

Lung Cancer Stem Cells

Subjects: Cell Biology

Contributor: Agata Raniszewska

CSCs are small numbers of cells that exist in the tumor microenvironment (TME). Lung cancer TME is composed of a various group of non-cancer cells, such as tumor-associated macrophages (TAMs stromal cells), regulatory T cells (Tregs), tumor-infiltrating lymphocytes (TILs), dendritic cells (DCs), natural killer (NK) cells, natural killer T (NKT), myeloid-derived suppressor cells (MDSCs), along with cancer cells: mature cancer cells and CSCs. As yet, the complexity of the interactions between the cells in the immune TME has not been exhaustively described.

Keywords: non-small lung cancer ; cancer stem cells ; liquid biopsy ; immunotherapy ; aging ; biopsy

1. Introduction

Lung cancer is still a leading cause of cancer-related deaths globally among men (23% of all cancer-related deaths) and women (22% of all cancer-related deaths) ^[1]. Worldwide, cigarette smoking alone accounts for over 80% of all lung cancer cases ^[2]. Other factors, such as air pollution, emission fuel combustion indoor, environmental exposure to radon and asbestos, contribute to the development of lung cancer ^[3]. As all these risk factors can be prevented through smoking cessation and air purification activities, it is possible to diminish lung cancer incidence and mortality through population-based preventive strategies ^[2].

Lung cancer is categorized into two main histological types: small cell lung carcinoma (SCLC, 15% of all lung cancers) and non-small cell lung carcinoma (NSCLC, 85% of all lung cancers). NSCLC comprises main histological subtypes: adenocarcinoma (ADC), squamous cell carcinoma (SQCLC) and large cell carcinoma (LCC) ^[4]. The collected data suggest that lung cancer is a group of histologically and molecularly heterogeneous disorders even within the same histologic subtype ^[5].

Most NSCLC patients are diagnosed at advanced stage, when the various treatments cannot be curative. Thus, this requires greater readiness of primary care physicians to carefully screen patients at high risk, even with non-specific symptoms ^[2]. In the course of the disease, lung cancer spreads when cells detach from a tumor and pass through the bloodstream or the lymph vessels to distant areas of the body and grow. The most common sites of the spread of lung cancer are the: lymph nodes, liver, bones, brain, adrenal glands ^[6].

2. Lung Cancer and Aging

Metastatic solid tumors, such as lung cancer, remain largely incurable despite improvements in cancer therapies. Aging is still the most important risk factor for lung cancer development. NSCLC is a heterogeneous illness with unique combinations of somatic molecular changes in individual patients, as well as significant differences in populations around the world with respect to mutation spectra and frequencies. Interestingly, it has been proven that aging is tightly associated with developing EGFR mutation in lung cancer ^{[7][8]}.

Alterations in these pathways have been described in different chronic lung diseases, including lung cancer ^{[9][10][11]}. Participation of stem cells in this process is highly probable. Depending on the nature and damage size, cytostatic, cytotoxic, or mutagenic lesions arising in stem cells have the potential to lead to cells senescence, apoptosis, or transformation (**Figure 1**). What is more, normal stem cells can become cancer stem cells (CSCs) through the acquisition of mutation and genetic or epigenetic alterations ^[12].

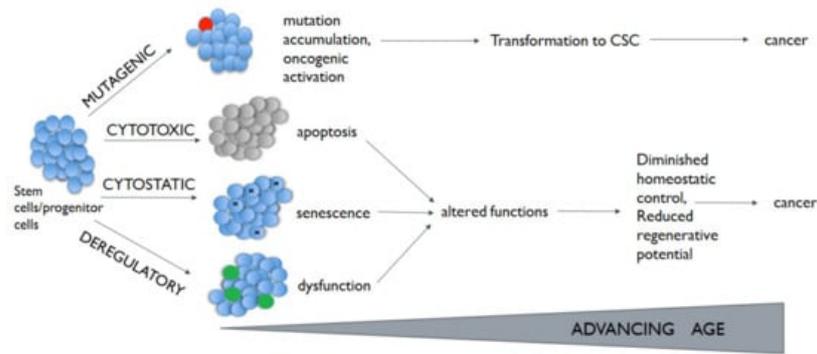


Figure 1. Age-related DNA damage in stem cells. Mutation accumulation and/or oncogenic activation leads to transformation to cancer stem cells (CSCs; marked red) and the initiation of cancer. Cytotoxic and cytostatic effects lead to stem cell senescence or apoptosis. It may result in reduction in the stem cell population. Deregulatory mechanisms lead to reduced homeostatic control and diminished regenerative potential.

Some author suggests that driver mutations increase the cancer cell stemness, even if it is not possible to be said unequivocally that a cell with a higher number of driver mutations corresponds to less uninvolved phenotype of the cells as in the case of normal stem cells [13][14]. There are some data confirming the presence of driver mutations in CSCs (e.g., KRAS in colorectal CSCs [15]). It should be pointed out that CSCs are a diverse population of cells with relatively high plasticity and a reversible phenotype. However, an insufficient number of pre-clinical and clinical studies on CSCs in NSCLC patients make it impossible to evaluate whether the new targeted therapy may reduce the number of lung CSCs.

3. Lung Cancer Stem Cells

Nevertheless, the identification of stem cell origin in the lungs presents a challenge, because the tracheal and bronchiolar epithelia are quiescent, having a low proliferative fraction [16]. Thus, in some simplification, the origin of lung CSCs has been traced back to cells on specific anatomical sites on lungs (Figure 2). The basal cells of proximal airway, such as trachea and bronchi, are linked with SQCLC showing stem-cell-like behavior [16]. ADC is linked with normal stem cells from the bronchoalveolar duct junction area [17].

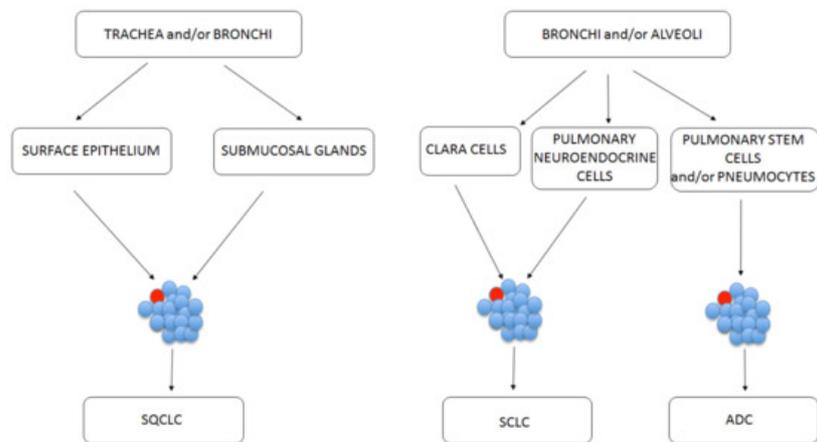


Figure 2. Possible CSC (marked red) initiation sites depending on the lung cancer subtype.

Although the present knowledge on lung CSCs functions is restricted, a number of CSC markers have been proposed. These are often markers belonging to CD (cluster of differentiation). Many studies have confirmed the presence of the following molecules on lung CSCs: CD133, CD44, CD90, epithelial cell adhesion molecule (EpCAM), C-X-C chemokine receptor type 4 (CXCR4)

We identified lung CSCs exhibiting CD133, CD44, CD90, EpCAM, CXCR4 in lung cancer patients in our previous studies [18][19][20]. The rationale for the identification of CSCs according to some authors is that they seem to be more stable, guarantee resistance to systemic therapies and, in our view, are capable of bringing information for the modification of immune response in the site of the tumor [16][21].

Patient-derived organoids can be applicable to identify treatment resistance signatures of cancer stem cells in treated organoids. Unfortunately, the majority of cancer organoid models are limited to adenocarcinomas [22]. However, as we broaden our understanding of tumor development and more, cancer stem cell markers are identified, organoids can only

become a stronger research tool covering all types of cancer [22]. Thus, combined with other in vivo experiments, such as xenotransplantation assays of CSCs, organoid cultures of human CSCs have a high potential to advance our understanding of human cancer biology [23].

It is thought to be related to their activation of different signaling pathways such as: Wnt (Wingless-type), Notch, and Hedgehog [24]. A growing body of evidence supports the association of Notch signaling dysregulation with various types of malignancies, including NSCLC [25]. Notch signaling pathway play a role in stem cell maintenance in NSCLC; aberration in that pathway may result in increasing the number of CSCs resistant to platinum-based therapy [26]. What is more, Hedgehog pathway is involved in tumor drug resistance in NSCLC, as cytotoxic chemotherapy, targeted therapies and radiotherapy [27].

4. Cancer Stem Cells and Tumor Microenvironment

Lung cancer TME is composed of a various group of non-cancer cells, such as tumor-associated macrophages (TAMs stromal cells), regulatory T cells (Tregs), tumor-infiltrating lymphocytes (TILs), dendritic cells (DCs), natural killer (NK) cells, natural killer T (NKT), myeloid-derived suppressor cells (MDSCs), along with cancer cells: mature cancer cells and CSCs [28][29]. Indeed, each cell involved, immune cell or tumor cell may influence the immunological behavior of the other cells, either distant or adjacent, within the TME. Some studies showed also that CSCs may activate mechanisms to circumvent a possible attack from the immune cells: loss of cancer antigen expression, initiation of oncolytic pathways, and promotion of immunosuppressive milieu [30]. It provides the ground to induce or recruit the differentiation of suppressive immune cells, including suppressive macrophages (M2 type) or Tregs [31][32].

MDSCs represent the heterogeneous group of immature myeloid cells (precursors of macrophages, dendritic cells and granulocytes). MDSCs show pro-angiogenic activity, induce the production of metalloproteinases and contribute to the formation of pre-metastatic niches that facilitate the colonization of cancer cells [33][34]. Reports from recent years indicate that MDSCs affect the expression of oncogenes in CSCs and induce their proliferation [35][33]. Furthermore, MDSCs can regulate CSCs by the secretion of pro-inflammatory cytokines: IL-1,

TME elicits differentiation of the CD4+ T cells into different subsets of T cells, particularly suppressive Tregs, and T helper 17 (Th17) cells. Despite this, the exact role of the last cells in tumor immunity is still unclear depending on tumor stages and histological subtypes. Intriguingly, recent reports demonstrate that Tregs, under certain circumstances, express IL-17, which together with hypoxia plays a pivotal role in the regulation of CSCs [36]. Nonetheless, the interactions between CSCs and Tregs, which play an important immunosuppressive role in the TME, are still poorly understood.

The understanding of the immunological profile of CSCs and their interaction within TME has provoked the investigation of the immunological targeting of these cells (**Table 1**).

Table 1. Recent advances in targeting CSCs by immunotherapy. DCs—dendritic cells; CAR T cells—chimeric antigen receptor T cells; ID8-T—epithelial ovarian cancer cell line.

Type of Immunotherapy	Condition	Study Intervention	Reference
DCs vaccination	SQCLC, melanoma	ALDH ^{high} CSC-pulsed DCs	[37]
DCs vaccination	Squamous cell cancer, melanoma	CSCs lysate-pulsed DCs	[38][39]
T-cell therapy	Colon cancer	CD8+ cytotoxic T-cells, specific for the CSCs antigen	[40]
T-cell therapy	Prostate cancer	CAR T-cells against EpCAM antigen	[41]
Virotherapy	Glioblastoma	Oncolytic adenovirus targeting CD133+ CSCs	[42]
Virotherapy	Ovarian cancer	Oncolytic vaccinia virus targeting ID8-T tumor model that harbors CSCs	[43]
Virotherapy	Hepatocellular carcinom	Oncolytic measles viruses: targeting CD133+ CSCs	[44]
Virotherapy	Breast cancer	Oncolytic vaccinia virus targeting ALDH ^{high} CSCs	[45]
Combined therapy	Bladder cancer	CSCs vaccine combined with anti-PD-1	[37]
Monoclonal antibody	Breast cancer	Anti-CD44 antibody	[46]
CSC-CAR T	Prostate	EpCAM-specific CAR T cell	[37]

Type of Immunotherapy	Condition	Study Intervention	Reference
Targeting signaling pathway	Lung cancer	Hedgehog pathway inhibitor	[47]
CSC-primed T cells	Lung cancer	CD8+ cytotoxic T-cells, ALDH ^{high} specific CSCs	[48]

5. Conclusions

Understanding the biology of cancer stem cells (progenitor cells, CSCs) is one of the greatest challenges in basic science and clinical oncology. The presented directions and research results show that these cells are significantly associated with the development of solid tumors, such as lung cancer. ADC, which is recently more and more frequent and is now precisely recognized, also in terms of molecular changes, needs special attention. Identification of CSCs in this type of cancer with the use of markers discussed above may contribute to the designation of new therapy directions. Investigation of somatic mutation in normal tissues and its role in tumor progression and aging will provide a new insight into cancer treatment. Direct studies of mutation load, mutation signatures, clonal dynamics, and cellular phenotypes will provide a bridge from epidemiological discoveries to mechanistic insights into the earliest stages of cancer. A liquid biopsy may improve the qualification of lung cancer patients to targeted therapies or immunotherapies, through the identification of appropriate tumor-specific targets and biomarkers and to better define the predictors of the response to modern therapies.

References

1. Siegel, R.L.; Miller, K.D.; Jemal, A. Cancer statistics, 2020. *CA Cancer J Clin.* 2020, 70, 7–30.
2. Wong, M.C.S.; Lao, X.Q.; Ho, K.F.; Goggins, W.B.; Tse, S. Incidence and mortality of lung cancer: Global trends and association with socioeconomic status. *Sci. Rep.* 2017, 7, 14300.
3. Raaschou-Nielsen, O.; Andersen, Z.J.; Beelen, R.; Samoli, E.; Stafoggia, M.; Wenmayr, G.; Hoffmann, B.; Fischer, P.; Nieuwenhuijsen, M.J.; Brunekreef, B.; et al. Air pollution and lung cancer incidence in 17 European cohorts: Prospective analyses from the European Study of cohorts for air pollution effects [ESCAPE]. *Lancet Oncol.* 2013, 14, 813–822.
4. Travis, W.D.; Brambilla, E.; Nicholson, A.G.; Yatabe, Y.; Austin, J.H.M.; Beasley, M.B.; Chirieac, L.R.; Dacic, S.; Duhig, E.; Flieder, D.B.; et al. WHO panel. The 2015 world health organization classification of lung tumors: Impact of genetic, clinical and radiologic advances since the 2004 classification. *J. Thorac. Oncol.* 2015, 10, 1243–1260.
5. Inamura, K. Lung cancer: Understanding its molecular pathology and the 2015 WHO classification. *Front. Oncol.* 2017, 7, 193.
6. Popper, H.H. Progression and metastasis of lung cancer. *Cancer Metastasis Rev.* 2016, 35, 75–91.
7. Choi, W.I.; Jeong, J.; Lee, C.W. Association between EGFR mutation and ageing, history of pneumonia and gastroesophageal reflux disease among patients with advanced lung cancer. *Eur. J. Cancer* 2019, 122, 101–108.
8. Choi, Y.H.; Lee, J.K.; Kang, H.J.; Lee, T.S.; Kim, H.R.; Kim, C.H.; Koh, J.S.; Baek, H.J.; Lee, J.C.; Na, I.I. Association between age at diagnosis and the presence of EGFR mutations in female patients with resected non-small cell lung cancer. *J. Thorac. Oncol.* 2010, 12, 1949–1952.
9. Hoffmeyer, K.; Raggioli, A.; Rudloff, S.; Anton, R.; Hierholzer, A.; Del Valle, I.; Hein, K.; Vogt, R.; Kemler, R. Wnt/ β -catenin signaling regulates telomerase in stem cells and cancer cells. *Science* 2012, 336, 1549–2554.
10. Yu, B.; Chang, J.; Liu, Y.; Li, J.; Kevork, K.; Al-Hezaimi, K.; Graves, D.T.; Park, N.H.; Wang, C.Y. Wnt4 signaling prevents skeletal aging and inflammation by inhibiting nuclear factor- κ B. *Nat. Med.* 2014, 20, 1009–1017, Erratum in: 2015, 21, 1101.
11. Castilho, R.M.; Squarize, C.H.; Chodosh, L.A.; Williams, B.O.; Gutkind, J.S. mTOR mediates Wnt-induced epidermal stem cell exhaustion and aging. *Cell Stem Cell.* 2009, 5, 279–289.
12. Santos Franco, S.; Raveh-Amit, H.; Kobilák, J.; Alqahtani, M.H.; Mobasher, A.; Dinnyes, A. The crossroads between cancer stem cells and aging. *BMC Cancer* 2015, 15 (Suppl. S1).
13. Lee, G.; Hall, R.R., III; Ahmed, A.U. Cancer stem cells: Cellular plasticity, niche, and its clinical relevance. *J. Stem. Cell Res. Ther.* 2016, 6, 363.
14. Ascolani, G.; Liò, P. Modeling breast cancer progression to bone: How driver mutation order and metabolism matter. *BMC Med. Genom.* 2019, 12 (Suppl. S6).

15. Hwang, J.H.; Yoon, J.; Cho, Y.H.; Cha, P.H.; Park, J.C.; Choi, K.Y. A mutant KRAS-induced factor REG4 promotes cancer stem cell properties via Wnt/ β -catenin signaling. *Int. J. Cancer* 2020, 146, 2877–2890.
16. Testa, U.; Castelli, G.; Pelosi, E. Lung cancers: Molecular characterization, clonal heterogeneity and evolution, and cancer stem cells. *Cancers* 2018, 10, 248.
17. Barr, M.P.; Gray, S.G.; Hoffmann, A.C.; Hilger, R.A.; Thomale, J.; O'Flaherty, J.D.; Fennell, D.A.; Richard, D.; O'Leary, J.J.; O'Byrne, K.J. Generation and characterisation of cisplatin-resistant non-small cell lung cancer cell lines displaying a stem-like signature. *PLoS ONE* 2013, 8, e54193, Erratum in: 2020, 21, e0233739.
18. Raniszewska, A.; Polubiec-Kownacka, M.; Rutkowska, E.; Domagała-Kulawik, J. PD-L1 expression on lung cancer stem cells in metastatic lymph nodes aspirates. *Stem Cell Rev.* 2019, 15, 324–330.
19. Raniszewska, A.; Vroman, H.; Dumoulin, D.; Cornelissen, R.; Aerts, J.G.J.V.; Domagała-Kulawik, J. PD-L1+ lung cancer stem cells modify the metastatic lymph-node immunomicroenvironment in nsclc patients. *Cancer Immunol. Immunother.* 2021, 70, 453–461.
20. Raniszewska, A.; Kwiecień, I.; Sokołowski, R.; Rutkowska, E.; Domagała-Kulawik, J. Immunomodulatory molecules on lung cancer stem cells from lymph nodes aspirates. *Cancers* 2020, 12, 838.
21. De Francesco, E.M.; Sotgia, F.; Lisanti, M.P. Cancer stem cells (CSCs): Metabolic strategies for their identification and eradication. *Biochem. J.* 2018, 475, 1611–1634.
22. Nagle, P.W.; Plukker, J.T.M.; Muijs, C.T.; van Luijk, P.; Coppes, R.P. Patient-derived tumor organoids for prediction of cancer treatment response. *Semin. Cancer Biol.* 2018, 53, 258–264.
23. Shimono, Y.; Mukohyama, J.; Isobe, T.; Johnston, D.M.; Dalerba, P.; Suzuki, A. Organoid culture of human cancer stem cells. In *Organoids, Methods in Molecular Biology*; Turksen, K., Ed.; Humana: New York, NY, USA, 2016; Volume 1576.
24. Morrison, R.; Schleicher, S.M.; Sun, Y.; Niermann, K.J.; Kim, S.; Spratt, D.E. Targeting the mechanisms of resistance to chemotherapy and radiotherapy with the cancer stem cell hypothesis. *J. Oncol.* 2011, 2011.
25. Galluzzo, P.; Bocchetta, M. Notch signaling in lung cancer. *Exp. Rev. Anticancer Ther.* 2011, 11, 533–540.
26. Zhang, Y.; Xu, W.; Guo, H.; Zhang, Y.; He, Y.; Lee, S.H.; Song, X.; Li, X.; Guo, Y.; Zhao, Y.; et al. NOTCH1 signaling regulates self-renewal and platinum chemoresistance of cancer stem-like cells in human non-small cell lung cancer. *Cancer Res.* 2017, 77, 3082–3091.
27. Cochrane, C.R.; Szczepny, A.; Watkins, D.N.; Cain, J.E. Hedgehog signaling in the maintenance of cancer stem cells. *Cancers* 2015, 7, 1554–1585.
28. Schreiber, R.D.; Old, L.J.; Smyth, M.J. Cancer immunoediting: Integrating immunity's roles in cancer suppression and promotion. *Science* 2011, 331, 1565–1570.
29. Ogino, S.; Galon, J.; Fuchs, C.S.; Dranoff, G. Cancer immunology--analysis of host and tumor factors for personalized medicine. *Nat. Rev. Clin. Oncol.* 2011, 8, 711–719.
30. Zhang, D.G.; Tang, K.; Rycaj, K. Cancer stem cells: Regulation programs, immunological properties and immunotherapy. *Semin. Cancer Biol.* 2018, 52, 94–106.
31. Maccalli, C.; Volonte, A.; Cimminiello, C.; Parmiani, G. Immunology of cancer stem cells in solid tumours. A review. *Eur. J. Cancer* 2014, 50, 649–655.
32. Quian, B.Z.; Pollard, J.W. Macrophage diversity enhances tumor progression and metastasis. *Cell* 2010, 141, 39–51.
33. Law, A.M.K.; Valdes-Mora, F.; Gallego-Ortega, D. Myeloid-derived suppressor cells as a therapeutic target for cancer. *Cells* 2020, 9, 561.
34. Groth, C.; Hu, X.; Weber, R.; Fleming, V.; Altevogt, P.; Utikal, J.; Umansky, V. Immunosuppression mediated by myeloid-derived suppressor cells (MDSCs) during tumour progression. *Br. J. Cancer* 2019, 120, 16–25.
35. Miranda-Lorenzo, I.; Dorado, J.; Lonardo, E.; Alcala, S.; Serrano, A.G.; Clausell-Tormos, J.; Cioffi, M.; Megias, D.; Zago, S.; Balic, A.; et al. Intracellular autofluorescence: A biomarker for epithelial cancer stem cells. *Nat. Methods* 2014, 11, 1161–1169.
36. Rezaiofii, A.; Ahmadian, E.; Aazami, H.; Solgi, G.; Ebrahimi, M. Gastric cancer stem cells effect on Th17/treg balance; A bench to bedside perspective. *Front. Oncol.* 2019, 9, 226.
37. Deng, Z.; Wu, Y.; Ma, W.; Zhang, S.; Zhang, Y.Q. Adoptive t-cell therapy of prostate cancer targeting the cancer stem cell antigen epcam. *BMC Immunol.* 2015, 16.
38. Dashti, A.; Ebrahimi, M.; Hadjati, J.; Memarnejadian, A.; Moazzeni, S.M. Dendritic cell based immunotherapy using tumor stem cells mediates potent antitumor immune responses. *Cancer Lett.* 2016, 374, 175–185.

39. Lu, L.; Tao, H.; Chang, A.E.; Hu, Y.; Shu, G.; Chen, Q.; Egenti, M.; Owen, J.; Moyer, J.S.; Prince, M.E.; et al. Cancer stem cell vaccine inhibits metastases of primary tumors and induces humoral immune responses against cancer stem cells. *Oncoimmunology* 2015, 4, e990767.
40. Miyamoto, S.; Kochin, V.; Kanaseki, T.; Hongo, A.; Tokita, S.; Kikuchi, Y.; Takaya, A.; Hirohashi, Y.; Tsukahara, T.; Terui, T.; et al. The antigen asb4 on cancer stem cells serves as a target for ctl immunotherapy of colorectal cancer. *Cancer Immunol. Res.* 2018, 6, 358–369.
41. Sato-Dahlman, M.; Miura, Y.; Huang, J.L.; Hajeri, P.; Jacobsen, K.; Davydova, J.; Yamamoto, M. Cd133-targeted oncolytic adenovirus demonstrates anti-tumor effect in colorectal cancer. *Oncotarget* 2017, 8, 76044–76056.
42. Gil, M.; Komorowski, M.P.; Seshadri, M.; Rokita, H.; McGray, A.J.; Opyrchal, M.; Odunsi, K.O.; Kozbor, D. Cxcl12/cxcr4 blockade by oncolytic virotherapy inhibits ovarian cancer growth by decreasing immunosuppression and targeting cancer-initiating cells. *J. Immunol.* 2014, 193, 5327–5337.
43. Bach, P.; Abel, T.; Hoffmann, C.; Gal, Z.; Braun, G.; Voelker, I.; Ball, C.R.; Johnston, I.C.; Lauer, U.M.; Herold-Mende, C.; et al. Specific elimination of cd133+ tumor cells with targeted oncolytic measles virus. *Cancer Res.* 2013, 73, 865–874.
44. Wang, H.; Chen, N.G.; Minev, B.R.; Szalay, A.A. Oncolytic vaccinia virus glv-1h68 strain shows enhanced replication in human breast cancer stem-like cells in comparison to breast cancer cells. *J. Transl. Med.* 2012, 10, 167.
45. Shi, X.; Zhang, X.; Li, J.; Mo, L.; Zhao, H.; Zhu, Y.; Hu, Z.; Gao, J.; Tan, W. Pd-1 blockade enhances the antitumor efficacy of gm-csf surface-modified bladder cancer stem cells vaccine. *Int. J. Cancer* 2018, 142, 2106–2117.
46. Aires, A.; Ocampo, S.M.; Simoes, B.M.; Josefa Rodriguez, M.; Cadenas, J.F.; Couleaud, P.; Spence, K.; Latorre, A.; Miranda, R.; Somoza, A.; et al. Multifunctionalized iron oxide nanoparticles for selective drug delivery to CD44-positive cancer cells. *Nanotechnology* 2016, 27, 65103.
47. Tian, F.; Mysliwicz, J.; Ellwart, J.; Gamarra, F.; Huber, R.M.; Bergner, A. Effects of the Hedgehog pathway inhibitor GD C-0449 on lung cancer cell lines are mediated by side populations. *Clin. Exp. Med.* 2012, 12, 25–30.
48. Luo, H.; Zeng, C.; Fang, C.; Seeruttun, S.R.; Lv, L.; Wang, W. A new strategy using ALDHhigh-CD8+T cells to inhibit tumorigenesis. *PLoS ONE* 2014, 9, e103193.

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