Immunosuppression and Liver Transplantation

Subjects: Transplantation Contributor: Tsvetelina Velikova

In liver transplant patients, solid tumors and post-transplant lymphoproliferative disorders (PTLD) have emerged as significant long-term mortality causes. Additionally, it is assumed that de novo malignancy (DNM) after liver transplantation (LT) is the second-leading cause of death after cardiovascular complications. Well-established risk factors for PTLD and solid tumors are calcineurin inhibitors (CNIs), tacrolimus (TAC), and cyclosporine, the cornerstones of all immunosuppressive (IS) therapies used after LT.

liver transplantation	de novo malignancy	immunosuppressive therapy	calcineurin inhibitors
tacrolimus cyclospe	orine carcinogenesis	immune tolerance	

1. Introduction

Liver transplantation (LT) is the treatment of choice for patients with end-stage liver disease, such as decompensated liver disease, cirrhosis, acute hepatic failure, and hepatocellular cancer ^[1]. Still, the primary causes of mortality during the early post-transplantation period are infections and surgical complications ^[2]. Additionally, in liver transplant patients, solid tumors and post-transplant lymphoproliferative disorders (PTLD) have emerged as significant long-term causes of death ^[3]. Furthermore, it was estimated that liver transplant recipients have a higher risk of developing de novo malignancy (DNM) compared to the general population, with standardized incidence ratios (SIRs) ranging from 2.3 to 4.3 ^{[4][5]}.

We have to emphasize that the liver is the only solid organ in which malignancy could be an indication for transplantation; thus, cancer recurrence after LT could be a possible scenario. DNM and recurrence malignancy after LT make this topic very complex and significant. Moreover, well-established risk factors for PTLD and solid tumors are calcineurin inhibitors (CNIs), tacrolimus (TAC), and cyclosporine, the cornerstones of all immunosuppressive therapies used after LT ^[G]. Nowadays, the IS regimen improvement is one of the most critical factors leading to long-term survival and favorable outcome in patients after LT ^{[Z][B]}.

However, on the one hand, the crucial effect of IS agents leads to a significant reduction in steroid-resistant rejection rates and rejection-related graft loss ^[9]. On the other hand, more prolonged patient survival after LT is correlated with prolonged exposure to IS. Since such long-time therapy with IS is associated with unwanted adverse events, it is considered a double-edged sword ^[10]. In line with this, to prevent adverse effects, especially those related to DNM, an optimal IS regimen plays a central role in long-term survival in patients after LT.

This paper aims to overview the refined mechanisms of IS-induced tumorigenesis after LT and the loss of immunocompetence facilitated by the host immune system due to prolonged IS therapy. In addition, we also discuss in detail the mechanisms of action in different types of IS regimen used after LT and their putative effect on DNM.

2. De Novo Malignancy in Patients after Liver Transplantation

Risk factors such as aging and lifestyle of recipients, sun exposure, alcohol, and tobacco consumption before and after LT are strongly associated with the development of DNM in these patients. Other important events, such as decreased immune surveillance, activation of oncogenic viruses, and direct impact of certain classes of IS therapy, play a pivotal role in tumorigenesis after LT ^[11]. It has been reported that the overall incidence of DNM in LT ranges from 3.1 to 14.4%, depending on the period of the study and duration of IS treatment ^{[3][12]}.

A large study by Altieri et al., including 1480 recipients, elucidated the most significant risk factors leading to DNM after LT. The authors demonstrated that recipient age, male gender, deceased donor, and the type of the initial liver disease are the most critical causes of post-LT DNM ^[13]. The most common malignancy after LT is non-melanoma skin cancers, but they do not affect patient survival. More interestingly, in alcohol consumers and long-time smokers, the risk of developing DNM increases significantly due to induced DNA damage ^{[14][15]}.

A great diversity of epidemiological data came from different registry databases or single-center retrospective studies. However, reliable numbers related to the incidence of DNM after LT were obtained. Although some reports announce PTLD with the highest frequency, other sources outline non-melanoma skin cancer as the most common. **Table 1** presents estimated SIRs for the incidence of DNM in LT recipients according to the reviews by Sherston et al.

Table 1. Estimated standard incidence ratio for each malignancy after LT (adapted from Sherston et al. and Shalaby et al.) [<u>12,17</u>].

Cancer Type	SIR
Kaposi's sarcoma	61.46–100
Skin (non-melanoma)	>13.85
Head and neck cancers	2.5–24.8
PTLD	3.9–21
Thyroid	0.65–4.6
Esophagus	12–23.4
Stomach	0.5–3.7

Cancer Type	SIR
Pancreas and biliary tract	1.1-6.4
Colorectal overall	1.24–12.5
Colorectal in IBD/PSC	3–5
Colorectal in non-IBD/PSC	1–1.8
Anal cancer	3.3–10
Lung, trachea, and bronchus	1.1–8
Urological tract	0.8–10.2
Kidney	1.8–30
Prostate (in males)	0.39–2.2
Cervix (in females)	1.03–30.7
Vulva (in females)	8–23.8
Breast (in females)	0.3–2.3

SIR—standardized incidence ratio.

3. Immunosuppression and Carcinogenesis

To elucidate the impact of IS on carcinogenesis after LT, we have to pay attention to the different immune mechanisms involved in preventing cancer development. The term "immunoediting" describes a process wherein the immune system remodels tumor immunogenicity and represses tumorigenesis. Immunoediting has three component phases: elimination, equilibrium, and escape ^[16]. These tumor cells, which successfully pass this process, might prevent their destruction by the host's immune system.

The first phase of "elimination" is characterized by the immune system's ability to recognize and eliminate modified cells by distinct immune system mechanisms ^[17]. It is assumed that a growing tumor mass is a factor that initiates this process by recruiting macrophages, dendritic cells, and infiltrating lymphocytes (natural killer (NK) cells and NK T cells that release interferon-gamma, INF-γ). The latter is the main cytokine that activates the release of chemokines (e.g., CXCL9, CXCL10, and CXCL11), aiming to suppress tumor angiogenesis and to induce tumor cells' apoptosis or programmed cell death (PCD) type I. Tumor-specific dendritic cells facilitate the cytotoxic T cells (CD8+), which infiltrate the tumor after recognition of tumor-associated antigens. This process further activates other immune mechanisms to help B cells to produce specific antibodies.

When the tumor cells bypass the elimination phase of the immune system, the equilibrium phase starts. This phase is characterized by lymphocytes' ability to recognize genetically unstable and mutated tumor cells and prevent tumor expansion. In the third phase of "immunoediting," the tumor cells evade the immune system and, in an uncontrolled manner, might cause malignant transformation and direct transfer to the escape phase ^{[18][19][20]}.

Additionally, different types of nonimmune surveillance mechanisms against tumors exist, such as genetic, intraand intercellular, and epigenetic surveillance. For example, genetic surveillance is based on DNA repair and checkpoint control of the cell cycle. I PCD, and intercellular mechanisms are associated with tumor microenvironment factors. Additionally, epigenetic surveillance is a compound process of gene regulation by changing chromatin architecture, influenced by different transcription factors ^[21]

All of the above-described mechanisms, which the immune system employs to control tumors, may be disturbed by the IS modalities, leading to cancer development, including in LT patients.

4. Immunosuppressive Drugs and Their Impact on the Immune System Tumor Control

As organ transplant recipients, liver transplant patients often receive a combination of two or more IS agents to prevent allograft rejection. Nowadays, IS agents are used after LT, including cyclosporine, TAC, sirolimus, MMF, basiliximab, and corticosteroids.

These drugs are commonly used for IS induction and are considered first-line agents to treat acute rejection ^[23]. Corticosteroids are a class of nonspecific anti-inflammatory drugs, which interfere with the cytokine gene transcription. Since they hamper T cells' activation, corticosteroids decrease the inflammatory response by inhibiting pro-inflammatory cytokines such as IL-1, IL-2, IL-6, etc. However, it was recently shown that many transplant centers prefer steroid-free IS regimens because of the tremendous number of steroid-related adverse events ^[24].

So far, there is no evidence suggesting that corticosteroids do not directly influence the risk of DNM development after LT ^[23]. Furthermore, it is well known that corticosteroids increase tumor cell resistance in solid tumors by disarming B and T lymphocytes, decrease expression of major histocompatibility class I antigen in vivo, and reduce immunosurveillance ^[25]. It was also demonstrated that corticosteroids could modulate the process of immunosurveillance and promote anti-apoptotic and proliferative effects in various cell types. However, further studies have to be conducted to prove the relationship between the potential contribution of corticosteroids and the risk of cancer development in transplanted patients.

In standard patients after LT, the first choice of IS, based on CNI, is often TAC, whereas cyclosporine could be optional ^[26]. IL-2 is a well-known cytokine that plays a significant role in communication between different immune cells and is essential for their activation ^[27]. After binding to the IL-2 receptor, which has three subunits (IL-2R α (CD25), IL-2R β (CD122), and IL-2R γ (CD132)), the activation of many critical inflammatory cells is initiated. For

instance, the expression of CD25 (IL-2R α) leads to the activation and expansion of immunosuppressive regulatory T cells (Treg) ^[28].

A study by Angriman et al. demonstrated that a reduced Treg subpopulation in mice receiving TAC, cyclosporine, and mTOR inhibitors correlated with the reduced pro-tumorigenic effect of these cells ^[29]. High expression of IL- $2R\beta$ and IL- $2R\gamma$ was detected in NK cells, monocytes, macrophages, and resting CD4+ and resting CD8+ T cells. The latter could distinguish tumor cells and exert antitumor cytotoxicity ^[30]. It was well established in many studies that chronic use of CNI leads to increased risk of renal dysfunction, neurotoxicity, metabolic disorders, and DNM [31][32].

Moreover, a correlation between CNI after LT and direct pro-oncogenic activity has been reported by different groups. Tumorigenesis and tumor growth in LT patients are thought to be linked with the length of IS therapy exposure and intensity ^[33].

Further analyses revealed that the delicate mechanisms in DNM tumorigenesis after LT are associated with processes such as hindering DNA repair mechanisms, promoting tumor cell invasiveness, and hampering apoptosis ^[34].

It is noteworthy that excessive angiogenesis plays a major role in tumor expansion, where CNI could maintain this tumor angiogenesis by stimulating vascular endothelial growth factor ^[35]. Another putative oncogenic pathway of CNI is associated with overexpression of the TGF β 1 gene, which leads to tumor cell invasion and promotion of metastatic potential. Therefore, more aggressive behavior and the earlier manifestation of an oncological disease have been observed in patients after LT ^{[36][37]}.

In patients with severe diabetes, the alternative IS treatment option is monotherapy with cyclosporine. Approximately a quarter of patients after LT, who received TAC plus steroids, are suitable for this entity. It is well known that diabetes is a risk factor for developing various neoplasm types; notably, hepatocellular carcinoma (HCC) They estimated that the mean blood concentration and exposure period of TAC were major factors for developing DNM after LT ^[38].

Based on the presented data, clinicians' selection of proper IS treatment should be grounded in certain variables. Primary liver diseases, pre-LT patient status, co-morbidities, and surgical complications are among the crucial factors that impact the final IS treatment choice. A plethora of studies report the adverse effects associated with prolonged usage of CNI. Hence, strategies to reduce these adverse effects, including dose reduction or switching to another IS class, should be kept in mind by clinicians ^[26].

MMF is a prodrug compound of mycophenolic acid (MPA), a selective and reversible uncompetitive inhibitor of inosine monophosphate dehydrogenase (IMPDH) ^[39]. This pathway is crucial for the de novo synthesis of guanosine nucleotides, and it is well known that T and B lymphocytes are more dependent on this pathway than other cells ^[40]. Most of the IS guidelines recommend MMF as IS combined with another IS class, such as CNI or

mTOR inhibitors. Furthermore, MMF exhibits its antitumor activity by interfering with the tumor growth and tumor cell adhesion to vascular endothelium ^[41].

A significant scientific challenge nowadays is defining whether MMF, as an IS agent after LT, provokes malignant transformation in the human body. So far, insufficient data related to the prooncogenic effect of MMF after LT have been published. Data from the Scientific Registry of Transplant Recipients indicate that MMF use in recipients of kidney grafts is correlated with a decreased risk of PTLD or any other malignancy ^[42].

An interesting study by Koehl et al. observed the potential of MMF in animal models to inhibit tumor cell expansion and tumor angiogenesis. The study results are controversial since MMF showed only a marginal effect on tumor growth ^[43]. However, these results correspond indirectly to the data mentioned above from the Scientific Registry of Transplant Recipients.

For the first time in clinical practice, mTOR inhibitors were used as anti-cancer drugs. In LT recipients, mTOR inhibitors proved their efficacy in the prevention of immunologic rejection in organ transplantation. Nowadays, about 4% of patients after LT receive mTOR inhibitors as IS treatment combined with MMF^[44]. Another important reason for the implication of mTOR inhibitors in the post-transplant period is avoiding and/or postponing HCC recurrence in this patient population.

This IS class inhibits serine–threonine kinase, which belongs to the family of phosphatidylinositol-3 kinase (PI3K). It is thought to be involved in the cell cycle, cellular metabolism, and apoptosis ^{[45][46]}. The refined mechanisms of mTOR inhibitors include obstructing the cell cycle in the G1 phase, consequently hampering protein synthesis (**Figure 1**) ^[47]. mTORC1, through phosphorylating different kinases, regulates the expression of proteins that interfere with cell proliferation and survival.

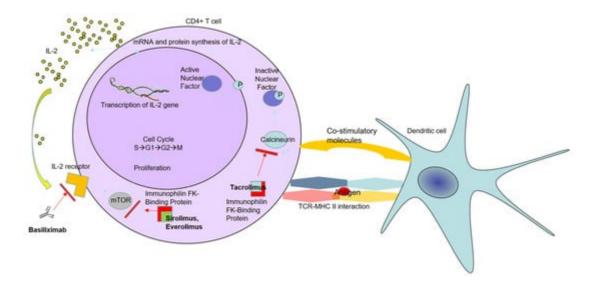


Figure 1. Mechanisms of action of tacrolimus, everolimus, and sirolimus. By attaching to immunophilin FK-binding protein and calcineurin enzyme in the T-cell cytoplasm, tacrolimus prevents the nuclear factor's dephosphorylation and transportation into the nucleus. Therefore, the synthesis of interleukin-2 (IL-2) is blocked, leading to interrupted

T-cell activation. Basiliximab, a chimeric mouse–human monoclonal antibody that binds to the IL-2 receptor (a.k.a., CD25), is another approach to block T-cell activation after liver transplantation. mTOR inhibitors such as everolimus and sirolimus, act as interrupting mTOR actions (e.g., regulating protein expression, cell proliferation, and survival). However, the mTORC2 complex may promote liver tumorigenesis by direct phosphorylation of the protein kinase Akt, essential for actin cytoskeleton regulation.

On the contrary, mTORC2 directly phosphorylates AKT (protein kinase B), therefore regulating the actin cytoskeleton and promoting liver tumorigenesis. It was established that mTORC1 is more sensitive to treatment with mTOR inhibitors than mTORC2, which is less sensitive to the drug ^[48]. As a general principle, mTOR inhibitors have many positive effects on patient outcomes and survival after LT. Thus, mTOR inhibitors showed promising therapeutic results in patients with DNM because of their ability to decrease vascular endothelial growth factor synthesis and interrupt angiogenesis ^[35].

Additionally, the antitumor effect of mTOR inhibitors has been comprehensively reported in some studies, including in patients with HCC. It was established that progression and spreading in HCC depend on the mTOR signaling pathway involved in cell metabolism and proliferation. Notably, more aggressive tumor progression and shorter survival in patients with HCC were associated with a constitutively activated PI3K/AKT/mTOR pathway ^[49]. Therefore, the available data from the literature confirmed that mTOR inhibitors could be a feasible treatment option to limit HCC recurrence and progression in LT recipients ^{[50][51][52]}.

proved that sirolimus-based IS showed significantly better outcomes for post-LT HCC patients ^[51]. Furthermore, another clinical study by Cholongitas et al., which included 3666 patients, showed the negative association between mTOR inhibitors and HCC recurrence after LT against patients on CNI therapy. Unfortunately, many clinical studies, especially retrospective ones, showed insufficient efficacy of mTOR inhibitors on HCC progression and recurrence after LT. Therefore, more studies have to be conducted to obtain a more efficient mTOR inhibitor dose, regimen, etc.

Another beneficial aspect of mTOR inhibitors in clinical practice was documented in patients who developed DNM after LT. in 850 LT patients compared LT patients with DNM who were switched to sirolimus monotherapy and LT patients who received standard IS regimens with CNI/MMF. The authors showed that sirolimus monotherapy is effective and safe, which further improves survival outcomes in LT recipients with DNM ^[53].

Regarding the incidence of skin cancer after LT, Funk-Debleds et al. demonstrated that conversion from CNI-based regimen to mTOR inhibitors could reduce the development of non-melanoma skin cancer ^[54]. The implication of mTOR inhibitors in LT leads to some benefits in renal function compared to CNI. Moreover, because of its protective cardiovascular effect, everolimus is recommended, especially in patients with diabetic nephropathy. This alternative IS regimen could be a feasible long-term treatment option for LT patients.

References

- 1. Fox, A.N.; Brown, R.S., Jr. Is the patient a candidate for liver transplantation? Clin. Liver Dis. 2012, 16, 435–448.
- Adam, R.; Karam, V.; Delvart, V.; O'Grady, J.; Mirza, D.; Klempnauer, J.; Castaing, D.; Neuhaus, P.; Jamieson, N.; Salizzoni, M.; et al. Evolution of indications and results of liver transplantation in Europe. A report from the European Liver Transplant Registry (ELTR). J. Hepatol. 2012, 57, 675– 688.
- Engels, E.A.; Pfeiffer, R.M.; Fraumeni, J.F., Jr.; Kasiske, B.L.; Israni, A.K.; Snyder, J.J.; Wolfe, R.A.; Goodrich, N.P.; Bayakly, A.R.; Clarke, C.A.; et al. Spectrum of cancer risk among US solid organ transplant recipients. JAMA 2011, 306, 1891–1901.
- Na, R.; Grulich, A.E.; Meagher, N.S.; McCaughan, G.W.; Keogh, A.M.; Vajdic, C.M. Comparison of de novo cancer incidence in Australian liver, heart and lung transplant recipients. Am. J. Transplant. 2013, 13, 174–183.
- Krynitz, B.; Edgren, G.; Lindelöf, B.; Baecklund, E.; Brattström, C.; Wilczek, H.; Smedby, K.E. Risk of skin cancer and other malignancies in kidney, liver, heart and lung transplant recipients 1970 to 2008—A Swedish population-based study. Int. J. Cancer 2013, 132, 1429–1438.
- Bakker, N.A.; van Imhoff, G.W.; Verschuuren, E.A.; van Son, W.J. Presentation and early detection of post-transplant lymphoproliferative disorder after solid organ transplantation. Transpl. Int. 2007, 20, 207–218.
- 7. Scientific Registry Transplant Recipients (SRTR). Transplant Center Search Results. Available online: (accessed on 28 June 2020).
- 8. European Liver Transplant Registry. Evolution of Liver Transplantation in Europe. Available online: (accessed on 28 June 2020).
- Adam, R.; McMaster, P.; O'Grady, J.G.; Castaing, D.; Klempnauer, J.L.; Jamieson, N.; Neuhaus, P.; Lerut, J.; Salizzoni, M.; Pollard, S.; et al. European Liver Transplant Association. Evolution of liver transplantation in Europe: Report of the European Liver Transplant Registry. Liver Transpl. 2003, 9, 1231–1243.
- 10. Di Maira, T.; Coelho, L.E.; Berenguer, M. Immunosuppression in liver transplant. Best Pract. Res. Clin. Gastroenterol. 2020, 46–47, 101681.
- 11. Sherston, S.N.; Carroll, R.P.; Harden, P.N.; Wood, K.J. Predictors of cancer risk in the long-term solid-organ transplant recipient. Transplantation 2014, 97, 605–611.
- Piselli, P.; Verdirosi, D.; Cimaglia, C.; Busnach, G.; Fratino, L.; Ettorre, G.M.; De Paoli, P.; Citterio, F.; Serraino, D. Epidemiology of de novo malignancies after solid-organ transplantation: Immunosuppression, infection and other risk factors. Best Pract. Res. Clin. Obstet. Gynaecol. 2014, 28, 1251–1265.

- Altieri, M.; Sérée, O.; Lobbedez, T.; Segol, P.; Abergel, A.; Blaizot, X.; Boillot, O.; Boudjema, K.; Coilly, A.; Conti, F.; et al. Risk factors of de novo malignancies after liver transplantation: A French national study on 11004 adult patients. Clin. Res. Hepatol. Gastroenterol. 2021, 45, 101514.
- Benlloch, S.; Berenguer, M.; Prieto, M.; Moreno, R.; San Juan, F.; Rayón, M.; Mir, J.; Segura, A.; Berenguer, J. De novo internal neoplasms after liver transplantation: Increased risk and aggressive behavior in recent years? Am. J. Transplant. 2004, 4, 596–604.
- Das, R.; Kundu, S.; Laskar, S.; Choudhury, Y.; Ghosh, S.K. Assessment of DNA repair susceptibility genes identified by whole-exome sequencing in head and neck cancer. DNA Repair. 2018, 66–67, 50–63.
- 16. Prendergast, G.C. Immune escape as a fundamental trait of cancer: Focus on IDO. Oncogene 2008, 27, 3889–3900.
- 17. Vajdic, C.M.; van Leeuwen, M.T. Cancer incidence and risk factors after solid organ transplantation. Int. J. Cancer 2009, 125, 1747–1754.
- 18. Lake, R.A.; van der Most, R.G. A better way for a cancer cell to die. N. Engl. J. Med. 2006, 354, 2503–2504.
- 19. Zitvogel, L.; Tesniere, A.; Kroemer, G. Cancer despite immunosurveillance: Immunoselection and immunosubversion. Nat. Rev. Immunol. 2006, 6, 715–727.
- 20. Dunn, G.P.; Koebel, C.M.; Schreiber, R.D. Interferons, immunity and cancer immunoediting. Nat. Rev. Immunol. 2006, 6, 836–848.
- 21. Glick, A.B.; Yuspa, S.H. Tissue homeostasis and the control of the neoplastic phenotype in epithelial cancers. Semin. Cancer Biol. 2005, 15, 75–83.
- Ferro, M.; Ungaro, P.; Cimmino, A.; Lucarelli, G.; Busetto, G.M.; Cantiello, F.; Damiano, R.; Terracciano, D. Epigenetic Signature: A New Player as Predictor of Clinically Significant Prostate Cancer (PCa) in Patients on Active Surveillance (AS). Int. J. Mol. Sci. 2017, 18, 1146.
- 23. Gutierrez-Dalmau, A.; Campistol, J.M. Immunosuppressive therapy and malignancy in organ transplant recipients: A systematic review. Drugs 2007, 67, 1167–1198.
- 24. Haller, M.C.; Royuela, A.; Nagler, E.V.; Pascual, J.; Webster, A.C. Steroid avoidance or withdrawal for kidney transplant recipients. Cochrane Database Syst. Rev. 2016, 8, CD005632.
- 25. Rutz, H.P.; Herr, I. Interference of glucocorticoids with apoptosis signaling and host-tumor interactions. Cancer Biol. Ther. 2004, 3, 715–718.
- 26. Cillo, U.; De Carlism, L.; Del Gaudio, M.; De Simone, P.; Fagiuoli, S.; Lupo, F.; Tisone, G.; Volpes, R. Immunosuppressive regimens for adult liver transplant recipients in real-life practice:
 Consensus recommendations from an Italian Working Group. Hepatol. Int. 2020, 14, 930–943.

- 27. Dougan, M.; Dranoff, G. Immune therapy for cancer. Annu. Rev. Immunol. 2009, 27, 83–117.
- Mortara, L.; Balza, E.; Bruno, A.; Poggi, A.; Orecchia, P.; Carnemolla, B. Anti-cancer Therapies Employing IL-2 Cytokine Tumor Targeting: Contribution of Innate, Adaptive and Immunosuppressive Cells in the Anti-tumor Efficacy. Front. Immunol. 2018, 9, 2905.
- 29. Angriman, I.; Furian, L.; Scarpa, M. Effects of immune suppression for transplantation on inflammatory colorectal cancer progression. Oncogenesis 2018, 7, 46.
- 30. Spolski, R.; Li, P.; Leonard, W.J. Biology and regulation of IL-2: From molecular mechanisms to human therapy. Nat. Rev. Immunol. 2018, 18, 648–659.
- 31. Burra, P.; Rodriguez-Castro, K.I. Neoplastic disease after liver transplantation: Focus on de novo neoplasms. World J. Gastroenterol. 2015, 21, 8753–8768.
- 32. Pillai, A.A.; Levitsky, J. Overview of immunosuppression in liver transplantation. World J. Gastroenterol. 2009, 15, 4225–4233.
- Hojo, M.; Morimoto, T.; Maluccio, M.; Asano, T.; Morimoto, K.; Lagman, M.; Shimbo, T.; Suthanthiran, M. Cyclosporine induces cancer progression by a cell-autonomous mechanism. Nature 1999, 397, 530–534.
- Yarosh, D.B.; Pena, A.V.; Nay, S.L.; Canning, M.T.; Brown, D.A. Calcineurin inhibitors decrease DNA repair and apoptosis in human keratinocytes following ultraviolet B irradiation. J. Investig. Dermatol. 2005, 125, 1020–1025.
- Guba, M.; von Breitenbuch, P.; Steinbauer, M.; Koehl, G.; Flegel, S.; Hornung, M.; Bruns, C.J.; Zuelke, C.; Farkas, S.; Anthuber, M.; et al. Rapamycin inhibits primary and metastatic tumor growth by antiangiogenesis: Involvement of vascular endothelial growth factor. Nat. Med. 2002, 8, 128–135.
- Maluccio, M.; Sharma, V.; Lagman, M.; Vyas, S.; Yang, H.; Li, B.; Suthanthiran, M. Tacrolimus enhances transforming growth factorbeta1 expression and promotes tumor progression. Transplantation 2003, 76, 597–602.
- 37. Verran, D.J.; Mulhearn, M.H.; Dilworth, P.J.; Balderson, G.A.; Munn, S.; Chen, J.W.; Fink, M.A.; Crawford, M.D.; McCaughan, G.W. Nature and outcomes of the increased incidence of colorectal malignancy after liver transplantation in Australasia. Med. J. Aust. 2013, 199, 610–612.
- Carenco, C.; Assenat, E.; Faure, S.; Duny, Y.; Danan, G.; Bismuth, M.; Herrero, A.; Jung, B.; Ursic-Bedoya, J.; Jaber, S.; et al. Tacrolimus and the risk of solid cancers after liver transplant: A dose effect relationship. Am. J. Transplant. 2015, 15, 678–686.
- 39. Allison, A.C.; Eugui, E.M. Mycophenolate mofetil and its mechanisms of action. Immunopharmacology 2000, 47, 85–118.

- 40. Cohn, R.G.; Mirkovich, A.; Caulfield, J.; Eugui, E.M. Apoptosis of human activated peripheral Tcells and Tlymphocytic and promonocytic cell lines induced by mycophenolic acid, the active metabolite of CellCept. In Proceedings of the Basic Sciences Symposium of the Transplantation Society, Monterey, CA, USA, 25–29 August 1999; p. 173.
- 41. Engl, T.; Makarević, J.; Relja, B.; Natsheh, I.; Müller, I.; Beecken, W.D.; Jonas, D.; Blaheta, R.A. Mycophenolate mofetil modulates adhesion receptors of the beta1 integrin family on tumor cells: Impact on tumor recurrence and malignancy. BMC Cancer 2005, 5, 4.
- 42. Francis, A.; Johnson, D.W.; Craig, J.; Teixeira-Pinto, A.; Wong, G. Post-transplant lymphoproliferative disease may be an adverse risk factor for patient survival but not graft loss in kidney transplant recipients. Kidney Int. 2018, 94, 809–817.
- 43. Koehl, G.E.; Wagner, F.; Stoeltzing, O.; Lang, S.A.; Steinbauer, M.; Schlitt, H.J.; Geissler, E.K. Mycophenolate mofetil inhibits tumor growth and angiogenesis in vitro but has variable antitumor effects in vivo, possibly related to bioavailability. Transplantation 2007, 83, 607–614.
- 44. De Simone, P.; Fagiuoli, S.; Cescon, M.; De Carlis, L.; Tisone, G.; Volpes, R.; Cillo, U.; Consensus Panel. Use of Everolimus in Liver Transplantation: Recommendations from a Working Group. Transplantation 2017, 101, 239–251.
- 45. Jacinto, E.; Hall, M.N. Tor signalling in bugs, brain and brawn. Nat. Rev. Mol. Cell Biol. 2003, 4, 117–126.
- Granata, S.; Dalla, G.A.; Carraro, A.; Brunelli, M.; Stallone, G.; Lupo, A.; Zaza, G. Sirolimus and Everolimus Pathway: Reviewing Candidate Genes Influencing Their Intracellular Effects. Int. J. Mol. Sci. 2016, 17, 735.
- 47. Laplante, M.; Sabatini, D.M. mTOR signaling in growth control and disease. Cell 2012, 149, 274–293.
- 48. Yang, H.; Rudge, D.G.; Koos, J.D.; Vaidialingam, B.; Yang, H.J.; Pavletich, N.P. mTOR kinase structure, mechanism and regulation. Nature 2013, 497, 217–223.
- 49. Hu, T.H.; Huang, C.C.; Lin, P.R.; Chang, H.W.; Ger, L.P.; Lin, Y.W.; Changchien, C.S.; Lee, C.M.; Tai, M.H. Expression and prognostic role of tumor suppressor gene PTEN/MMAC1/TEP1 in hepatocellular carcinoma. Cancer 2003, 97, 1929–1940.
- 50. Cholongitas, E.; Mamou, C.; Rodríguez-Castro, K.I.; Burra, P. Mammalian target of rapamycin inhibitors are associated with lower rates of hepatocellular carcinoma recurrence after liver transplantation: A systematic review. Transpl. Int. 2014, 27, 1039–1049.
- 51. Toso, C.; Merani, S.; Bigam, D.L.; Shapiro, A.M.; Kneteman, N.M. Sirolimus-based immunosuppression is associated with increased survival after liver transplantation for hepatocellular carcinoma. Hepatology 2010, 51, 1237–1243.

- 52. Treiber, G. mTOR inhibitors for hepatocellular cancer: A forward-moving target. Expert Rev. Anticancer Ther. 2009, 9, 247–261.
- Jiménez-Romero, C.; Manrique, A.; Marqués, E.; Calvo, J.; Sesma, A.G.; Cambra, F.; Abradelo, M.; Sterup, R.M.; Olivares, S.; Justo, I.; et al. Switching to sirolimus monotherapy for de novo tumors after liver transplantation. A preliminary experience. Hepatogastroenterology 2011, 58, 115–121.
- Funk-Debleds, P.; Ducroux, E.; Guillaud, O.; Ursic-Bedoya, J.; Decullier, E.; Vallin, M.; Euvrard, S.; Pageaux, G.P.; Boillot, O.; Dumortier, J. Subsequent nonmelanoma skin cancers and impact of immunosuppression in liver transplant recipients. J. Am. Acad. Dermatol. 2018, 79, 84–91.

Retrieved from https://encyclopedia.pub/entry/history/show/27469