chromosome Inactivating mutation, Tumor suppressor <3 Du et al. [54] seems that Bategina, with SCLC have the the the the transfer of CTC, which was the parent of t clinically evaluating the boots and provide the boots and provided t OSA1, P270, 2254 amino regulating regulating alterations, bave per philosophilic alterations, bave per philosophilic and the philosophilic alteration of the p and the occulrent of State of State relapse [60]. remodeling and rich interaction aene expression: involved in various domain 1A Furthermore, the characterization of extracellular vesicles, such as exocements, paresets, to be a promising tool and alternative source for various analytes in liquid biopsies ^[61]. This approach the product of the product of the significantly contribute to the identification of new biomarkers for the diagnosis and monitoring of Seyman at the development of promising prognostic models. Emerging predictive and prognostic biomark differentiational and indispensable for selecting thermast suitable therapeutic option for batimets with the mutation, Protein tyrosine <3 Sato et al. HPTPDELTA, PTPD, 9 at position deletion phosphatase To date, generations of Moral were noticed in about 20% of patients with SLEC, representing the third most a role in regulating common genetised areasenality following TP53 and RB1 and a potential biomacklesign at a generation of the second MSI-H have receptor type as potential predictive biomarkers for response to minute checkpoint inhibitors (ICIs) in protein typesine patients with the set of the set agents. The recentor type defitific evidence confirms its importance as a differentiation pandictive biomarker for several migration. therapeutics, including platinum and PARP inhibitors [63]. Expression of SLFN11 in CTCs provides a potential biomarker of sensitivity for TSRAMS, MBXHET chemotherapy drugs and bdy (ADP hose) pulline suppresses infibition in SCL et patients RAD54, RAD54L, chromosome deletion [62]. Therefore the set of the se tissue sampling, ZNF-HX, MRX52, alpha Xa21.1. remodeling and the regulation of gene thalassemia/mental expression. ATRX is retardation Table 2. Potential biomarkers in small cell luingotarcinoma. syndrome X-linked, maintaining the chromatin stability and Biomarker remodeler, ATRX Potential Application References Type chromatin remodeler telomeres and in Biomarker for SCECH Biographisas Delta-like ligand 3 Tumor-specific marker Chen et al. [58] DLL3 Circulating tumor cells (CTC) Liquid biopsy biomarker Prognostic biomarker for therapy evaluation Roumeliotou et al. [59] of therapy efficacy **Circulating tumor DNA** Liquid biopsy biomarker Biomarker for treatment efficacy and relapse Almodovar et al. [60] (ctDNA) detection Zhang et al. [61] Exosomes Extracellular vesicles Non-invasive biomarkers for prognosis MYC proto-oncogene/bHLH Genetic alteration Potential biomarker for targeted therapy Taniguchi et al. [62] transcription factor (MYC) Programmed death-ligand 1 Immune checkpoint Potential biomarker for immunotherapy Taniguchi et al. [62] (PD-L1) protein response Tumor mutational burden Mutation load of a tumor Potential biomarker for immunotherapy Taniguchi et al. [62] and (TMB) response Li et al. [65] Microsatellite instability Genetic marker of Potential biomarker for immunotherapy Taniguchi et al. [62] and (MSI-H) Microsatellite Instability response Chang et al. [66] Schlafen 11 Liquid biopsy biomarker Potential biomarker for the response on DNA Taniquchi et al. (SLFN11) damaging chemotherapy and PARP [62] and Zhang et al. [63] inhibition

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