VLP-Based Vaccines for Trypanosomatid Diseases

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Virus-like particle (VLP) technology is involved in many vaccines, including those undergoing studies related to COVID-19. The potential use of VLPs as vaccine adjuvants opens an opportunity for the use of protozoan antigens for the development of vaccines against diseases caused by *Trypanosoma cruzi*, *Leishmania* spp., and *Trypanosoma brucei*.

Keywords: virus-like particles ; vaccine ; trypanosomatids ; Chagas disease ; leishmaniasis ; African trypanosomiasis

1. Introduction

Trypanosoma cruzi, *Trypanosoma brucei*, and *Leishmania* spp. are flagellate parasites that belong to the Trypanosomatidae family and have a complex life cycle between invertebrate and mammalian hosts, where they cause Chagas disease, African trypanosomiasis, and leishmaniasis, respectively (<u>Figure 1</u>). These diseases mainly affect economically vulnerable populations, leading to thousands of annual deaths and major public health problems ^[1].



Figure 1. Schematic representation of the biological cycle of some trypanosomatids: *Trypanosoma brucei* (lilac), *Trypanosoma cruzi* (blue), and *Leishmania* spp. (red). Invertebrate hosts: *Phlebotomine* (P), *Triatoma* sp. (T), and *Glossina* sp. (G). The different evolutionary forms of these protozoa are represented: bloodstream form (BSF); procyclic form (PCF); metacyclic forms (M); amastigotes (A); promastigotes (P); epimastigotes (E); and trypomastigotes (T).

To ensure survival during infection, these protozoa have developed sophisticated evasion mechanisms to circumvent the immune system and virulence factors that facilitate the maintenance of the parasite-host interaction. Their invasion and embedment in various tissues lead to the establishment of the chronicity of these diseases. By invading specific tissues and regions of the body and staying within them, parasites hamper the accessibility of drugs that are traditionally used to treat these diseases, which leads to low efficiency in the chronic stage and, in many cases, high toxicity. Therefore, the mammalian inability to eliminate these parasites efficiently and the lack of adequate treatment require the search for new technologies to prevent and combat these infectious diseases [2](3)(4).

In this scenario, the development of vaccines to control these infectious diseases, despite representing a major challenge from a biotechnological point of view, is an urgent need. Due to the biological complexity of these protozoa, characterized mainly by different evolutionary forms, the search for antigenic candidates for the design of vaccine prototypes represents a major issue. On the other hand, during the maintenance of the infection, these parasites cause immunity response polarization, a phenomenon that governs the resistance and susceptibility mechanisms of the infected hosts. Thus, the application of technology involving virus-like particles (VLPs) has become an interesting tool for the development of vaccines in the context of infections caused by trypanosomatids, since VLPs have the immunoadjuvant ability to manipulate immune response polarization [5].

VLPs used as vaccine adjuvants provide an alternative to current chemical adjuvants, along with an ability to carry foreign epitopes; they constitute platforms for the development of new prototypes and vaccine protocols. VLPs' immunostimulatory properties are similar to natural infections of the viruses from which they are derived, and the preservation of the native conformation is, in addition to the primary sequence, a major factor responsible for this property [G][Z].

2. Application of the Use of VLPs in the Development of New Vaccines

The structural conformation of VLPs mimics the morphology and structures of viral particles without the presence of the virus genome, and therefore, they are not infectious particles. Thus, the main advantages of using VLPs as vaccine prototypes are safety, since they are not infectious viral particles, and the ability to induce a robust immune response that is highly similar to that elicited by a natural viral infection. These two characteristics allow VLPs to be used in the development of vaccines ^[8].

Additionally, there are several ways to produce recombinant VLPs, mainly using gene expression systems in bacteria, yeasts, insect cells, and mammalian cells, among others ^[9]. The strategy for the construction and production of recombinant VLPs depends on viral biology, mainly the presence or absence of a viral envelope, and the choice of the best system for gene expression and purification of recombinant particles ^[10].

Regarding VLP-based vaccines, about a dozen are currently approved and available in Europe and the USA. These include Recombivax HB[®] ^[11] and Engerix[®], both against hepatitis B ^[12], and Gardasil[®] against human papillomavirus (HPV) ^[13]. CervarixTM against HPV is relevant to highlight, which contains VLPs produced in insect cells used in the vaccine market ^{[14][15]}. Regulatory agencies have recognized and approved this production system for its capacity to generate biotechnological products in a safe, effective, and economical way ^[16].

During the past few decades, studies on different VLP systems used against important human diseases—such as Ebola ^[17], Chikungunya ^[18], Dengue ^[19], influenza A ^[20], respiratory syncytial virus ^[21], and Zika ^[22]—as well as a therapeutic strategy for cancer of the breast ^[23], have demonstrated the potential of this technology for the production of new vaccines. More recently, two of the 64 vaccine projects in progress are based on VLPs, including those related to the COVID-19 pandemic ^[24]. This demonstrates the potential and popularization of the use of VLPs for the development of new vaccines.

In the context of the use of VLPs for the development of vaccines against protozoa, it was observed that the immunostimulatory property of VLPs with targeted epitopes is able to induce the production of protective antibodies and a high survival rate of animals infected with *Toxoplasma gondii*, the etiologic agent of toxoplasmosis ^{[25][26]}. In the context of malaria, caused by a protozoan of the genus *Plasmodium*, the presence of lasting immunity is observed when different vaccine prototypes are produced using antigens associated with VLPs ^{[27][28][29]}.

3. Vaccines to Control Diseases Caused by Trypanosomatids: What Do We Need and Where Are We?

The main challenge in the development of trypanosomatid vaccines is the adequate identification of important antigenic candidates for the generation and maintenance of protective immunity. A second challenge is related to the type and nature of the immune response induced during the immunization process. However, several studies have been performed in order to identify potential antigenic candidates for these protozoa, as well as mechanisms of activation of immunity for the development of vaccines (reviewed in Reference ^[30]). Therefore, understanding the cellular and molecular biology of these protozoa becomes essential to guide new studies in the field of vaccinology to control these parasitic diseases.

Currently, several tools are being used in the development of vaccines in the context of trypanosomatids, such as immunoinformatic, genomic, and proteomic techniques that mainly contribute to the identification of new antigenic targets and epitopes of these parasites ^{[31][32]}, as well as in the development of various immunization protocols in experimental models ^{[33][34][35][36][37][38]}. Several proteins and virulence factors are biologically and structurally shared among protozoa of the trypanosomatid family and therefore could be used as possible targets for the development of vaccines, such as surface glycoproteins ^{[39][40][41]}, cysteine proteases ^{[42][43]}, and metalloproteases ^{[44][45][46]}.

Additionally, for the development of vaccines against these trypanosomatids of medical importance, VLPs are presented as alternatives to induce immunity in a polarized way, since the mechanisms of resistance and susceptibilities of these infections are related to the immune response polarization that occurs during a natural infection. Thus, due to their constitution, they may have the ability to express specific components in amounts comparable to high quantities of these parasites and, furthermore, can present other immunogenic components, thus allowing a stronger general immune response of the organism ^[47].

The immune response polarization of the Th1 type (CD4⁺ T helper cells) is important for controlling infections caused by trypanosomatids, mainly due to the production of interferon- γ (IFN- γ) and tumor necrosis factor α (TNF- α). These cytokines induce the activation of macrophages and, consequently, an increase in the production of reactive oxygen species, which control the parasitic load during infection. Trypanosomatid parasites have the ability to reduce Th1-type immune response polarization by enhancing the production of IL-10 and IL-17 cytokines due to the release of numerous virulence factors ^[48]. On the other hand, the polarization of the Th2-type immune response is responsible for triggering the polyclonal activation of B cells that result in hypergammaglobulinemia, a phenomenon characterized by the increased production of antibodies with low specificity and affinity ^[49]. Therefore, during the parasite—host interaction, trypanosomatids have the ability to cause immune response polarization, which facilitates the parasitism relationship.

Despite the importance of VLP-based vaccines as a strategy to induce Th1-type immune response polarization, this methodology is still poorly studied in trypanosomatids. Indeed, only one study was found on Chagas disease ^[50], and only two used leishmaniasis ^{[51][52]} models. However, to date, no study has been presented in the context of African trypanosomiasis.

In recent decades, several studies have demonstrated the ability of VLPs to stimulate the activation of B cells and dendritic cells in the context of MHC class II and, consequently, the production of cytokines related to the activation of CD4⁺ T helper cells, which strongly favors the induction of humoral and cellular immunity ^{[8][53]}. In contrast, the presentation of antigens in the context of MHC class I favors the induction of cellular immunity mediated by CD8⁺ T cells and favors the induction of Th1 cells ^[54]. This phenomenon of immune response polarization (Th1 versus Th2) is an important opportunity for biomedical research, since the mechanisms of resistance and susceptibility to infections caused by trypanosomatids depend on the immune response polarization. The importance of the Th1 immunity profile during infections by *T. cruzi* and *Leishmania* spp. is due to the intracellular forms of these parasites during parasitism in the host ^[55]. In contrast, *T. brucei* does not develop an intracellular phase of infection; however, this protozoan has an antigenic variation mechanism that favors the chronicity of the infection in the host.

4. Development of Vaccines Based on VLPs: A Checklist

Based on the previous discussion, for the use of VLPs as a strategy for the development of vaccines against trypanosomatids, some criteria must be considered, represented schematically in <u>Figure 2</u>.



Figure 2. Representation of a rational scheme for the development of new vaccines based on virus-like particles (VLPs).

According to the illustration, there are some essential steps in the checklist:

- Should the chosen virus be enveloped or not? Enveloped VLPs are inherited from structured viral proteins, thus generating a particle with target antigens integrated on the external surface, whereas non-enveloped VLPs are based on the expression of one or more necessary viral proteins without inheriting them from a host ^[56]. Both have the additional possibility to integrate antigens ^[57] and additional adjuvants ^[7] in vaccine formulations.
- Which expression system is ideal? It should be noted that, depending on the system, there will be limitations such as cost and difficulty in scheduling ^[58], as well as a higher level of purification when faced with contamination from cells ^[59]. Some systems differ from mammalian cells mainly in the protein glycosylation phase ^[60] and do not give rise to a eukaryotic expression environment. The insect cell system has come to solve some problems in the production of

particulates, as it has characteristics such as a eukaryotic environment, which is required for the glycosylation of proteins: this post-translational characteristic is also found in human cells, but the cost of their application is higher ^[61]. In addition, there is malleability in large-scale cultivation and the possibility of using it for the simultaneous expression of many proteins that facilitate the assembly of VLPs, which requires a structure with a high level of safety and a special culture system ^[58].

- The structural modification of the conformational and immunological characteristics of the VLP can adjust the relative degree of the immune response for a balanced induction of humoral and cellular immunity ^[62], thereby generating an improved candidate capable of redirecting the immune response against specific targets when combined with antigens ^[63]. Thus, this modulates the balance of Th1 and Th2 immune responses to increase the specificity and affinity towards the parasite ^[48]. Therefore, conformational changes and the aggregation of antigens are necessary to analyze changes in stability that can compromise the effectiveness of VLP vaccines ^[64].
- Before choosing the antigens, it is necessary to assess the immunological challenges found in the parasites that cause Chagas disease, leishmaniasis, and sleeping sickness. They have specific mechanisms of escape from the immune system that end up reducing the specific response of the immune system. Based on this assessment, one can then envision a single vaccine for three diseases or specific vaccines for each one, as VLPs will act as transporters of antigens, with greater security than using soluble antigens ^[G]. The applicability of antigens presented in various evolutionary forms of the protozoan, as well as the attempt to administer immunizations with more than one antigen to cover all forms of life and increase the antigenic repertoire, must be considered in studies on *T. cruzi*, given the genetic variability of this parasite ^[65]. In addition, the use of the same antigens for both prophylactic and therapeutic strategies should be considered, taking into account that prophylactic and therapeutic vaccines can prevent infection and interfere with the progression of infection, respectively. For an infected person, a therapeutic vaccine with or without an association with available drugs can have a significant impact on preventing complications, as has already been reported for Chagas disease ^[66].
- After the production and necessary testing of these particles in isolation and in association with antigens, in vitro and in vivo tests need to be performed, for example, endotoxin evaluation ^[67]. In addition, a basic toxicity assessment must be performed, according to the WHO guidelines for non-clinical evaluation of vaccines ^[68]. This step can also assist in understanding the humoral and cellular immunogenicity of the isolated particles without antigen association. The choice of adjuvant must include consideration of the cost–benefit trade-off of co-stimulation, as VLPs can assume the role of self-adjuvant due to their particulate and multivalent nature, causing efficient incorporation by antigen-presenting cells (APCs) ^{[69][70]}.
- In vivo tests on immunization strategies with these specific antigens and/or with a chimeric VLP should be performed by firstly establishing the animal model, time, and doses, characterizing the humoral and cellular immune responses, and importantly, considering different routes of administration to understand whether this influences the stability of VLPs ^[16].
- The animals challenges (experimental infection) used in experiments are necessary to understand, for example, whether the formulation of specific antigens with VLP technology helps or not in the survival of those infected. After preclinical trials proceed as expected, clinical trials have a significant step in producing results that confirm a specific, lasting, and harmless response in humans.

Due to the time spent in pre-clinical and clinical trials in the context of safety for humans, the ideal model at the present moment would be based on known and/or safe marketed VLPs that could be directed against these three diseases in a formulation system. Based on the already-known mechanisms of VLP technology and the deep understanding of the immune response to these parasites, it is still a utopian model of how this strategy can act in practice.

Here, we propose VLPs that carry antigens of a single individual parasite, thereby becoming chimeric VLPs. According to some studies, intramuscular application has the highest efficiency ^[16], as it penetrates the epidermis to reach deeper layers, with a slow and gradual release of the formulation. Upon entering the organism, VLPs will be recognized as foreign bodies by APCs (antigen-presenting cells) that will mediate responses resulting from the formulation presented. In the ideal model, VLPs in association with antigens are processed and presented by MHC class II molecules and cause the stimulation of CD4+ T-helper lymphocytes, which will activate B cells and, in turn, stimulate the production of specific IgG antibodies.

When processed by APCs and presented by MHC class I to CD8+ T cells, a cytotoxic induction will be observed. Another expected response would be after the recognition of these antigens by CD4+ T cells: there will be an intensification in the production of cytokines, such as interferon- and TNF-, which will induce a more aggressive response by macrophages with a high production of ROS and activate CD8+ T cells^[71].

5. Final Considerations and Conclusions

As presented above, VLPs have contributed to the development of new vaccines to control numerous pathogens of medical importance. Therefore, we believe that this technology is promising as a vaccine adjuvant for the manipulation of the immune response during the induction of immunity, mainly in the context of complex diseases, such as infections caused by trypanosomatids. Finally, VLP-based vaccines, when properly designed and formulated, represent a promising platform for chemical or genetic conjugation of parasitic antigens and the development of multivalent vaccines against protozoa.

References

- 1. DNDi—Drugs for Neglected Diseases Initiative. Diseases. 2019. Available online: (accessed on 10 December 2020).
- 2. Pérez-Molina, J.; Molina, I. Chagas disease. Lancet 2018, 391, 82-94.
- 3. Burza, S.; Croft, S.L.; Boelaert, M. Leishmaniasis. Lancet 2018, 392, 951–970.
- 4. Kennedy, P.G.E. Update on human African trypanosomiasis (sleeping sickness). J. Neurol. 2019, 266, 2334–2337.
- 5. Chroboczek, J.; Szurgot, I.; Szolajska, E. Virus-like particles as vaccine. Acta Biochim. Pol. 2014, 61, 531–539.
- Crisci, E.; Bárcena, J.; Montoya, M. Virus-like particles: The new frontier of vaccines for animal viral infections. Veter Immunol. Immunopathol. 2012, 148, 211–225.
- Gomes, A.C.; Flace, A.; Saudan, P.; Zabel, F.; Cabral-Miranda, G.; El Turabi, A.; Manolova, V.; Bachmann, M.F. Adjusted Particle Size Eliminates the Need of Linkage of Antigen and Adjuvants for Appropriated T Cell Responses in Virus-Like Particle-Based Vaccines. Front. Immunol. 2017, 8, 226.
- 8. Grgacic, E.V.; Anderson, D.A. Virus-like particles: Passport to immune recognition. Methods 2006, 40, 60–65.
- 9. Zeltins, A. Construction and Characterization of Virus-Like Particles: A Review. Mol. Biotechnol. 2012, 53, 92–107.
- Pijlman, G.P. Enveloped virus-like particles as vaccines against pathogenic arboviruses. Biotechnol. J. 2015, 10, 659– 670.
- 11. Venters, C.; Graham, W.; Cassidy, W. Recombivax-HB: Perspectives past, present and future. Expert Rev. Vaccines 2004, 3, 119–129.
- Hieu, N.T.; Kim, K.H.; Janowicz, Z.; Timmermans, I. Comparative efficacy, safety and immunogenicity of Hepavax-Gene and Engerix-B, recombinant hepatitis B vaccines, in infants born to HBsAg and HBeAg positive mothers in Vietnam: An assessment at 2 years. Vaccine 2002, 20, 1803–1808.
- 13. McLemore, M.R. Gardasil®: Introducing the New Human Papillomavirus Vaccine. Clin. J. Oncol. Nurs. 2006, 10, 559– 560.
- 14. Paavonen, J.; Jenkins, D.; Bosch, F.X.; Naud, P.; Salmerón, J.; Wheeler, C.M.; Chow, S.-N.; Apter, D.L.; Kitchener, H.C.; Castellsague, X.; et al. Efficacy of a prophylactic adjuvanted bivalent L1 virus-like-particle vaccine against infection with human papillomavirus types 16 and 18 in young women: An interim analysis of a phase III double-blind, randomised controlled trial. Lancet 2007, 369, 2161–2170.
- Herrero, R.; Hildesheim, A.; Rodríguez, A.C.; Wacholder, S.; Bratti, C.; Solomon, D.; González, P.; Porras, C.; Jiménez, S.; Guillen, D.; et al. Rationale and design of a community-based double-blind randomized clinical trial of an HPV 16 and 18 vaccine in Guanacaste, Costa Rica. Vaccine 2008, 26, 4795–4808.
- 16. Qian, C.; Liu, X.; Xu, Q.; Wang, Z.; Chen, J.; Li, T.; Zheng, Q.; Yu, H.; Gu, Y.; Li, S.; et al. Recent Progress on the Versatility of Virus-Like Particles. Vaccines 2020, 8, 139.
- Singh, K.; Marasini, B.; Chen, X.; Ding, L.; Wang, J.-J.; Xiao, P.; Villinger, F.; Spearman, P. A Bivalent, Spherical Virus-Like Particle Vaccine Enhances Breadth of Immune Responses against Pathogenic Ebola Viruses in Rhesus Macaques. J. Virol. 2020, 94.
- Metz, S.W.; Pijlman, G.P. Production of Chikungunya Virus-Like Particles and Subunit Vaccines in Insect Cells. In Chikungunya Virus: Methods and Protocols; Humana Press: Totowa, NJ, USA, 2016; Volume 1426, pp. 297–309.
- 19. Metz, S.W.; Thomas, A.; White, L.; Stoops, M.; Corten, M.; Hannemann, H.; De Silva, A.M. Dengue virus-like particles mimic the antigenic properties of the infectious dengue virus envelope. Virol. J. 2018, 15, 1–10.
- 20. Pillet, S.; Couillard, J.; Trépanier, S.; Poulin, J.-F.; Yassine-Diab, B.; Guy, B.; Ward, B.J.; Landry, N. Immunogenicity and safety of a quadrivalent plant-derived virus like particle influenza vaccine candidate—Two randomized Phase II clinical trials in 18 to 49 and ≥50 years old adults. PLoS ONE 2019, 14, e0216533.

- Huertas-Díaz, M.C.; Phan, S.; Elson, A.; Nuñez, I.; Wei, H.; Sakamoto, K.; He, B. Parainfluenza virus 5 (PIV5) amplifying virus-like particles expressing respiratory syncytial virus (RSV) antigens protect mice against RSV infection. Vaccine 2019, 37, 2925–2934.
- 22. Boigard, H.; Alimova, A.; Martin, G.R.; Katz, A.; Gottlieb, P.; Galarza, J.M. Zika virus-like particle (VLP) based vaccine. PLoS Negl. Trop. Dis. 2017, 11, e0005608.
- 23. Jiménez-Chávez, Á.d.J.; Moreno-Fierros, L.; Bustos-Jaimes, I. Therapy with multi-epitope virus-like particles of B19 parvovirus reduce tumor growth and lung metastasis in an aggressive breast cancer mouse model. Vaccine 2019, 37, 7256–7268.
- 24. World Health Organization. DRAFT Landscape of COVID-19 Candidate Vaccines. 2021. Available online: (accessed on 5 January 2021).
- 25. Lee, S.-H.; Kang, H.-J.; Lee, D.-H.; Kang, S.-M.; Quan, F.-S. Virus-like particle vaccines expressing Toxoplasma gondii rhoptry protein 18 and microneme protein 8 provide enhanced protection. Vaccine 2018, 36, 5692–5700.
- 26. Choi, W.H.; Park, J.S. Immunogenicity and Protective Effect of a Virus-Like Particle Containing the SAG1 Antigen of Toxoplasma gondii as a Potential Vaccine Candidate for Toxoplasmosis. Biomedicines 2020, 8, 91.
- 27. Chichester, J.A.; Green, B.J.; Jones, R.M.; Shoji, Y.; Miura, K.; Long, C.A.; Lee, C.K.; Ockenhouse, C.F.; Morin, M.J.; Streatfield, S.J.; et al. Safety and immunogenicity of a plant-produced Pfs25 virus-like particle as a transmission blocking vaccine against malaria: A Phase 1 dose-escalation study in healthy adults. Vaccine 2018, 36, 5865–5871.
- Janitzek, C.M.; Peabody, J.; Thrane, S.; Carlsen, P.H.R.; Theander, T.G.; Salanti, A.; Chackerian, B.; Nielsen, M.A.; Sander, A.F. A proof-of-concept study for the design of a VLP-based combinatorial HPV and placental malaria vaccine. Sci. Rep. 2019, 9, 5260.
- Yenkoidiok-Douti, L.; Williams, A.E.; Canepa, G.E.; Molina-Cruz, A.; Barillas-Mury, C. Engineering a Virus-Like Particle as an Antigenic Platform for a Pfs47-Targeted Malaria Transmission-Blocking Vaccine. Sci. Rep. 2019, 9, 16833– 16839.
- 30. De Castro, M.C.A.B.; de Albuquerque, S.D.C.G.; Tavares, D.C.; De Paiva-Cavalcanti, M.; Pereira, V.A. Vaccines Against Trypanosomatids. In Tropical Diseases: An Overview of Major Diseases Occurring in the Americas; Bentham eBooks; Betham Science Publishers: Sharjah, United Arab Emirates, 2017; pp. 88–113.
- 31. Michel-Todó, L.; Bigey, P.; Reche, P.; Pinazo, M.-J.; Gascón, J.; Alonso-Padilla, J. Design of an Epitope-Based Vaccine Ensemble for Animal Trypanosomiasis by Computational Methods. Vaccines 2020, 8, 130.
- 32. Oli, A.N.; Obialor, W.O.; Ifeanyichukwu, M.O.; Odimegwu, D.C.; Okoyeh, J.N.; Emechebe, G.O.; Adejumo, S.A.; Ibeanu, G.C. Immunoinformatics and Vaccine Development: An Overview. ImmunoTargets Ther. 2020, 9, 13–30.
- 33. Rios, L.E.; Vázquez-Chagoyán, J.C.; Pacheco, A.O.; Zago, M.P.; Garg, N.J. Immunity and vaccine development efforts against Trypanosoma cruzi. Acta Trop. 2019, 200, 105168.
- 34. Dumonteil, E.; Herrera, C.; Tu, W.; Goff, K.; Fahlberg, M.; Haupt, E.; Kaur, A.; Marx, P.A.; Ortega-Lopez, J.; Hotez, P.J.; et al. Safety and immunogenicity of a recombinant vaccine against Trypanosoma cruzi in Rhesus macaques. Vaccine 2020, 38, 4584–4591.
- 35. Bivona, A.E.; Alberti, A.S.; Cerny, N.; Trinitario, S.N.; Malchiodi, E.L. Chagas disease vaccine design: The search for an efficient Trypanosoma cruzi immune-mediated control. Biochim. Biophys. Acta Mol. Basis Dis. 2020, 1866, 165658.
- 36. Ghorbani, M.; Farhoudi, R. Leishmaniasis in humans: Drug or vaccine therapy? Drug Des. Dev. Ther. 2017, 12, 25-40.
- 37. Thomaz-Soccol, V.; Da Costa, E.S.F.; Karp, S.G.; Letti, L.A.J.; Soccol, C.R.; Soccol, F.T. Recent Advances in Vaccines Against Leishmania Based on Patent Applications. Recent Patents Biotechnol. 2017, 11, 21–32.
- Black, S.J.; Mansfield, J.M. Prospects for vaccination against pathogenic African trypanosomes. Parasite Immunol. 2016, 38, 735–743.
- Jackson, A.P. The Evolution of Amastin Surface Glycoproteins in Trypanosomatid Parasites. Mol. Biol. Evol. 2009, 27, 33–45.
- 40. Jackson, A.P. Gene family phylogeny and the evolution of parasite cell surfaces. Mol. Biochem. Parasitol. 2016, 209, 64–75.
- 41. Pech-Canul, Á.D.L.C.; Monteón, V.; Solís-Oviedo, R.-L. A Brief View of the Surface Membrane Proteins fromTrypanosoma cruzi. J. Parasitol. Res. 2017, 2017, 1–13.
- 42. Rafati, S.; Fasel, N.; Masina, S. Leishmania Cysteine Proteinases: From Gene to Subunit Vaccine. Curr. Genom. 2003, 4, 253–261.
- 43. Abaza, S.Z. Expression of cysteine proteinases and cystatins in parasites and use of cysteine proteinase inhibitors in parasitic diseases. Part III: Protozoa (3): Kinetoplastids. Parasitol. United J. 2019, 12, 163–186.

- 44. Soares, R.P.; Altoé, E.C.F.; Ennes-Vidal, V.; Da Costa, S.M.; Rangel, E.F.; De Souza, N.A.; Da Silva, V.C.; Volf, P.; D'Avila-Levy, C.M. In Vitro Inhibition of Leishmania Attachment to Sandfly Midguts and LL-5 Cells by Divalent Metal Chelators, Anti-gp63 and Phosphoglycans. Protist 2017, 168, 326–334.
- 45. Moreno, C.J.G.; Torres, T.; Silva, M.S. Variable Surface Glycoprotein from Trypanosoma brucei Undergoes Cleavage by Matrix Metalloproteinases: An in silico Approach. Pathogens 2019, 8, 178.
- 46. Parthasarathy, A.; Kalesh, K. Defeating the trypanosomatid trio: Proteomics of the protozoan parasites causing neglected tropical diseases. RSC Med. Chem. 2020, 11, 625–645.
- 47. Sarkar, B.; Islam, S.S.; Zohora, U.S.; Ullah, M.A. Virus like particles-A recent advancement in vaccine development. Korean J. Microbiol. 2019, 55, 327–343.
- 48. Silva-Barrios, S.; Charpentier, T.; Stäger, S. The Deadly Dance of B Cells with Trypanosomatids. Trends Parasitol. 2018, 34, 155–171.
- 49. Silva-Barrios, S.; Stäger, S. Hypergammaglobulinemia sustains the development of regulatory responses during chronic Leishmania donovani infection in mice. Eur. J. Immunol. 2019, 49, 1082–1091.
- Fernández-Presas, A.M.; Padilla-Noriega, L.; Robert, L.; Jiménez, J.A.; Solano, S.; Delgado, J.; Tato, P.; Molinari, J.L.; Becker, I. Enveloped and non-enveloped viral-like particles in Trypanosoma cruzi epimastigotes. Rev. Inst. Med. Trop. São Paulo 2017, 59.
- 51. Moura, A.P.V.; Santos, L.C.B.; Brito, C.R.N.; Valencia, E.; Junqueira, C.; Filho, A.A.P.; Sant'Anna, M.R.V.; Gontijo, N.F.; Bartholomeu, D.C.; Fujiwara, R.T.; et al. Virus-like Particle Display of the α-Gal Carbohydrate for Vaccination againstLeishmaniaInfection. ACS Cent. Sci. 2017, 3, 1026–1031.
- 52. Cecilio, P.; Pérez-Cabezas, B.; Fernandez, L.; Moreno, J.; Carrillo, E.; Requena, J.M.; Fichera, E.; Reed, S.G.; Coler, R.N.; Kamhawi, S.; et al. Pre-clinical antigenicity studies of an innovative multivalent vaccine for human visceral leishmaniasis. PLoS Negl. Trop. Dis. 2017, 11, e0005951.
- 53. Chackerian, B. Virus-like particles: Flexible platforms for vaccine development. Expert Rev. Vaccines 2007, 6, 381–390.
- 54. Deml, L.; Speth, C.; Dierich, M.P.; Wolf, H.; Wagner, R. Recombinant HIV-1 Pr55gag virus-like particles: Potent stimulators of innate and acquired immune responses. Mol. Immunol. 2005, 42, 259–277.
- 55. Pavanelli, W.R. Citocinas e óxido nítrico na Doença de Chagas. Biosaúde 2016, 10, 147–156.
- 56. Zlotnick, A.; Mukhopadhyay, S. Virus assembly, allostery and antivirals. Trends Microbiol. 2011, 19, 14–23.
- Visciano, M.; Diomede, L.; Tagliamonte, M.; Tornesello, M.L.; Asti, V.; Bomsel, M.; Buonaguro, F.; Lopalco, L.; Buonaguro, L. Generation of HIV-1 Virus-Like Particles expressing different HIV-1 glycoproteins. Vaccine 2011, 29, 4903–4912.
- 58. Naskalska, A.; Pyrć, K. Virus like particles as immunogens and universal nanocarriers. Pol. J. Microbiol. 2015, 64, 3– 13.
- 59. Hervas-Stubbs, S.; Rueda, P.; Lopez, L.; Leclerc, C. Insect Baculoviruses Strongly Potentiate Adaptive Immune Responses by Inducing Type I IFN. J. Immunol. 2007, 178, 2361–2369.
- 60. Balke, I.; Zeltins, A. Recent Advances in the Use of Plant Virus-Like Particles as Vaccines. Viruses 2020, 12, 270.
- Roy, P.; Noad, R. Virus-Like Particles as a Vaccine Delivery System: Myths and Facts. In Advances in Experimental Medicine and Biology; Springer: Berlin, Germany, 2009; Volume 655, pp. 145–158.
- Jeong, H.; Seong, B.L. Exploiting virus-like particles as innovative vaccines against emerging viral infections. J. Microbiol. 2017, 55, 220–230.
- Perotti, M.; Perez, L. Virus-Like Particles and Nanoparticles for Vaccine Development against HCMV. Viruses 2019, 12, 35.
- 64. Jain, N.K.; Sahni, N.; Kumru, O.S.; Joshi, S.B.; Volkin, D.B.; Middaugh, C.R. Formulation and stabilization of recombinant protein based virus-like particle vaccines. Adv. Drug Deliv. Rev. 2015, 93, 42–55.
- Rodríguez-Morales, O.; Monteón-Padilla, V.; Carrillo-Sánchez, S.C.; Rios-Castro, M.; Martínez-Cruz, M.; Carabarin-Lima, A.; Arce-Fonseca, M. Experimental Vaccines against Chagas Disease: A Journey through History. J. Immunol. Res. 2015, 2015, 1–8.
- 66. Beaumier, C.M.; Gillespie, P.M.; Strych, U.; Hayward, T.; Hotez, P.J.; Bottazzi, M.E. Status of vaccine research and development of vaccines for Chagas disease. Vaccine 2016, 34, 2996–3000.
- 67. Brito, L.A.; Singh, M. COMMENTARY: Acceptable Levels of Endotoxin in Vaccine Formulations During Preclinical Research. J. Pharm. Sci. 2011, 100, 34–37.

- 68. World Health Organization. WHO guidelines on nonclinical evaluation of vaccines. World Health Organ. Tech. Rep. Ser. 2005, 927, 55.
- 69. Yan, D.; Wei, Y.-Q.; Guo, H.-C.; Sun, S.-Q. The application of virus-like particles as vaccines and biological vehicles. Appl. Microbiol. Biotechnol. 2015, 99, 10415–10432.
- 70. Frietze, K.M.; Peabody, D.S.; Chackerian, B. Engineering virus-like particles as vaccine platforms. Curr. Opin. Virol. 2016, 18, 44–49.
- 71. Serradell, M.C.; Rupil, L.L.; Martino, R.A.; Prucca, C.G.; Carranza, P.G.; Saura, A.; Fernández, E.A.; Gargantini, P.R.; Tenaglia, A.H.; Petiti, J.P.; et al. Efficient oral vaccination by bioengineering virus-like particles with protozoan surface proteins. Nat. Commun. 2019, 10, 361.

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