

Arthrocnemum indicum (Willd.) Moq. Extracts

Subjects: Allergy

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Aromatic medicinal plants (AMP) with multiple targets might play a role in drug discovery and development due to their potential health-promoting effects and are a source of new pharmaceutical substances.

Keywords: Arthrocnemum indicum extracts ; halophytes ; phytochemicals ; antimicrobial ; antioxidant ; antidiabetic ; cytotoxicity

1. Introduction

Aromatic medicinal plants (AMP) with multiple targets might play a role in drug discovery and development due to their potential health-promoting effects and are a source of new pharmaceutical substances ^{[1][2][3]}. Herbal extracts and their phytochemicals have been extensively used in folkloric medicine to cure, heal, or reduce the aggressiveness of disease and treat various ailments and health disorders ^{[4][5][6][7]}. Among them, halophyte plants, known for their high salt tolerance, that grow in tidal flats, sand dunes, saline depressions, in deserts, or rocky coasts, have the potential to develop several physiological traits ^[8]. They can attenuate and protect cells from the damage caused by the accumulation of reactive oxygen species (ROS), including superoxide anion (O_2^-), singlet oxygen (O_2), peroxide (O_2^{-2}), hydrogen peroxide (H_2O_2), hydroxyl radicals (OH^*), and hydroxyl (OH^-) ions and can maintain ion homeostasis ^{[9][10]}. In addition, they can promote several biological activities implicated in preventing cancer, chronic inflammation, cardiovascular disorders, and neurodegenerative disease ^{[11][12][13][14]}. Oxidative stress has been implicated in Alzheimer's disease (AD); memory impairment in AD patients is related to the decline in the acetylcholine (ACh) level in the cholinergic system ^[15]. Therefore, AChE inhibitors are used for stabilizing the ACh neurotransmitter levels in the synaptic cleft ^[15]. On the other hand, a large number of halophyte plants have been traditionally used to reduce blood pressure (*Salsola kali* L., Chenopodiaceae), for the treatment of cancer (*Artemisia scopariae* Waldst. and Kit., Asteraceae), and microbial infections (e.g., *M. edule*, Aizoaceae) ^[16] as well as antioxidant, anti-inflammatory, and antitumoral activities ^[17].

Arthrocnemum indicum (macrostachyum) (**Figure 1**) is a stem-succulent perennial, greenish-pinkish, shrubby halophyte plant that belongs to the family of Amaranthaceae (Chenopodiaceae). These species of plant are low shrubs that grow up to 1.5 m, much-branched from the base, and frequently form mats. This plant is abundant in saltmarshes along the coastlines of Europe, South-West Asia, and North Africa ^[18]. In folkloric medicine, *A. indicum* has been commonly used to treat poisonous bites and stings and possesses beneficial effects against numerous other diseases ^[16]. The antiproliferative effect of *A. indicum* shoot (leaves and stems) extracts was compared to the control, and the results are very encouraging ^[8].



Figure 1. *A. indicum* plant.

2. Discussion

Plant-based bioactive compounds containing substantial quantities of polyphenols have been gaining much attention nowadays. Our obtained antioxidant results revealed a broad variability in antioxidant values depending on the methods used because antioxidants may exert their effect through various mechanisms. This variability was attributed to the interference of the reaction mechanism and the tested solvents. Typically, the nature of the active molecules present in the samples as well as the presence of phenolic compounds with a certain structure and particular hydroxyl position in the molecule, which can act as a proton donor and show radical scavenging activity. Parallel to that, our antibacterial results showed that Gram-negative bacteria are more resistant than Gram-positive bacteria to the various extracts, especially the ethanol, due to their distinctive structure and to the bacterium's outer-membrane barrier for Gram-negative bacteria. Their resistance was amplified via chromosomal mutations and lateral gene transfers.

The ethanol extract of *A. indicum* shoots as the most active was dominated by TPC and TFC, which was well supported by LC-MS analysis with the major secondary metabolite being *trans*-ferulic acid with the contents of 7432.51 ± 27.407 $\mu\text{g/g}$ extract, followed by *p*-coumaric acid (5982.57 ± 1.37 $\mu\text{g/g}$ extract), respectively. TPC and TFC are widely present in plant extracts and have been considered significant contributors to their biological activities, exclusively due to their unique redox properties [19]. Therefore, polyphenols containing hydrogen-donating groups have the ability to react with oxidants [20]. Phenolic compounds can also intervene as a potential free radical scavenger by blocking the ROS-induced cytotoxicity and simultaneously decreasing lipid peroxidation and DNA damage [21]. The high level of TPC in ethanol, 80%, might be related to its capacity to solubilize more secondary metabolites displaying a polar character and the higher solubility of a lot of extractable bioactive molecules in this solvent. The highest antioxidant activity of *A. indicum* shoot extracts towards the DPPH test may be due to its polyphenol contents. These two compounds might be greatly involved in the biological activity of this extract, with the others minor by the synergism effect. *Trans*-ferulic acid (4-hydroxy-3-methoxycinnamic acid), which is known for its potent antioxidant activity, is found in many food products and fruits and is used in cosmetology [22]. The safety of ferulic acid has been demonstrated with evidence that a high level of ferulic acid (0.5 and 1 mM) does not affect the cell viability in 786-O human renal cancer cells [23]. Besides that, the anticancer activity of ferulic acid has been proven against different cancer cells, including breast cancer cells (MCF-7) and liver cancer cells (HepG2) [24], human urinary bladder carcinoma (T24) [25], human osteosarcoma (143B and MG63) [26], human breast cancer (MDA-MB-231) [27], and human renal adenocarcinoma (ACHN) cells [28]. Additionally, the inhibition of A549 and HT29-D4 cancer cells was induced by ferulic acid [29]. Ferulic acid has been proven previously for its antioxidant activity, which was mainly related to its resonance stabilization [30]. Ferulic acid helps in neutralizing the free radicals. Bami et al. [30] suggested ferulic acid can hamper oxidative stress and regulate the levels of protein nitrotyrosine, malondialdehyde, blood urea nitrogen, myeloperoxidase, total antioxidant status, and creatinine in rats treated with cisplatin. Alam et al. [31] reported that ferulic acid improves cardiovascular and kidney structure. It was able to decrease the hydrophobicity of *P. aeruginosa* [32]. In the study of Ijabadeniyi et al. [33], ferulic acid was well demonstrated for its antimicrobial activity [33]. Merkl et al. [34] stated that ferulic acid could inhibit the growth of *E. coli*. In addition to all of the above, ferulic acid has been proven for its neuroprotective and antidiabetic properties as well as having high synergistic interaction with hypoglycemic drugs [35][36][37].

The second major identified compound in *A. indicum* shoot ethanol extract, *p*-coumaric acid (4-hydroxycinnamic acid), is a natural ligand abundant in many fruits, vegetables, and cereals with diverse health benefits. The safety of *p*-coumaric acid has been investigated, and the results outlined no significant cytotoxicity [38]. Previous studies have demonstrated the significant relationship between *p*-coumaric acid and antioxidant and antihyperlipidemic activities [39]. The authors suggested that *p*-coumaric is a potent antioxidant with potential therapeutic efficacy for treating hyperlipidemia symptoms [40]. Kilic et al. [39] reported that it is a good scavenger. Besides that, the antimicrobial role of *p*-coumaric acid has been proven. Boz et al. [41] demonstrated the antimicrobial activity of *p*-coumaric acid allows the disrupting of bacterial cell membranes [41]. *p*-coumaric acid was found to inhibit the proliferation and migration of cancer cells [42]. Moreover, the chemopreventive effects of *p*-coumaric acid on colon cancer have been illustrated [43].

The third predominant compound was found to be rutin (3,30,40,5,7-pentahydroxyflavone-3-rhamnoglucoside), which exists in high levels in ethanolic extract (4108.17 ± 14.31 $\mu\text{g/g}$ extract) and acetonic extract (7987.96 ± 18.73 $\mu\text{g/g}$ extract), and must be taken into account. Rutin has been verified for its carcinogenicity, and data showed no carcinogenic potential in non-inbred golden hamsters. In fact, the flavonol rutin has been studied for its antidiabetic effect. It was added for glycemic control by increasing the insulin receptor kinase property [44]. Also, it possesses a protective effect on hepatic and cardiac toxicity [45]. The pharmacological properties of rutin have also been widely studied, including its antileukemic potential [46], anti-inflammatory, antimicrobial, anticarcinogenic, neuroprotective, antithrombotic, and antiviral activities [47][48].

References

1. Ben Mefteh, F.; Daoud, A.; Bouket, A.C.; Thissera, B.; Kadri, Y.; Cherif-Silini, H.; Eshelli, M.; Alenezi, F.N.; Vallat, A.; Oszako, T.; et al. Date Palm Trees Root-Derived Endophytes as Fungal Cell Factories for Diverse Bioactive Metabolites. *Int. J. Mol. Sci.* 2018, 19, 1986.
2. Hajlaoui, H.; Arraouadi, S.; Noumi, E.; Aouadi, K.; Adnan, M.; Khan, M.A.; Kadri, A.; Snoussi, M. Antimicrobial, Antioxidant, Anti-Acetylcholinesterase, Antidiabetic, and Pharmacokinetic Properties of *Carum carvi* L. and *Coriandrum sativum* L. Essential Oils Alone and in Combination. *Molecules* 2021, 26, 3625.
3. Mseddi, K.; Alimi, F.; Noumi, E.; Veettil, V.N.; Deshpande, S.; Adnan, M.; Hamdi, A.; Elkahoui, S.; Alghamdi, A.; Kadri, A.; et al. *Thymus musilii* Velen. as a promising source of potent bioactive compounds with its pharmacological properties: In vitro and in silico analysis. *Arab. J. Chem.* 2020, 13, 6782–6801.
4. Daoud, A.; Ben Mefteh, F.; Mnafigui, K.; Turki, M.; Jmal, S.; Ben Amar, R.; Ayadi, F.; ElFeki, A.; Abid, L.; Rateb, M.E.; et al. Cardiopreventive effect of ethanolic extract of date palm pollen against isoproterenol induced myocardial infarction in rats through the inhibition of the angiotensin-converting enzyme. *Exp. Toxicol. Pathol.* 2017, 69, 656–665.
5. Gad-Elkareem, M.A.M.; Abdelgadir, E.H.; Badawy, O.M.; Kadri, A. Potential antidiabetic effect of ethanolic and aqueous-ethanolic extracts of *Ricinus communis* leaves on streptozotocin-induced diabetes in rats. *PeerJ* 2019, 7, e6441.
6. Felhi, S.; Hajlaoui, H.; Ncir, M.; Bakari, S.; Ktari, N.; Saoudi, M.; Gharsallah, N.; Kadri, A. Nutritional, phytochemical and antioxidant evaluation and FT-IR analysis of freeze-dried extracts of *Ecballium elaterium* fruit juice from three localities. *Food Sci. Technol.* 2016, 36, 646–655.
7. Felhi, S.; Daoud, A.; Hajlaoui, H.; Mnafigui, K.; Gharsallah, N.; Kadri, A. Solvent extraction effects on phytochemical constituents profiles, antioxidant and antimicrobial activities and functional group analysis of *Ecballium elaterium* seeds and peels fruits. *Food Sci. Technol.* 2017, 37, 483–492.
8. Ksouri, R.; Ksouri, W.M.; Jallali, I.; Debez, A.; Magné, C.; Hiroko, I.; Abdely, C. Medicinal halophytes: Potent source of health promoting biomolecules with medical, nutraceutical and food applications. *Crit. Rev. Biotechnol.* 2012, 32, 289–326.
9. Aouadi, K.; Hajlaoui, H.; Arraouadi, S.; Ghannay, S.; Snoussi, M.; Kadri, A. HPLC/MS Phytochemical Profiling with Antioxidant Activities of *Echium humile* Desf. Extracts: ADMET Prediction and Computational Study Targeting Human Peroxiredoxin 5 Receptor. *Agronomy* 2021, 11, 2165.
10. Sajkowska-Kozielewicz, J.J.; Kozielewicz, P.; Barnes, N.M.; Wawer, I.; Paradowska, K. Antioxidant, cytotoxic, and antiproliferative activities and total polyphenol contents of the extracts of *Geissospermum reticulatum* bark. *Oxid. Med. Cell. Longev.* 2016, 2016, 2573580.
11. Hajlaoui, H.; Arraouadi, S.; Mighri, H.; Chaaibia, M.; Gharsallah, N.; Ros, G.; Nieto, G.; Kadri, A. Phytochemical Constituents and Antioxidant Activity of *Oudneya Africana* L. Leaves Extracts: Evaluation Effects on Fatty Acids and Proteins Oxidation of Beef Burger during Refrigerated Storage. *Antioxidants* 2019, 8, 442.
12. Felhi, S.; Saoudi, M.; Daoud, A.; Hajlaoui, H.; Ncir, M.; Chaabane, R.; El Feki, A.; Gharsallah, N.; Kadri, A. Investigation of phytochemical contents, in vitro antioxidant and antibacterial behavior and in vivo anti-inflammatory potential of *Ecballium elaterium* methanol fruits extract. *Food Sci. Technol.* 2017, 37, 558–563.
13. Bakari, S.; Hajlaoui, H.; Daoud, A.; Mighri, H.; Ross-Garcia, J.M.; Gharsallah, N.; Kadri, A. Phytochemicals, antioxidant and antimicrobial potentials and LC-MS analysis of hydroalcoholic extracts of leaves and flowers of *Erodium glaucophyllum* collected from Tunisian Sahara. *Food Sci. Biotechnol.* 2018, 38, 310–317.
14. Bakari, S.; Daoud, A.; Felhi, S.; Smaoui, S.; Gharsallah, N.; Kadri, A. Proximate analysis, mineral composition, phytochemical contents, antioxidant and antimicrobial activities and GC-MS investigation of various solvent extracts of cactus cladode. *Food Sci. Technol.* 2017, 27, 286–293.
15. Takomthong, P.; Waiwut, P.; Yenjai, C.; Sombatsri, A.; Reubroycharoen, P.; Lei, L.; Lai, R.; Chaiwiwatrakul, S.; Boonyarat, C. Multi-Target Actions of Acridones from *Atalantia monophylla* towards Alzheimer's Pathogenesis and Their Pharmacokinetic Properties. *Pharmaceutics* 2021, 14, 888.
16. Hurtado-Fernandez, E.; Pacchiarotta, T.; Mayboroda, O.A.; Fernandez-Gutierrez, A.; Carrasco-Pancorbo, A. Quantitative characterization of important metabolites of avocado fruit by gas chromatography coupled to different detectors (APCI-TOF MS and FID). *Food Res. Int.* 2014, 62, 801–811.
17. Liebezeit, G.; Künnemann, T.D.; Gad, G. Biotechnological potential of North sea salt marsh plants—a review of traditional knowledge. *Prog. Ind. Microbiol.* 1999, 70, 77–84.

18. Redondo-Gómez, S.; Mateos-Naranjo, E.; Figueroa, M.E.; Davy, A.J. Salt stimulation of growth and photosynthesis in an extreme halophyte *Arthrocnemum macrostachyum*. *Plant Biol.* 2010, 12, 79–87.
19. Muflihah, Y.M.; Gollavelli, G.; Ling, Y.-C. Correlation Study of Antioxidant Activity with Phenolic and Flavonoid Compounds in 12 Indonesian Indigenous Herbs. *Antioxidants* 2021, 10, 1530.
20. Kumar, A.; Kaushik, P.; Incerpi, S.; Pedersen, J.Z.; Goel, S.; Prasad, A.K.; Rohil, V.; Parmar, V.S.; Saso, L.; Len, C. Evaluation of the Free Radical Scavenging Activities of Ellagic Acid and Ellagic Acid Peracetate by EPR Spectrometry. *Molecules* 2021, 26, 4800.
21. Majtan, J. Honey: An immunomodulator in wound healing. *Wound Repair Regen.* 2014, 22, 187–192.
22. Kelainy, E.G.; Laila, I.M.I.; Ibrahim, S.R. The effect of ferulic acid against lead-induced oxidative stress and DNA damage in kidney and testes of rats. *Environ. Sci. Poll. Res.* 2019, 26, 31675–31684.
23. Caparica, R.; Júlio, A.; Baby, A.R.; de Almeida, T.S.; Costa, J.G. In vitro cytotoxicity assessment of ferulic, caffeic and p-coumaric acids on human renal cancer cells. *Biomed. Biopharm. Res.* 2020, 17, 63–74.
24. ElKhazendar, M.; Chalak, J.; El-Huneidi, W.; Vinod, A.; Abdel-Rahman, W.M.; Abu-Gharbieh, E. Antiproliferative and proapoptotic activities of ferulic acid in breast and liver cancer cell lines. *Trop. J. Pharm. Res.* 2019, 18, 2571–2576.
25. Peng, C.C.; Chyau, C.C.; Wang, H.E.; Chang, C.H.; Chen, K.C.; Chou, K.Y.; Peng, R.Y. Cytotoxicity of ferulic acid on T24 cell line differentiated by different microenvironments. *BioMed Res. Int.* 2013, 2013, 579859.
26. Wang, T.; Gong, X.; Jiang, R.; Li, H.; Du, W.; Kuang, G. Ferulic acid inhibits proliferation and promotes apoptosis via blockage of PI3K/Akt pathway in osteosarcoma cell. *Am. J. Transl. Res.* 2016, 8, 968–980.
27. Zhang, X.; Lin, D.; Jiang, R.; Li, H.; Wan, J.; Li, H. Ferulic acid antitumor activity and inhibits metastasis in breast cancer cells by regulating epithelial to mesenchymal transition. *Oncol. Rep.* 2016, 36, 271–278.
28. Karimvand, M.N.; Kalantar, H.; Khodayar, M.J. Cytotoxic and apoptotic effects of ferulic acid on renal carcinoma cell line (ACHN). *Jundishapur J. Nat. Pharm. Prod.* 2021, 15, e81969.
29. Bouzaiane, N.N.; Kilani Jaziri, S.; Kovacic, H.; Chekir-Ghedira, L.; Ghedira, K.; Luis, J. The effects of caffeic, coumaric and ferulic acids on proliferation, superoxide production, adhesion and migration of human tumor cells in vitro. *Eur. J. Pharmacol.* 2015, 766, 99–105.
30. Bami, E.; Ozakpinar, O.B.; Ozdemir-Kumral, Z.N.; Köroglu, K.; Ercan, F.; Cirakli, Z.; Sekerler, T.; Izzettin, F.V.; Sancar, M.; Okuyan, B. Protective effect of ferulic acid on cisplatin induced nephrotoxicity in rats. *Environ. Toxicol. Pharmacol.* 2017, 54, 105–111.
31. Alam, M.A.; Sernia, C.; Brown, L. Ferulic Acid Improves Cardiovascular and Kidney Structure and Function in Hypertensive Rats. *J. Cardiovasc. Pharmacol.* 2013, 61, 240–249.
32. Borges, A.; Ferreira, C.; Saavedra, M.J.; Simões, M. Antibacterial Activity and Mode of Action of Ferulic and Gallic Acids Against Pathogenic Bacteria. *Microb. Drug Resist.* 2013, 19, 256–265.
33. Ijabadeniyi, O.A.; Govender, A.; Olagunju, O.F.; Oyediji, A.B. The antimicrobial activity of two phenolic acids against foodborne *Escherichia coli* and *Listeria monocytogenes* and their effectiveness in a meat system. *Ital. J. Food Sci.* 2021, 33, 39–45.
34. Merkl, R.; Hrádková, I.; Filip, V.; Šmidrkal, J. Antimicrobial and antioxidant properties of phenolic acids alkyl esters. *Czech J. Food Sci.* 2010, 28, 275–279.
35. Ren, Z.; Zhang, R.; Li, Y.; Li, Y.; Yang, Z.; Yang, H. Ferulic acid exerts neuroprotective effects against cerebral ischemia/reperfusion-induced injury via antioxidant and anti-apoptotic mechanisms in vitro and in vivo. *Int. J. Mol. Med.* 2017, 40, 1444–1456.
36. Narasimhan, A.; Chinnaiyan, M.; Karundevi, B. Ferulic acid exerts its antidiabetic effect by modulating insulin-signalling molecules in the liver of high-fat diet and fructose-induced type-2 diabetic adult male rat. *Appl. Physiol. Nutr. Metab.* 2015, 40, 769–781.
37. Prabhakar, P.K.; Prasad, R.; Ali, S.; Doble, M. Synergistic interaction of ferulic acid with commercial hypoglycemic drugs in streptozotocin induced diabetic rats. *Phytomedicine* 2013, 20, 488–494.
38. Boo, Y.C. p-Coumaric Acid as An Active Ingredient in Cosmetics: A Review Focusing on its Antimelanogenic Effects. *Antioxidants* 2019, 8, 275.
39. Yingbin, S.; Xun, S.; Li, L.; Jian, S.; Yogini, J.; Junqing, H.; Chun, L.; Wenjian, Y.; Leonard, W.; Hui, Z.; et al. Protective effects of p-coumaric acid against oxidant and hyperlipidemia-an in vitro and in vivo evaluation. *Biomed. Pharmacother.* 2019, 111, 579–587.
40. Kilic, I.; Yesiloglu, Y. Spectroscopic studies on the antioxidant activity of p-coumaric acid. *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* 2013, 115, 719–724.

41. Boz, H. p-Coumaric acid in cereals: Presence, antioxidant and antimicrobial effects. *Int. J. Food Sci. Technol.* 2015, 50, 2323–2328.
42. Roy, N.; Narayanankutty, A.; Nazeem, P.A.; Valsalan, R.; Babu, T.D.; Mathew, D. Plant Phenolics Ferulic Acid and P-Coumaric Acid Inhibit Colorectal Cancer Cell Proliferation through EGFR Down-Regulation. *Asian Pac. J. Cancer Prev.* 2016, 17, 4019–4023.
43. Sharma, S.H.; Rajamanickam, V.; Nagarajan, S. Antiproliferative effect of p-Coumaric acid targets UPR activation by downregulating Grp78 in colon cancer. *Chem. Biol. Interact.* 2018, 291, 16–28.
44. Hsu, C.Y.; Shih, H.Y.; Chia, Y.C.; Lee, C.H.; Ashida, H.; Lai, Y.K.; Weng, C.F. Rutin potentiates insulin receptor kinase to enhance insulin-dependent glucose transporter 4 translocation. *Mol. Nutr. Food Res.* 2014, 58, 1168–1176.
45. Fernandes, A.A.; Novelli, E.L.; Okoshi, K.; Okoshi, M.P.; Di Muzio, B.P.; Guimaraes, J.F.; Fernandes Junior, A. Influence of rutin treatment on biochemical alterations in experimental diabetes. *Biomed. Pharmacother.* 2010, 64, 214–219.
46. Lin, J.P.; Yang, J.S.; Lin, J.J.; Lai, K.C.; Lu, H.F.; Ma, C.Y.; Sai-Chuen Wu, R.; Wu, K.C.; Chueh, F.S.; Gibson Wood, W.; et al. Rutin inhibits human leukemia tumor growth in a murine xenograft model in vivo. *Environ. Toxicol.* 2012, 27, 480–484.
47. Agrawal, P.K.; Agrawal, C.; Blunden, G. Rutin: A potential antiviral for repurposing as a SARS-CoV-2 main protease (Mpro) inhibitor. *Nat. Prod. Commun.* 2021, 16, 1–12.
48. Ganeshpurkar, A.; Saluja, A.K. The pharmacological potential of rutin. *Saudi Pharm. J.* 2017, 25, 149–164.

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