Wickerhamomyces Yeast Killer Toxins

Subjects: Toxicology | Infectious Diseases Contributor: Laura Giovati

Bacteriocins are ribosomally synthesized antimicrobial peptides produced from a wide variety of bacteria that inhibit the growth of similar or closely related bacterial strains. A similar phenomenon of competition is present in yeasts, based on the production of killer toxins (KTs, or mycocins) that are secreted proteins or glycoproteins capable of killing susceptible microorganisms with various mechanisms of action, through interaction with specific superficial receptors. Possible implications and applications of the yeast killer phenomenon in the fight against infectious diseases are reviewed in this work, with particular reference to some wide-spectrum killer toxins (KTs) produced by *Wickerhamomyces anomalus* and other related species.

Keywords: killer yeasts ; killer toxins ; antimicrobial activity ; medical applications ; Wickerhamomyces anomalus

1. Introduction

Competition for space, nutrients and other resources in the environment is a rather common mechanism through which different species of microorganisms can interact with or prevail over others, determining the composition of microbial communities within different ecological niches. The production and excretion of molecules with toxic activity, such as bacteriocins and yeast killer toxins, can be included among the mechanisms by which some microorganisms can harm or kill their competitors, contributing to natural selection.

Bacteriocins are ribosomally synthesized antimicrobial peptides produced from a wide variety of bacteria that inhibit the growth of similar or closely related bacterial strains. Broad-spectrum bacteriocins have also been described. Based on their structure, mode of action, mechanism of biosynthesis and self-immunity, bacteriocins could deserve serious consideration as potential alternatives to traditional antimicrobials for use in agriculture, food storage, veterinary and even human medicine ^{[1][2][3]}.

A similar phenomenon of competition is present in yeasts, based on the production of killer toxins (KTs, or mycocins) that are secreted proteins or glycoproteins capable of killing susceptible microorganisms with various mechanisms of action, through interaction with specific superficial receptors. Interestingly, killer yeasts are immune to their own KTs. Since the first description of the "killer phenomenon" in Saccharomyces cerevisiae, by Bevan and Makower in 1963 ^[4], more than 100 different species of yeasts belonging to more than forty genera, including basidiomycetes and ascomycetes, have been described as killer yeasts, thus attesting to the widespread diffusion of the phenomenon and its ecological relevance ^{[5][6]}. Numerous studies over the years have greatly contributed to clarifying the molecular characteristics of various KTs, their physiology and mode of action and the genetic determinants encoding for their production. As relatively simple eukaryotic cells, killer yeasts have also represented a valid model to study relevant aspects of eukaryotic cell biology, such as mechanisms of processing and extracellular secretion of proteins. Comprehensive reviews have been published on these subjects to which the reader is referred ^{[5][6][[7][8][9][10][11][12]}.

Although the spectrum of antimicrobial activity of KTs was initially considered limited to susceptible cells belonging to the same species as the producing yeast or to closely related species, it is now known that some KTs may be active against a great variety of eukaryotic and prokaryotic microorganisms. Taxonomically unrelated fungi, bacteria, protozoa and even achlorophyllous saprophytic algae are among the described KT-susceptible strains, including spoilage and pathogenic microorganisms involved in relevant plant, animal and human infections ^[6]. The mode of action, which still needs to be clarified for many KTs, is variable and includes cell wall or membrane damages as well as arrest of the cell cycle ^[10].

2. Wickerhamomyces anomalus Killer Strains and WaKTs

In the last two decades, there has been a continuous reclassification of yeasts, including KT-producing strains, into new genera and species. For example, based on nuclear DNA reassociation studies and phylogenetic analysis of gene sequences, species of the genus Hansenula have been reassigned to the genus Pichia and then to the genus

Wickerhamomyces ^{[13][14][15][16]}. Among the species belonging to this genus, W. anomalus has aroused particular interest in microbiology and biotechnology fields, food production and biopreservation, as well as the development of innovative therapeutics, due to its specific characteristics, adaptation properties, frequent detection in natural environments (plants, soil, fruits, animals) and involvement in various fermentation processes ^[12]. Another key feature of W. anomalus is its capability to produce and secrete KTs, characterized by a broad spectrum of activity, comprising relevant plant, animal and human pathogens ^{[18][19]}. Many KTs exert their optimal activity at acidic pH and temperatures below 30 °C, which could be a problem for biomedical application ^[5]. Over the years, various KTs produced by W. anomalus (named WaKTs or, previously, PaKTs and HaKTs) have been described ^{[20][21][22][23]}. Notably, some killer strains can produce more than one toxin with different characteristics ^[24]. The most attractive features of some of these chromosomally encoded KTs ^[25] are the broad spectrum of activity and their mechanism of action, mediated by the interaction with specific cell wall receptors. In most cases, 1,3 or 1,6 β-glucans are the potential receptors and/or targets of KTs, sometimes characterized by exo-β-glucanase activity ^{[20][21][22][28]}.

3. Killer Toxins' Medical Applications

Three possible medical applications of W. anomalus and related species killer strains or their KTs have been tested and suggested: **1**. direct use of killer yeasts as biological competitors; **2**. direct use of KTs as potential antimicrobial molecules with broad activity; **3**. production and use of immunological derivatives of KTs. In the following paragraphs, killer strains and KTs for these applications are described (summarized in **Table 1**).

Table 1. Killer strains and KTs for applications in the medical field.

| Killer Toxin (KT) | KT Producer | MM (kDa) 1 | Target | Function | Application | References |
|---------------------------------|-------------------------------------|------------------|--|---|--------------------|--|
| WaF17.12 | Wickerhamomyces anomalus F17.12 | 140 | Plasmodium berghei | β-glucanase activity | Killer yeast | [29][30][31][32] |
| Wa96603KT | W. anomalus ATCC 96603 | 220 | Bacteria, fungi, protozoa, viruses | Various/nd | KT and derivatives | [<u>33][34][35][36]</u> [<u>37][38][39]</u> |
| HM-1 | W. saturnus var. mrakii IFO 0895 | 10.7 | Yeasts | Inhibition of β-1,3-glucan synthase | KT and derivatives | [<u>40][41][42][43]</u> [<u>44][45][46][47]</u> |
| Panomycocin | W. anomalus NCYC 434 | 49 | Human dermatophytes, Candida spp. | β-glucanase activity | KT | [20][27][48][49] [50][51] |
| Wa1F1-KT | W. anomalus 1F1 | 160– 170 | Candida spp. | β-glucanase activity | КТ | [23] |
| WA40M1, WA45M2 and WA92M3 | W. anomalus WA40, WA45 and WA92 | nd | C. albicans, Acinetobacter baumannii | nd | КТ | [<u>52][53]</u> |
| КТ | W. anomalus LMA- 827 | nd | Listeria sp. | Pore formation | КТ | [54] |
| КТ | W. anomalus YF07b | 67 | Candida spp. | Membrane permeabilization | КТ | [55] |

| Killer Toxin (KT) | KT Producer | MM (kDa) 1 | Target | Function | Application | References |
|----------------------|--------------------|------------------|-----------------------|-------------------------|-------------|--------------|
| Mycocin | W. anomalus tp2-15 | 45, 50 | Candida mesorugosa | β-glucanase activity | KT | [<u>56]</u> |

¹ MM, molecular mass (kDa); nd, not determined.

4. Conclusions

A direct use of killer yeasts or their KTs as competitors against microbial pathogens is limited by several problems, except perhaps in the interesting option of their exploitation in vector insects for the symbiotic control of arthropod-borne diseases. KT-derived peptides or immunological derivatives of KTs, such as KT-mimicking Abs and their fragments, have been extensively described. KT- and Ab-derived peptides, once selected and tested, can be easily managed in terms of synthesis, quantitative production and sequence modification, in order to improve their activity and delivery systems, in view of a possible therapeutic use.

In the 21st century, infectious diseases still represent an important challenge for human health, despite the improvements in hygiene, healthcare and socioeconomic status and the extraordinary success of preventative and therapeutic approaches. Globalization and climate changes are favoring the emergence and re-emergence of new and old etiologic agents, often characterized by intrinsic or acquired resistance to anti-infective agents. The growing crowd of immunocompromised or otherwise debilitated individuals represent a further, dramatic challenge for the treatment of infectious diseases. All this severely limits the available therapeutic armamentarium, strongly highlighting the need to develop new therapeutic tools and approaches. In this scenario, KT- and Ab-derived peptides can provide leading structures for the rational design of novel, target-directed compounds applicable, by their own or in synergy with existing agents, in the field of human, animal and even plant infectious diseases.

References

- 1. Cotter, P.D.; Hill, C.; Ross, R.P. Bacteriocins: Developing innate immunity for food. Nat. Rev. Microbiol. 2005, 3, 777–78 8.
- 2. Nishie, M.; Nagao, J.; Sonomoto, K. Antibacterial peptides "bacteriocins": An overview of their diverse characteristics a nd applications. Biocontrol Sci. 2012, 17, 1–16.
- 3. Cotter, P.D.; Ross, R.P.; Hill, C. Bacteriocins—A viable alternative to antibiotics? Nat. Rev. Microbiol. 2013, 11, 95–105.
- Bevan, E.A.; Makower, M. The physiological Basis of the Killer Character in Yeast. In Genetics Today: Proceedings of t he XIth International Congress of Genetics, The Hague, The Netherlands, September 1963; Pergamon Press: Oxford, UK, 1963; Volume I, pp. 202–203.
- 5. Magliani, W.; Conti, S.; Gerloni, M.; Bertolotti, D.; Polonelli, L. Yeast killer systems. Clin. Microbiol. Rev. 1997, 10, 369–400.
- 6. Klassen, R.; Schaffrath, R.; Buzzini, P.; Ganter, P.F. Antagonistic interactions and killer yeasts. In Yeasts in Natural Eco systems: Ecology; Buzzini, P., Lachance, M.A., Yurkov, A., Eds.; Springer: Cham, Switzerland, 2017; pp. 229–275.
- 7. Marquina, D.; Santos, A.; Peinado, J.M. Biology of killer yeasts. Int. Microbiol. 2002, 5, 65-71.
- 8. Golubev, W.I. Antagonistic interactions among yeasts. In Biodiversity and Ecophysiology of Yeasts. The Yeast Handboo k, Péter, G., Rosa, C., Eds.; Springer: Berlin, Germany, 2006; pp. 197–219.
- 9. Schmitt, M.J.; Breinig, F. Yeast viral killer toxins: Lethality and self-protection. Nat. Rev. Genet. 2006, 4, 212-221.
- 10. Liu, G.; Chi, Z.; Wang, G.; Wang, Z.-P.; Li, Y.; Chi, Z.-M. Yeast killer toxins, molecular mechanisms of their action and th eir applications. Crit. Rev. Biotechnol. 2013, 35, 222–234.
- Shaffrath, R.; Meinhardt, F.; Klassen, R. Yeast killer toxins: Fundamentals and applications. In Physiology and Genetic s. The Mycota: A Comprehensive Treatise on Fungi as Experimental Systems for Basic and Applied Research; Anke, T., Schuffler, A., Eds.; Springer: Cham, Switzerland, 2018; pp. 87–118.
- 12. Boynton, P.J. The ecology of killer yeasts: Interference competition in natural habitats. Yeast 2019, 36, 473-485.

- 13. Kurtzman, C.P.; Robnett, C.J.; Basehoar-Powers, E. Phylogenetic relationships among species of Pichia, Issatchenkia and Williopsis determined from multigene sequence analysis, and the proposal of Barnettozyma gen. nov., Lindnera ge n. nov. and Wickerhamomyces gen. nov. FEMS Yeast Res. 2008, 8, 939–954.
- 14. Kurtzman, C.P. Phylogeny of the ascomycetous yeasts and the renaming of Pichia anomala to Wickerhamomyces ano malus. Antonie Leeuwenhoek 2011, 99, 13–23.
- Daniel, H.-M.; Redhead, S.A.; Schnürer, J.; Naumov, G.I.; Kurtzman, C.P. (2049–2050) Proposals to conserve the nam e Wickerhamomyces against Hansenula and to reject the name Saccharomyces sphaericus (Ascomycota: Saccharomy cotina). Taxon 2012, 61, 459–461.
- 16. May, T.W. Report of the Nomenclature Committee for Fungi-20. IMA Fungus 2017, 8, 189-203.
- 17. Walker, G.M. Pichia anomala: Cell physiology and biotechnology relative to other yeasts. Antonie Leeuwenhoek 2011, 99, 25–34.
- 18. Passoth, V.; Fredlund, E.; Druvefors, U.; Schnürer, J. Biotechnology, physiology and genetics of the yeast Pichia anom ala. FEMS Yeast Res. 2006, 6, 3–13.
- 19. Passoth, V.; Olstorpe, M.; Schnürer, J. Past, present and future research directions with Pichia anomala. Antonie van L eeuwenhoek 2011, 99, 121–125.
- 20. Izgü, F.; Altınbay, D.; Acun, T. Killer toxin of Pichia anomala NCYC 432; purification, characterization and its exo-β-1,3-g lucanase activity. Enzym. Microb. Technol. 2006, 39, 669–676.
- 21. De Ingeniis, J.; Raffaelli, N.; Ciani, M.; Mannazzu, I. Pichia anomala DBVPG 3003 Secretes a Ubiquitin-Like Protein Th at Has Antimicrobial Activity. Appl. Environ. Microbiol. 2009, 75, 1129–1134.
- 22. Coda, R.; Cassone, A.; Rizzello, C.G.; Nionelli, L.; Cardinali, G.; Gobbetti, M. Antifungal Activity of Wickerhamomyces a nomalus and Lactobacillus plantarum during Sourdough Fermentation: Identification of Novel Compounds and Long-Te rm Effect during Storage of Wheat Bread. Appl. Environ. Microbiol. 2011, 77, 3484–3492.
- 23. Giovati, L.; Santinoli, C.; Ferrari, E.; Ciociola, T.; Martin, E.; Bandi, C.; Ricci, I.; Epis, S.; Conti, S. Candidacidal Activity of a Novel Killer Toxin from Wickerhamomyces anomalus against Fluconazole-Susceptible and -Resistant Strains. Toxi ns 2018, 10, 68.
- Farkas, Z.; Márki-Zay, J.; Kucsera, J.; Vágvölgyi, C.; Golubev, I.W.; Pfeiffer, I. Characterization of two different toxins of Wickerhamomyces anomalus (Pichia Anomala) VKM Y-159. Acta Biol. Hung. 2012, 63, 277–287.
- Schneider, J.; Rupp, O.; Trost, E.; Jaenicke, S.; Passoth, V.; Goesmann, A.; Tauch, A.; Brinkrolf, K. Genome sequence of Wickerhamomyces anomalus DSM 6766 reveals genetic basis of biotechnologically important antimicrobial activitie s. FEMS Yeast Res. 2012, 12, 382–386.
- 26. Sawant, A.D.; Ahearn, D.G. Involvement of a cell wall receptor in the mode of action of an anti-Candida toxin of Pichia anomala. Antimicrob. Agents Chemother. 1990, 34, 1331–1335.
- Izgü, F.; Altınbay, D.; Türeli, A.E. In vitro activity of panomycocin, a novel exo-β-1,3-glucanase isolated from Pichia ano mala NCYC 434, against dermatophytes. Mycoses 2007, 50, 31–34.
- 28. Izgü, F.; Altınbay, D.; Türeli, A.E. In Vitro Susceptibilities of Candida spp. to Panomycocin, a Novel Exo-β-1,3-Glucanas e Isolated from Pichia anomala NCYC 434. Microbiol. Immunol. 2007, 51, 797–803.
- Cappelli, A.; Ulissi, U.; Valzano, M.; Damiani, C.; Epis, S.; Gabrielli, M.G.; Conti, S.; Polonelli, L.; Bandi, C.; Favia, G.; e t al. A Wickerhamomyces anomalus Killer Strain in the Malaria Vector Anopheles stephensi. PLoS ONE 2014, 9, e9598 8.
- 30. Cappelli, A.; Valzano, M.; Cecarini, V.; Bozic, J.; Rossi, P.; Mensah, P.; Amantini, C.; Favia, G.; Ricci, I. Killer yeasts exe rt anti-plasmodial activities against the malaria parasite Plasmodium berghei in the vector mosquito Anopheles stephen si and in mice. Parasites Vectors 2019, 12, 329.
- Valzano, M.; Cecarini, V.; Cappelli, A.; Capone, A.; Bozic, J.; Cuccioloni, M.; Epis, S.; Petrelli, D.; Angeletti, M.; Eleuteri, A.M.; et al. A yeast strain associated to Anopheles mosquitoes produces a toxin able to kill malaria parasites. Malar. J. 2016, 15, 1–9.
- 32. Cecarini, V.; Cuccioloni, M.; Bonfili, L.; Ricciutelli, M.; Valzano, M.; Cappelli, A.; Amantini, C.; Favia, G.; Eleuteri, A.M.; A ngeletti, M.; et al. Identification of a Killer Toxin from Wickerhamomyces anomalus with β-Glucanase Activity. Toxins 20 19, 11, 568.
- 33. Polonelli, L.; Morace, G. Reevaluation of the yeast killer phenomenon. J. Clin. Microbiol. 1986, 24, 866–869.
- Séguy, N.; Polonelli, L.; Dei-Cas, E.; Cailliez, J.C. Effect of a killer toxin of Pichia anomala to Pneumocystis. Perspectiv es in the control of pneumocystosis. FEMS Immunol. Med. Microbiol. 1998, 22, 145–149.

- Polonelli, L.; Magliani, W.; Conti, S.; Bracci, L.; Lozzi, L.; Neri, P.; Adriani, D.; De Bernardis, F.; Cassone, A. Therapeutic Activity of an Engineered Synthetic Killer Antiidiotypic Antibody Fragment against Experimental Mucosal and Systemic Candidiasis. Infect. Immun. 2003, 71, 6205–6212.
- Magliani, W.; Conti, S.; Travassos, L.R.; Polonelli, L. From yeast killer toxins to antibiobodies and beyond. FEMS Micro biol. Lett. 2008, 288, 1–8.
- Magliani, W.; Conti, S.; Ciociola, T.; Giovati, L.; Zanello, P.P.; Pertinhez, T.; Spisni, A.; Polonelli, L. Killer peptide: A novel paradigm of antimicrobial, antiviral and immunomodulatory auto-delivering drugs. Futur. Med. Chem. 2011, 3, 1209–12 31.
- 38. Polonelli, L.; Magliani, W.; Ciociola, T.; Giovati, L.; Conti, S. From Pichia anomala killer toxin through killer antibodies to killer peptides for a comprehensive anti-infective strategy. Antonie Leeuwenhoek 2010, 99, 35–41.
- Giovati, L.; Santinoli, C.; Mangia, C.; Vismarra, A.; Belletti, S.; D'Adda, T.; Fumarola, C.; Ciociola, T.; Bacci, C.; Maglian i, W.; et al. Novel Activity of a Synthetic Decapeptide Against Toxoplasma gondii Tachyzoites. Front. Microbiol. 2018, 9, 753.
- 40. Yamamoto, T.; Uchida, K.; Hiratani, T.; Miyazaki, T.; Yagiu, J.; Yamaguchi, H. In vitro activity of the killer toxin from yeas t Hansenula mrakii against yeasts and molds. J. Antibiot. 1988, 41, 398–403.
- 41. Kasahara, S.; Ben Inoue, S.; Mio, T.; Yamada, T.; Nakajima, T.; Ichishima, E.; Furuichi, Y.; Yamada, H. Involvement of c ell wall β-glucan in the action of HM-1 killer toxin. FEBS Lett. 1994, 348, 27–32.
- 42. Takasuka, T.; Komiyama, T.; Furuichi, Y.; Watanabe, T. Cell wall synthesis specific cytocidal effect of Hansenula mrakii t oxin-1 on Saccharomyces cerevisiae. Cell Mol. Biol. Res. 1995, 41, 575–581.
- 43. Komiyama, T.; Ohta, T.; Urakami, H.; Shiratori, Y.; Takasuka, T.; Satoh, M.; Watanabe, T.; Furuichi, Y. Pore Formation o n Proliferating Yeast Saccharomyces cerevisiae Cell Buds by HM-1 Killer Toxin. J. Biochem. 1996, 119, 731–736.
- Selvakumar, D.; Karim, N.; Miyamoto, M.; Furuichi, Y.; Komiyama, T. Recombinant Single-Chain Anti-idiotypic Antibody: An Effective Fungal β-1,3-Glucan Synthase Inhibitor. Biol. Pharm. Bull. 2006, 29, 1848–1853.
- 45. Selvakumar, D.; Miyamoto, M.; Furuichi, Y.; Komiyama, T. Inhibition of Fungal β-1,3-Glucan Synthase and Cell Growth by HM-1 Killer Toxin Single-Chain Anti-Idiotypic Antibodies. Antimicrob. Agents Chemother. 2006, 50, 3090–3097.
- 46. Selvakumar, D.; Miyamoto, M.; Furuichi, Y.; Komiyama, T. Inhibition of β-1,3-Glucan Synthase and Cell Growth of Crypt ococcus species by Recombinant Single-chain Anti-idiotypic Antibodies. J. Antibiot. 2006, 59, 73–79.
- Kabir, M.E.; Karim, N.; Krishnaswamy, S.; Selvakumar, D.; Miyamoto, M.; Furuichi, Y.; Komiyama, T. Peptide derived fro m anti-idiotypic single-chain antibody is a potent antifungal agent compared to its parent fungicide HM-1 killer toxin pep tide. Appl. Microbiol. Biotechnol. 2011, 92, 1151–1160.
- Izgü, F.; Altinbay, D. Isolation and Characterization of the K5-Type Yeast Killer Protein and Its Homology with an Exo-β-1,3-glucanase. Biosci. Biotechnol. Biochem. 2004, 68, 685–693.
- Izgü, F.; Altinbay, D.; Sertkaya, A. Enzymic Activity of the K5-Type Yeast Killer Toxin and Its Characterization. Biosci. Bi otechnol. Biochem. 2005, 69, 2200–2206.
- 50. Izgü, F.; Bayram, G.; Tosun, K.; Izgü, D. Stratum corneum lipid liposome-encapsulated panomycocin: Preparation, char acterization, and the determination of antimycotic efficacy against Candida spp. isolated from patients with vulvovaginiti s in an in vitro human vaginal epithelium tissue model. Int. J. Nanomed. 2017, 12, 5601–5611.
- 51. Muhammed, M.T.; Son, D.; Izgü, F. Three dimensional structure prediction of panomycocin, a novel Exo-β-1,3-glucanas e isolated from Wickerhamomyces anomalus NCYC 434 and the computational site-directed mutagenesis studies to en hance its thermal stability for therapeutic applications. Comput. Biol. Chem. 2019, 80, 270–277.
- 52. Paris, A.P.; Persel, C.; Serafin, C.F.; Simão, R.D.C.G.; Gandra, R.F. Susceptibility of Candida albicans Isolated from Blo od to Wickerhamomyces anomalous Mycocins. Curr. Microbiol. 2016, 73, 878–884.
- Junges, D.S.B.; Delabeneta, M.F.; Rosseto, L.R.B.; Nascimento, B.L.; Paris, A.P.; Persel, C.; Loth, E.A.; Simão, R.D.C.
 G.; Menolli, R.A.; Paula, C.R.; et al. Antibiotic Activity of Wickerhamomyces anomalus Mycocins on Multidrug-Resistant Acinetobacter baumannii. Microb. Ecol. 2020, 80, 278–285.
- 54. Hatoum, R.; Labrie, S.; Fliss, I. Identification and Partial Characterization of Antilisterial Compounds Produced by Dairy Yeasts. Probiotics Antimicrob. Proteins 2013, 5, 8–17.
- 55. Guo, F.-J.; Ma, Y.; Xu, H.-M.; Wang, X.-H.; Chi, Z.-M. A novel killer toxin produced by the marine-derived yeast Wickerh amomyces anomalus YF07b. Antonie Leeuwenhoek 2012, 103, 737–746.
- 56. Tay, S.-T.; Lim, S.-L.; Tan, H.-W. Growth inhibition of Candida species by Wickerhamomyces anomalus mycocin and a l actone compound of Aureobasidium pullulans. BMC Complement. Altern. Med. 2014, 14, 439.

Retrieved from https://encyclopedia.pub/entry/history/show/35683