Mutations of SARS-CoV-2 RBD

Subjects: Virology Contributor: Ahmed Alaofi

The genetic mutation of SARS-CoV-2 -especially RBD- might be linked to the viral properties that influence the viral transmission mode and severity of COVID-19 as well as RBD conformation. One of the dominant variants during COVID-19 pandemic has been the D614G mutation (not in the RBD region) of S glycoprotein; several reports have claimed that this mutation is able to increase the infectivity and stability of SARS-CoV-2. Up until now, most neutralizing antibodies against SARS-CoV-2 have been targeting its RBD. However, there have been several mutations reported in SARS-CoV-2 RBD, such as N501Y, L452R, S477N, E484K, A502S, N439K, S494P, T478K, K417N, and K417T. These mutations pose a threat due to their role in host cell entry via the hACE2 receptor, which might strengthen SARS-CoV-2 infectivity, conformation and stability of RBD, viral load, or resistance against neutralizing antibodies.

Keywords: wild-type RBD ; mutant RBDs ; SARS-CoV-2 ; molecular dynamics simulations ; RBD flexibility ; principle component analysis ; free energy landscape

1. Overview

The receptor-binding domain (RBD) of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) mediates the viral-host interaction and is a target for most neutralizing antibodies. Nevertheless, SARS-CoV-2 RBD mutations pose a threat due to their role in host cell entry via the human angiotensin-converting enzyme 2 receptor that might strengthen SARS-CoV-2 infectivity, viral load, or resistance against neutralizing antibodies. To understand the molecular structural link between RBD mutations and infectivity, the top five mutant RBDs (i.e., N501Y, E484K L452R, S477N, and N439K) were selected based on their recorded case numbers. These mutants along with wild-type (WT) RBD were studied through all-atom molecular dynamics (MD) simulations of 100 ns. The principal component analysis and the free energy landscape were used too. Interestingly, N501Y, N439K, and E484K mutations were observed to increase the rigidity in some RBD regions while increasing the flexibility of the receptor-binding motif (RBM) region, suggesting a compensation of the entropy penalty. However, S477N and L452R RBDs were observed to increase the flexibility of the RBM region while maintaining similar flexibility in other RBD regions in comparison to WT RBD. Therefore, both mutations (especially S477N) might destabilize the RBD structure, as loose conformation compactness was observed. The destabilizing effect of S477N RBD was consistent with previous work on S477N mutation. Finally, the free energy landscape results showed that mutations changed WT RBD conformation while local minima were maintained for all mutant RBDs. In conclusion, RBD mutations definitely impact the WT RBD structure and conformation as well as increase the binding affinity to angiotensin-converting enzyme receptor.

2. Genetic Mutation of SARS-CoV-2

The pandemic of coronavirus disease 2019 (COVID-19) has infected over 173 million individuals (at the time of this writing) and caused millions of deaths around the globe ^[1]. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), responsible for COIVD-19, is a single-stranded positive-strand RNA virus that belongs to the Coronaviridae family ^[2]. Coronaviruses (CoVs) were previously known to be present in the environment and to infect humans, although the earlier infections resulted in mild symptoms and were limited to local areas. However, deadly human CoVs, such as SARS-CoV and Middle East respiratory syndrome coronavirus (MERS-CoV) as well as SARS-CoV-2, have appeared in the past two decades. These CoVs are more severe and cover more ground in every passing phase, as they can cause deadly pneumonia in humans along with other gastrointestinal diseases ^{[3][4][5]}. SARS-CoV-2 is characterized by efficient transmission and its ability to rapidly spread worldwide despite its lower mortality rate (3.3%) in comparison to SARS-CoV (10%) and MERS-CoV (37%) ^[6].

Structural components of SARS-CoV-2 have been extensively studied ^{[7][8][9][10][11]}. On mature virus, the spike (S) glycoprotein on the surface of SARS-CoV-2 is composed of an extracellular domain (EC), transmembrane (anchor) domain, and short intracellular tail domain (IC) ^{[11][12]}. EC domain has two functional subunits: a receptor-binding subunit

(S1) and a membrane-fusion (S2) subunit ^[13]. The host cell (cellular) proteases cleave S protein at the boundary between S1–S2 site and S2' site during host–virus membrane fusion ^{[11][14]}. Further, the S1 subunit compromises the receptorbinding domain (RBD) that is essential for receptor binding and contributes to stabilizing the S2 subunit that harbors the fusion machinery. After the S1 subunit binds to the cellular receptor, subsequent structural rearrangements of metastable S glycoprotein occur to allow fusion between the viral and the host cell membranes. The structural rearrangements can be explained by the conformational dynamics behavior of the S glycoprotein trimer that eventually results in an open (standing) conformation in order to successfully achieve binding and fusion events ^[Z]. In fact, S glycoprotein is a target for immune cells that neutralize the virus, as many vaccines have been developed based on the antigenicity of S glycoprotein. SARS-CoV-2 RBD is known to bind to human angiotensin converting enzyme 2 (AEC2), specifically through the receptor-binding motif (RMB) of RBD, to mediate the viral–host interaction. Moreover, the RBM bears a flexible nature and contains most of the SARS-CoV-2 RBD residues that bind directly to ACE2 receptor ^[10]. However, RBD must adopt a specific conformation (up conformation) to bind efficiently to ACE2 ^[15].

Importantly, the genetic mutation of SARS-CoV-2 might be linked to the viral properties that influence the viral transmission mode and severity of COVID-19 as well as RBD conformation ^{[15][16]}. One of the dominant variants during COVID-19 pandemic has been the D614G mutation (not in the RBD region) of S glycoprotein; several reports have claimed that this mutation is able to increase the infectivity and stability of SARS-CoV-2 ^{[12][18][19][20][21]}. Up until now, most neutralizing antibodies against SARS-CoV-2 have been targeting its RBD ^{[22][23][24][25][26]}. However, there have been several mutations reported in SARS-CoV-2 RBD, such as N501Y, L452R, S477N, E484K, A502S, N439K, S494P, T478K, K417N, and K417T. These mutations pose a threat due to their role in host cell entry via the hACE2 receptor, which might strengthen SARS-CoV-2 infectivity, conformation and stability of RBD, viral load, or resistance against neutralizing antibodies including one authorized by the U.S. Food and Drug Administration (FDA) for emergency use ^[32]. It is clear that these mutation sites are mostly located in the RBM in the RBD region of SARS-CoV-2, which has shown a flexible nature. Importantly, the molecular dynamics and flexibility of the RBD region might have contributed to



SARS-CoV-2 infectivity [31][32].

3. Conclusions

Mutations in proteins can affect protein conformation, folding, and stability, and can eventually influence protein–protein interactions and protein thermodynamics ^[33]. There are several observed mutations in the RBD of SARS-CoV-2 S glycoprotein that improve its infectivity and strengthen the viral binding interaction to ACE2 receptor ^[27]. These mutations compromise the neutralizing ability of anti-SARS-CoV-2 antibodies; therefore, it is necessary to study the mutation effects on the RBD structure (e.g., conformation, stability, dynamics, etc.) ^{[24][26]}. Several structural and dynamic studies at the molecular level show that SARS-CoV-2 RBDs have to adapt open conformation (also known as "up" or "standing") to effectively bind to ACE2 receptors ^{[8][16][34][35]}. So far, mutations in non-RBD residues, such as the D614G variant, can populate RBD open conformation rather than closed conformations ^[36]. Most of the virulent point mutations occur in the RBM loop that directly binds hACE2 and are more prone to conformational variations; thus, these mutations may have the ability to generate a more stable complex with high binding affinity ^[37]. For instance, N439K, L452R, T478I, and E484D mutations on RBM have significant free energy changes, and they constitute approximately 58% of all mutations on RBD.

frequency of S477N, N439K, V483A, and V367F), clearly indicating the natural selection of mutations with stronger transmissibility ^[27].

Here, we studied five critical mutant RBDs according to the RBD mutation tracking website (CovMT) ^[38] by utilizing the allatom MD simulation technique. The N501Y flexibility in loop Y473–C489 of RBM was comparable to WT RBD; thus, suggesting that tyrosine mutation did not alter neither the loop Y473–C489 flexibility or the whole RBD conformational compactness. Only N501Y RBD showed different loop Y495–Y508 conformations. The mutation of alanine instead of tyrosine at 501 (i.e., N501A) shows an increase in loop Y473–C489 flexibility and conformational compactness according to a related study ^[34]. Moreover, the same loop Y473–C489 in SARS-CoV RBD showed a higher flexibility in comparison to SARS-CoV-2 RBD ^{[34][35]}. This suggests that the higher infectivity of the N501Y variant might be attributed to an improvement in the N501Y RBD conformation and therefore a higher affinity to ACE2 receptor. Previously, substitution mutations in the SARS-CoV-2 RBD, N501, L452, N439, E484, T470, and Q498 have been shown to enhance binding affinity for hACE2, thereby increasing infectivity and transmissibility in comparison to the natural SARS-CoV-2 ^[39].

On the other hand, the loop Y473–C489 flexibility was increased in L452R, N439K, and E484K RBDs, and these mutations were associated with higher infectivity and binding affinity to ACE2. This was consistent with previously reported structural analyses that showed that the RBM region has the highest flexibility ^{[9][32][40]}. However, the observed rigidity in some parts of RBDs (i.e., non-RBM regions) in N439K and E484K RBDs might compensate for the entropy penalty due to flexibility in the loop Y473–C489. Therefore, the N501Y, N439K, and E484K mutations studied in this work have insignificant changes in the overall RBD flexibility. This was indicated by the SARS-CoV-2 mutations may augment the conformational sampling to avoid the entropy cost upon interaction with ACE2 receptor. In contrast, S477N and L452R RBDs showed comparable flexibility to WT RBD in non-RBM regions but higher flexibility in the RBM regions, as well as a loose conformational compactness. Our results were consistent with a pervious study that showed that S477N has a destabilizing effect on RBD structure and therefore less prone to develop disease

PCA, FEL, and porcupine plot results suggested that the destabilizing effect could be noticed in the loop Y473–C489 of S477N RBD as compared to WT RBD. However, the conformational sampling of energetically favorable conformations of mutant RBDs showed local minima, thereby indicating stable structures for these mutant RBDs. The COVDI-19 pandemic presents a continuing threat to global health due to its ongoing critical mutations. To conquer this pandemic, it is necessary to investigate the effects of SARS-CoV-2 mutations at all possible levels, such as structural, functional, and activity levels. Moreover, RBD mutations directly affect the ability of SARS-CoV-2 to binding to ACE2 receptor and therefore affect its infectivity. Our structural investigation of critical mutant RBDs along with WT RBD showed that RBD mutations have a direct impact on the molecular structural of SARS-CoV-2. These data might be helpful for researchers investigating antiviral agents and vaccine research and development against SARS-CoV-2, especially mutant SARS-CoV-2 viruses.

References

- 1. WHO Coronavirus (COVID-19) Dashboard. Available online: https://covid19.who.int (accessed on 6 June 2021).
- Shang, J.; Wan, Y.; Luo, C.; Ye, G.; Geng, Q.; Auerbach, A.; Li, F. Cell Entry Mechanisms of SARS-CoV-2. Proc. Natl. Acad. Sci. USA 2020, 117, 11727–11734.
- 3. Gao, H.; Yao, H.; Yang, S.; Li, L. From SARS to MERS: Evidence and Speculation. Front. Med. 2016, 10, 377–382.
- 4. Walls, A.C.; Park, Y.-J.; Tortorici, M.A.; Wall, A.; McGuire, A.T.; Veesler, D. Structure, Function, and Antigenicity of the SARS-CoV-2 Spike Glycoprotein. Cell 2020, 181, 281–292.
- 5. Ye, Z.-W.; Yuan, S.; Yuen, K.-S.; Fung, S.-Y.; Chan, C.-P.; Jin, D.-Y. Zoonotic Origins of Human Coronaviruses. Int. J. Biol. Sci. 2020, 16, 1686–1697.
- 6. Hu, T.; Liu, Y.; Zhao, M.; Zhuang, Q.; Xu, L.; He, Q. A Comparison of COVID-19, SARS and MERS. PeerJ 2020, 8.
- 7. Wrapp, D.; Wang, N.; Corbett, K.S.; Goldsmith, J.A.; Hsieh, C.-L.; Abiona, O.; Graham, B.S.; McLellan, J.S. Cryo-EM Structure of the 2019-NCoV Spike in the Prefusion Conformation. Science 2020, 367, 1260–1263.
- Yan, R.; Zhang, Y.; Li, Y.; Ye, F.; Guo, Y.; Xia, L.; Zhong, X.; Chi, X.; Zhou, Q. Structural Basis for the Different States of the Spike Protein of SARS-CoV-2 in Complex with ACE2. Cell Res. 2021, 31, 717–719.
- 9. Shang, J.; Ye, G.; Shi, K.; Wan, Y.; Luo, C.; Aihara, H.; Geng, Q.; Auerbach, A.; Li, F. Structural Basis of Receptor Recognition by SARS-CoV-2. Nature 2020, 581, 221–224.

- 10. Lan, J.; Ge, J.; Yu, J.; Shan, S.; Zhou, H.; Fan, S.; Zhang, Q.; Shi, X.; Wang, Q.; Zhang, L.; et al. Structure of the SARS-CoV-2 Spike Receptor-Binding Domain Bound to the ACE2 Receptor. Nature 2020, 581, 215–220.
- Yang, J.; Petitjean, S.J.L.; Koehler, M.; Zhang, Q.; Dumitru, A.C.; Chen, W.; Derclaye, S.; Vincent, S.P.; Soumillion, P.; Alsteens, D. Molecular Interaction and Inhibition of SARS-CoV-2 Binding to the ACE2 Receptor. Nat. Commun. 2020, 11, 4541.
- 12. Li, F. Receptor Recognition Mechanisms of Coronaviruses: A Decade of Structural Studies. J. Virol. 2015, 89, 1954– 1964.
- 13. Wang, Q.; Zhang, Y.; Wu, L.; Niu, S.; Song, C.; Zhang, Z.; Lu, G.; Qiao, C.; Hu, Y.; Yuen, K.-Y.; et al. Structural and Functional Basis of SARS-CoV-2 Entry by Using Human ACE2. Cell 2020, 181, 894–904.
- 14. Li, W. Delving Deep into the Structural Aspects of a Furin Cleavage Site Inserted into the Spike Protein of SARS-CoV-2: A Structural Biophysical Perspective. Biophys. Chem. 2020, 264, 106420.
- 15. Henderson, R.; Edwards, R.J.; Mansouri, K.; Janowska, K.; Stalls, V.; Gobeil, S.M.C.; Kopp, M.; Li, D.; Parks, R.; Hsu, A.L.; et al. Controlling the SARS-CoV-2 Spike Glycoprotein Conformation. Nat. Struct. Mol. Biol. 2020, 27, 925–933.
- 16. Mansbach, R.A.; Chakraborty, S.; Nguyen, K.; Montefiori, D.; Korber, B.; Gnanakaran, S. The SARS-CoV-2 Spike Variant D614G Favors an Open Conformational State. bioRxiv 2020.
- 17. Plante, J.A.; Liu, Y.; Liu, J.; Xia, H.; Johnson, B.A.; Lokugamage, K.G.; Zhang, X.; Muruato, A.E.; Zou, J.; Fontes-Garfias, C.R.; et al. Spike Mutation D614G Alters SARS-CoV-2 Fitness. Nature 2021, 592, 116–121.
- Korber, B.; Fischer, W.M.; Gnanakaran, S.; Yoon, H.; Theiler, J.; Abfalterer, W.; Hengartner, N.; Giorgi, E.E.; Bhattacharya, T.; Foley, B.; et al. Tracking Changes in SARS-CoV-2 Spike: Evidence That D614G Increases Infectivity of the COVID-19 Virus. Cell 2020, 182, 812–827.
- 19. Fernández, A. Structural Impact of Mutation D614G in SARS-CoV-2 Spike Protein: Enhanced Infectivity and Therapeutic Opportunity. ACS Med. Chem. Lett. 2020, 11, 1667–1670.
- 20. Zhang, L.; Jackson, C.B.; Mou, H.; Ojha, A.; Rangarajan, E.S.; Izard, T.; Farzan, M.; Choe, H. The D614G Mutation in the SARS-CoV-2 Spike Protein Reduces S1 Shedding and Increases Infectivity. BioRxiv Prepr. Serv. Biol. 2020.
- 21. Becerra-Flores, M.; Cardozo, T. SARS-CoV-2 Viral Spike G614 Mutation Exhibits Higher Case Fatality Rate. Int. J. Clin. Pract. 2020, 74, e13525.
- Starr, T.N.; Greaney, A.J.; Hilton, S.K.; Ellis, D.; Crawford, K.H.D.; Dingens, A.S.; Navarro, M.J.; Bowen, J.E.; Tortorici, M.A.; Walls, A.C.; et al. Deep Mutational Scanning of SARS-CoV-2 Receptor Binding Domain Reveals Constraints on Folding and ACE2 Binding. Cell 2020, 182, 1295–1310.
- 23. Cao, Y.; Su, B.; Guo, X.; Sun, W.; Deng, Y.; Bao, L.; Zhu, Q.; Zhang, X.; Zheng, Y.; Geng, C.; et al. Potent Neutralizing Antibodies against SARS-CoV-2 Identified by High-Throughput Single-Cell Sequencing of Convalescent Patients' B Cells. Cell 2020, 182, 73–84.
- 24. Shi, R.; Shan, C.; Duan, X.; Chen, Z.; Liu, P.; Song, J.; Song, T.; Bi, X.; Han, C.; Wu, L.; et al. A Human Neutralizing Antibody Targets the Receptor-Binding Site of SARS-CoV-2. Nature 2020, 584, 120–124.
- 25. Potently Neutralizing and Protective Human Antibodies against SARS-CoV-2 PubMed. Available online: https://pubmed.ncbi.nlm.nih.gov/32668443/ (accessed on 2 June 2021).
- Pinto, D.; Park, Y.-J.; Beltramello, M.; Walls, A.C.; Tortorici, M.A.; Bianchi, S.; Jaconi, S.; Culap, K.; Zatta, F.; De Marco, A.; et al. Cross-Neutralization of SARS-CoV-2 by a Human Monoclonal SARS-CoV Antibody. Nature 2020, 583, 290–295.
- 27. Chen, J.; Wang, R.; Wang, M.; Wei, G.-W. Mutations Strengthened SARS-CoV-2 Infectivity. J. Mol. Biol. 2020, 432, 5212–5226.
- Ozono, S.; Zhang, Y.; Ode, H.; Sano, K.; Tan, T.S.; Imai, K.; Miyoshi, K.; Kishigami, S.; Ueno, T.; Iwatani, Y.; et al. SARS-CoV-2 D614G Spike Mutation Increases Entry Efficiency with Enhanced ACE2-Binding Affinity. Nat. Commun. 2021, 12, 848.
- 29. Emergence of RBD Mutations in Circulating SARS-CoV-2 Strains Enhancing the Structural Stability and Human ACE2 Receptor Affinity of the Spike Protein | BioRxiv. Available online: https://www.biorxiv.org/content/10.1101/2020.03.15.991844v4 (accessed on 2 June 2021).
- 30. Wang, Y.; Liu, M.; Gao, J. Enhanced Receptor Binding of SARS-CoV-2 through Networks of Hydrogen-Bonding and Hydrophobic Interactions. Proc. Natl. Acad. Sci. USA 2020, 117, 13967–13974.
- 31. Spinello, A.; Saltalamacchia, A.; Magistrato, A. Is the Rigidity of SARS-CoV-2 Spike Receptor-Binding Motif the Hallmark for Its Enhanced Infectivity? Insights from All-Atom Simulations. J. Phys. Chem. Lett. 2020, 11, 4785–4790.

- Thomson, E.C.; Rosen, L.E.; Shepherd, J.G.; Spreafico, R.; da Silva Filipe, A.; Wojcechowskyj, J.A.; Davis, C.; Piccoli, L.; Pascall, D.J.; Dillen, J.; et al. Circulating SARS-CoV-2 Spike N439K Variants Maintain Fitness While Evading Antibody-Mediated Immunity. Cell 2021, 184, 1171–1187.
- 33. Zhou, G.; Chen, M.; Ju, C.J.T.; Wang, Z.; Jiang, J.-Y.; Wang, W. Mutation Effect Estimation on Protein-Protein Interactions Using Deep Contextualized Representation Learning. NAR Genom. Bioinforma. 2020, 2, Iqaa015.
- Dehury, B.; Raina, V.; Misra, N.; Suar, M. Effect of Mutation on Structure, Function and Dynamics of Receptor Binding Domain of Human SARS-CoV-2 with Host Cell Receptor ACE2: A Molecular Dynamics Simulations Study. J. Biomol. Struct. Dyn. 2020, 1–15.
- 35. Williams, J.K.; Wang, B.; Sam, A.; Hoop, C.L.; Case, D.A.; Baum, J. Molecular Dynamics Analysis of a Flexible Loop at the Binding Interface of the SARS-CoV-2 Spike Protein Receptor-Binding Domain. bioRxiv 2021.
- 36. Gobeil, S.M.-C.; Janowska, K.; McDowell, S.; Mansouri, K.; Parks, R.; Manne, K.; Stalls, V.; Kopp, M.F.; Henderson, R.; Edwards, R.J.; et al. D614G Mutation Alters SARS-CoV-2 Spike Conformation and Enhances Protease Cleavage at the S1/S2 Junction. Cell Rep. 2021, 34, 108630.
- Gan, H.H.; Twaddle, A.; Marchand, B.; Gunsalus, K.C. Structural Modeling of the SARS-CoV-2 Spike/Human ACE2 Complex Interface Can Identify High-Affinity Variants Associated with Increased Transmissibility. J. Mol. Biol. 2021, 433, 167051.
- 38. Alam, I.; Radovanovic, A.; Incitti, R.; Kamau, A.A.; Alarawi, M.; Azhar, E.I.; Gojobori, T. CovMT: An Interactive SARS-CoV-2 Mutation Tracker, with a Focus on Critical Variants. Lancet Infect. Dis. 2021, 21, 602.
- Yi, C.; Sun, X.; Ye, J.; Ding, L.; Liu, M.; Yang, Z.; Lu, X.; Zhang, Y.; Ma, L.; Gu, W.; et al. Key Residues of the Receptor Binding Motif in the Spike Protein of SARS-CoV-2 That Interact with ACE2 and Neutralizing Antibodies. Cell. Mol. Immunol. 2020, 17, 621–630.
- 40. De Oliveira, C.C.S.; Pereira, G.R.C.; De Alcantara, J.Y.S.; Antunes, D.; Caffarena, E.R.; De Mesquita, J.F. In Silico Analysis of the V66M Variant of Human BDNF in Psychiatric Disorders: An Approach to Precision Medicine. PLoS ONE 2019, 14, e0215508.

Retrieved from https://encyclopedia.pub/entry/history/show/32289