

Hibernation

Subjects: **Biophysics**

Contributor: Anggraeini Puspitasari

Torpor and hibernation are natural physiological processes. Torpor refers to a period of metabolic suppression with a duration from a few hours to several weeks. The state of torpor is probably older in evolutionary terms and was likely a survival strategy of protomammals. Hibernation is a more elaborate behavior, structured in many long bouts of torpor separated by brief interbouts of arousal.

hibernation

torpor

space

radiation protection

1. Hibernation and Survival in Extreme Environments

In space, the quantity and quality of radiation are not the same as on Earth. Charged particles from the Sun and distant galaxies hit astronauts continuously. However, there are animals on Earth that can live in adverse conditions and environments, even with high radiation doses. The way that these animals manage to survive in these conditions could help us understand the molecular mechanisms behind these remarkable peculiarities ^{[1][2][3]}. Among them, species of interest that can survive radiation are hibernators.

During their inactive state, hibernators become more radioresistant ^{[1][2][3]}. Hibernation is a biological condition in which vital functions are minimized, the heartbeat decreases, metabolism is reduced, and body temperature is lowered. The drop range of the temperature depends on the size of the animal. For example, in bears, the body temperature during hibernation is lowered by a few degrees, while the arctic ground squirrel's goes down to almost 0 °C during its inactive state. Hibernation is not a constant and immutable physiological process. During the process, the animal goes through a series of arousals in which its body temperature returns to normothermic values rapidly and for about 24 h. Technically, bouts of hypometabolism are called torpor, while hibernation is a sequence of torpor bouts separated by brief interbout arousals. Hibernation can last from a few hours to many months.

Like every other mammal, humans are homeothermic (warm-blooded) animals and maintain a constant body temperature. Warm-blooded animals do not depend on the environment to regulate their body temperature, as do ectotherms. A higher body temperature improves brain and muscle function and allows warm-blooded animals to be active every moment of the day. The downside is that in order to maintain a high and stable body temperature, an extensive and constant intake of energy throughout the year is required; therefore, large available food supplies are needed. This makes homeothermic animals vulnerable to extreme environmental changes. For example, humans cannot survive more than 8–21 days without food ^[4]. However, this is not the case with hibernators.

Hibernators in their active state also have a high metabolism, keeping their body temperature constant, but they can survive for many months without food and water in their inactive state.

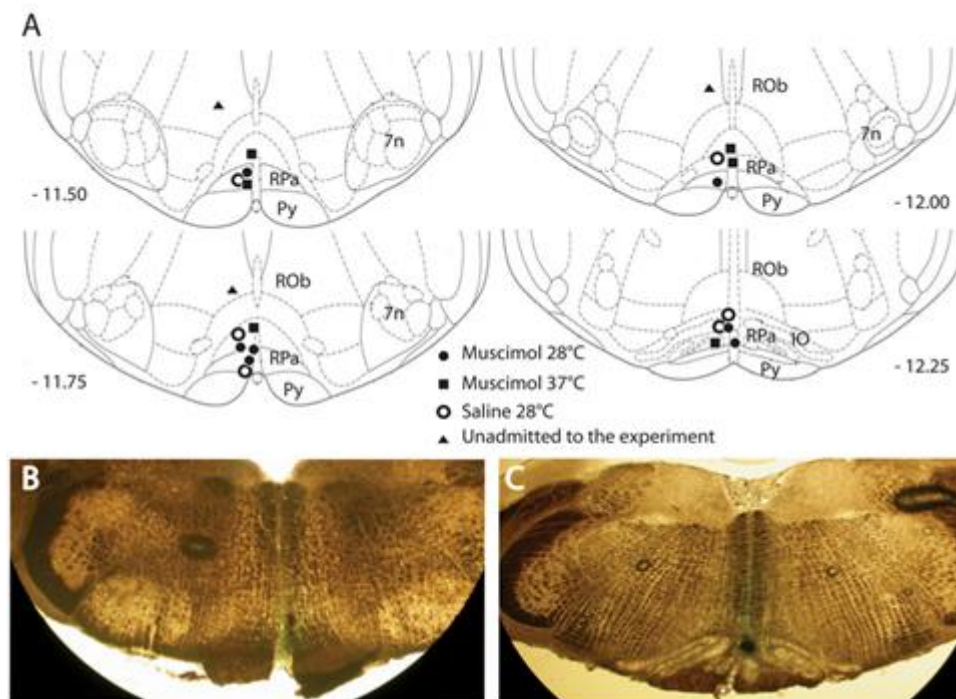


Figure 2. Distribution and locations of microinjections of GABA-A agonist muscimol in the brainstem. A key area in the central nervous pathways for thermoregulatory cold defense is the rostral ventromedial medulla (RVMM), a region including the raphe pallidus (RPa). (A) The location of every injection site, marked with fast green after each experimental procedure, was schematically plotted on atlas drawings [5] at four rostrocaudal levels of the RVMM. (B,C) Examples of marked sites at two rostrocaudal levels: 7n = nucleus of cranial nerve VII; IO = inferior olive; Py = pyramid; and Rob = raphe obscurus. This was adapted from [19]. Copyright 2013, Society for Neuroscience.

2. Genomic Instability and DNA Repair of a Hibernator

The human metabolism is finely organized and coordinated because of its complex machinery. It becomes a challenge when we have to adjust to or survive in extreme conditions. Astronauts require a high level of physical fitness to perform their mission. The need to be well trained and in good health compared with other occupations is fundamental. However, they still cannot escape from the risks of radiation-induced carcinogenesis. Radiation carcinogenesis is a slow process. Normal living cells damaged by ionizing radiation start a progressive genotypic change, in turn causing a drastic change in their phenotype. Regular cell cycle control points, cell contact inhibition, and apoptosis-programmed death are lost, and cells become malignant. An epidemiological study showed that leukemia has been linked with external radiation exposure in Japanese atomic bomb survivors and medically exposed persons, and skin cancer is linked with radiation exposure in radiologists [6]. Even though astronauts are exposed to much lower doses, there are still not many studies defining the risk of carcinogenesis. Other epidemiological studies show that cancer initiation processes dominate radiation risk after exposure in young

people, and radiation could promote preexisting malignant cells after exposure at later ages [6]. This means that if astronauts have an unknown preexisting condition, space flight could promote cancer later in life.

The mechanisms by which radiation can produce carcinogenic changes are postulated as the induction of (1) mutations, including alterations in the structure of single genes or chromosomes; (2) changes in gene expression without mutations; and (3) oncogenic viruses, which in turn can cause neoplasia [7]. Cytogenetic analysis of the lymphocytes of astronauts provides a direct measurement of space radiation damage. Chromosome exchanges were measured in the blood lymphocytes of eight crew members after their respective space missions using fluorescence in situ hybridization. The analysis showed significant increases in chromosome aberrations. The presence of cytogenetic damage was observed after long-duration and repeated missions [8][9]. Thus, evidence shows that space travelers may have genomic instability or the mutation of cells, which may lead to carcinogenesis.

Hibernation might be able to mitigate the radiation-induced genomic instability. There is an interesting study on hibernators, such as arctic ground squirrels, that are able to avoid genome instability during torpor–arousal cycles through status-specific combinations of strategies for preventing DNA damage and promoting efficient DNA repair, paired with anti-apoptotic environments. The hypothalamus, as the center of thermoregulation, plays an important role in hibernation initiation, and the defense mechanism of the hypothalamus of the ground squirrel is of interest and might be the key to the DNA repair mechanism function. These adaptations include upregulated genomic protective measures, specifically proteins involved in the detection and response to double-strand breaks (e.g., RAD50, NBN, and ATM) [10], which are important for ATM activation by DNA damage [11]. Interestingly, when the synthetic torpor rats were acutely exposed to 3 Gy of X-rays, ATM-related genes were downregulated in the testis and the liver [12]. ATM primarily initiates cellular responses to radiation-induced double-strand breaks [13], and this finding is crucial for the response to radiation-induced DNA damage. Thus, studies showed that the expression of DNA damage-related genes might be different depending on the species, stressor, and organs. Considering the occupational hazard of astronauts, radiation-induced damage is still unavoidable. Therefore, understanding the mechanism of how hibernators can adapt and repair damage efficiently will be useful for astronauts.

3. Potential of Hibernation to Protect Higher Brain Function from Radiation Effects

Several studies on rodents have shown that radiation, including cosmic rays, can damage synaptic integrity and induce neuroinflammation [14][15][16]. Inflammation persists for more than six months after exposure [16]. The molecular changes in synapses have been shown to affect neuronal function, resulting in behavioral changes. Furthermore, the effects of radiation on a smaller scale, such as DNA damage, can lead to synaptic dysfunction and neurodegeneration. The study of gene expression in the hypothalamus of arctic ground squirrels showed that during hibernation, they have strategies to prevent DNA damage by performing efficient DNA repair [32]. Neurons, as non-cycling cells, are generally known to have very high radioresistance, since most of the irradiated cells die due to mitotic catastrophe [17]. However, that is not entirely true, since a neuron's sensitivity to radiation depends on its developmental stage. Studies using 7 days in vitro (DIV) of immature primary hippocampal neurons and 21

DIV of mature neurons showed that mature neurons are more resistant 24 h after exposure to 50 Gy of X-rays in terms of cell death [18]. However, 30 Gy of X-rays has been shown to affect the morphology of cells [19]. In contrast, immature neurons are relatively sensitive to radiation, and they go into apoptosis after being exposed to ionizing radiation [18][20]. A long-term study of immature neurons showed delayed cell death, a change in the dendritic morphology, and critical synaptic proteins PSD-95 and drebrin three weeks after 0.5 Gy and 1 Gy of X-rays [21]. Cell death of immature neurons is not the only underlying cause of cognitive impairment. In studies using 10 Gy of X-rays, in the acute phase, transient synaptic dysfunction occurred, leading to temporary cognitive impairment, which occurred only within 24 h [22][23]. Although those studies used much higher doses than the dose received in space, the results showed that the effects of radiation on synaptic function also need to be investigated, and hibernation might be able to mitigate radiation-induced synaptic dysfunction.

Brain activity was reported to change dramatically during hibernation. The electroencephalograms (EEGs) were nearly constant in hibernating ground squirrels and hamsters at their lowest body temperatures [24][25]. In spontaneous neuronal activity, the firing rates were systematically reduced with decreasing body temperatures. Neurons stopped firing at a body temperature of 15–18 °C, remained silent for 10–28 h (deep torpor), and only began firing again when the body temperature increased [26][27]. Furthermore, in addition to the findings on the upregulation of DNA damage-related genes, in the hypothalamus of ground squirrels, the cerebral cortex showed remodeling and plasticity during hibernation, along with evidence of synapse functional organization, which was not seen in the hypothalamus [10]. These data imply that neuronal activity in hibernating animals is highly dependent on body and brain temperature, and different processes take place in different parts of the brain during hibernation. The dynamic change of neuronal activity is related to synaptic plasticity. An actin-binding protein, drebrin, plays a crucial role in synaptic plasticity [28]. A radiation-induced decrease of drebrin was prevented by the administration of an N-methyl-D-aspartic acid (NMDA) receptor antagonist, MK-801, before radiation [23]. Although the dose in the study used was 10 Gy of X-rays, this shows that NMDA receptor-induced toxicity is one of the underlying causes of radiation-induced synaptic dysfunction [29]. Referring to a study using hippocampal slices of hibernators, radiation-induced NMDA toxicity might be avoided during hibernation. A study on NMDA in hippocampal slices of hibernating ground squirrel neurons after 24 h in culture showed higher resistance than euthermic or non-hibernating animals. It also showed that inhibition of the Na⁺/K⁺ pump did not lead to increased cell death in the hippocampal slices [30], which means that the hibernation process may protect the hippocampus from radiation-induced neuronal cell death.

4. Cardiovascular Function During Hibernation

Astronauts in space are subjected to microgravity, which causes muscle atrophy and cardiovascular problems. Cardiovascular issues experienced by most astronauts emerge after space flight [31]. Although there are no changes in baroreflexes or cardiac function during flights, studies showed that six months after returning to Earth, astronauts could experience a slightly increased heart rate. Moreover, it seems that even a short-duration flight—around 10 days—can result in a marked loss of cardiac muscle mass [32]. The loss of cardiac muscle might be in response to a decreased physiological load, which in turn underlies the decrease or loss of plasma volume during

spaceflight [33]. This phenomenon might be preventable if we could control the cardiac output and heart rate efficiently.

A study of grizzly bears (*Ursus arctos horribilis*) and American black bears (*U. americanus*) suggested substantial cardiac adaptations during hibernation, characterized by a marked decrease in cardiac output caused by profound bradycardia. Furthermore, the bears presented severe respiratory sinus arrhythmia and a preserved left ventricular ejection fraction. The measurement of grizzly bears showed that myocardial contractility was significantly lower in all bears during hibernation than during the active period [34]. There are dramatic changes in physiological and molecular parameters during winter hibernation in some hibernators, like the ground squirrels (*Ictidomys tridecemlineatus*). Different studies have demonstrated reductions in phosphorylated Bcl-2 antagonist of cell death (p-BAD)-mediated pro-apoptotic signaling during hibernation, with active caspase-9 protein levels increasing only during the interbout arousal. *I. tridecemlineatus* has natural tissue protection mechanisms during hibernation, mainly due to cellular regulation through a phosphorylation-mediated signaling cascade [35]. This reveals the mechanism behind these mammals' resilience to cardiac stresses during hibernation that would otherwise be damaging, but which might be useful for protecting astronauts involved in extended interplanetary missions.

5. Immune Suppression During Hibernation

It has been reported that some astronauts experience allergy-like symptoms during spaceflight [36]. The primary lymphoid organs, such as bone marrow and the thymus, are affected by gravitational change during spaceflight. In rodents, short- and long-term spaceflight cause functional changes of the thymus and lead to changes in immune signaling and cell proliferation [37][38]. These changes may affect acquired immune responses to pathogens, allergens, and tumors [30]. Changes in the immune system during space missions might explain the astronauts' symptoms. A previous study showed that ionizing radiation reduces circulating T and B cell populations. In contrast, macrophages and natural killer and dendritic cells are more radioresistant [35]. Among the immune cells, T cells play a central role in the host's adaptive immunity against many intracellular pathogens and clearing viruses. Reduced T cells might underlie the reactivation and shedding of latent human herpesviruses, such as varicella-zoster virus, Epstein–Barr virus, and human cytomegalovirus, as happened during the Russian Soyuz and International Space Station missions [39][40].

It has been reported that low body temperatures in hibernators such as brown bears (*U. arctos*) might also be correlated to suppression of their immune systems [41]. During hibernation, the animals can suppress their immunity. A study showed that circulating leukocytes drop by ~90% during entrance into torpor, driven by a low body temperature. In hibernation, there is a reduced capacity to induce an immune response [42]. The immune system is not able to attack a bacterial infection during hibernation, but will react strongly upon arousal [43]. Those studies may provide clues about how hibernation can efficiently control the immune system and facilitate future space missions.

6. Thermoregulation and Muscle Preservation During Hibernation

Previous reports showed that astronauts' core body temperatures increased significantly, and did so even more with exercise ^[44]. Temperature plays an essential role in radiation-induced damage and enzymatic processes. A low temperature also influences radiation sensitivity, due to the change in activity of several enzymes when exposed to different temperatures. Exposure to ionizing radiation at low temperatures has been shown to lower the activity of enzymes, resulting in decreased radiation sensitivity^{[45][46]}. A study showed that an enzyme such as malate dehydrogenase, which is an essential enzyme for protection against oxidative damage, is inactive when it is irradiated at lower temperatures ^[45], which might be harmful. On the other hand, the enzyme lactate dehydrogenase (LDH), which may cause tissue damage, is shown to be temperature-dependent or radiation-sensitive [64]. Those studies indicated that the drop in temperature during torpor might be somewhat radiation protective by deactivating enzymes or may have no protection against radiation.

Exercise is an important activity of astronauts to preserve their physical health, muscle mass, and cardiovascular health ^[44]. As previously mentioned, exercise is important in space and affects the astronauts' thermoregulation ^{[44][47]}. One attractive advantage of hibernation is that it preserves physical health, including muscle mass. Naturally, muscle mass will be reduced when muscles are not used for a long time. However, hibernating mammals demonstrate limited muscle loss over prolonged immobile intervals during the winter ^[48]. Studies to understand muscle preservation in hibernating animals show that bears reabsorb their urea, which is used to form new amino acids ^[49]. This helps them minimize the loss of lean muscle during this prolonged inactivity. However, this is not the case for humans. Muscle atrophy in astronauts during space travel is caused by microgravity or the absence of gravity beyond low Earth orbit. Muscles such as the calf and quadriceps, as well as back and neck muscles, are commonly called antigravity muscles. In microgravity, since these muscles are not being used, they atrophy. Muscle loss is also presumably caused by changes in muscle metabolism, namely the process of building and breaking down proteins. Experiments performed during long-term missions onboard the Russian Mir revealed a decrease of about 15% in the rate of protein synthesis and alterations in the structure and function of skeletal muscle fiber in humans ^{[50][51]}. Therefore, understanding how hibernators preserve their muscle might help astronauts keep their strength and physical health.

References

1. Musacchia, X.J.; Barr, R.E. Survival of whole-body-irradiated hibernating and active ground squirrels; *Citellus tridecemlineatus*. *Radiat. Res.* 1968, 33, 348–356.
2. Jaroslow, B.N.; Fry, R.J.; Suhrbier, K.M.; Sallese, A.R. Radiosensitivity of ileum crypt cells in hibernating, arousing, and awake ground squirrels (*Citellus tridecemlineatus*). *Radiat. Res.* 1976,

66, 566–575.

3. Kuskin, S.M.; Wang, S.C.; Rugh, R. Protective effect of artificially induced hibernation against lethal doses of whole body X-irradiation in CF male mice. *Am. J. Physiol.* 1959, 196, 1211–1213, doi:10.1152/ajplegacy.1959.196.6.1211.
4. Miller, I. *A History of Force Feeding: Hunger Strikes, Prisons and Medical Ethics*; Palgrave Macmillan: Basingstoke, UK, 2016; pp. 1917–1923.
5. Paxinos, G.; Watson, G. *The Rat Brain in Stereotaxic Coordinates*, 6th ed.; Elsevier: San Diego, CA, USA, 2007; pp. 172–178.
6. Shuryak, I.; Sachs, R.K.; Brenner, D.J. Cancer risks after radiation exposure in middle age. *J. Natl. Cancer Inst.* 2010, 102, 1628–1636, doi:10.1093/jnci/djq346.
7. Hendee, W.R. Estimation of radiation risks. BEIR V and its significance for medicine. *JAMA* 1992, 268, 620–624.
8. George, K.; Durante, M.; Wu, H.; Willingham, V.; Badhwar, G.; Cucinotta, F.A. Chromosome aberrations in the blood lymphocytes of astronauts after space flight. *Radiat. Res.* 2001, 156, 731–738, doi:10.1667/0033-7587(2001)156[0731:caitbl]2.0.co;2.
9. George, K.; Rhone, J.; Beitman, A.; Cucinotta, F.A. Cytogenetic damage in the blood lymphocytes of astronauts: Effects of repeat long-duration space missions. *Mutat. Res.* 2013, 756, 165–169, doi:10.1016/j.mrgentox.2013.04.007.
10. Schwartz, C.; Hampton, M.; Andrews, M.T. Seasonal and regional differences in gene expression in the brain of a hibernating mammal. *PLoS ONE* 2013, 8, e58427, doi:10.1371/journal.pone.0058427.
11. Uziel, T.; Lerenthal, Y.; Moyal, L.; Andegeko, Y.; Mittelman, L.; Shiloh, Y. Requirement of the MRN complex for ATM activation by DNA damage. *EMBO J.* 2003, 22, 5612–5621, doi:10.1093/emboj/cdg541.
12. Tinganelli, W.; Hitrec, T.; Romani, F.; Simoniello, P.; Squarcio, F.; Stanzani, A.; Piscitiello, E.; Marchesano, V.; Luppi, M.; Sioli, M.; et al. Hibernation and radioprotection: Gene expression in the liver and testicle of rats irradiated under synthetic torpor. *Int. J. Mol. Sci.* 2019, 20, 352, doi:10.3390/ijms20020352.
13. Pietzner, J.; Merscher, B.M.; Baer, P.C.; Duecker, R.P.; Eickmeier, O.; Fußbroich, D.; Bader, P.; Del Turco, D.; Henschler, R.; Zielen, S.; et al. Low-dose irradiation prior to bone marrow transplantation results in ATM activation and increased lethality in ATM-deficient mice. *Bone Marrow Transplant.* 2016, 51, 560–567, doi:10.1038/bmt.2015.334.
14. Acharya, M.M.; Patel, N.H.; Craver, B.M.; Tran, K.K.; Giedzinski, E.; Tseng, B.P.; Parihar, V.K.; Limoli, C.L. Consequences of low dose ionizing radiation exposure on the hippocampal

- microenvironment. PLoS ONE 2015, 10, e0128316, doi:10.1371/journal.pone.0128316.
15. Parihar, V.K.; Limoli, C.L. Cranial irradiation compromises neuronal architecture in the hippocampus. *Proc. Natl. Acad. Sci. USA* 2013, 110, 12822–12827, doi:10.1073/pnas.1307301110.
 16. Parihar, V.K.; Maroso, M.; Syage, A.; Allen, B.D.; Angulo, M.C.; Soltesz, I.; Limoli, C.L. Persistent nature of alterations in cognition and neuronal circuit excitability after exposure to simulated cosmic radiation in mice. *Exp. Neurol.* 2018, 305, 44–55, doi:10.1016/j.expneurol.2018.03.009.
 17. Hartwell, L.H.; Kastan, M.B. Cell cycle control and cancer. *Science* 1994, 266, 1821–1828, doi:10.1126/science.7997877.
 18. Kudo, S.; Suzuki, Y.; Noda, S.E.; Mizui, T.; Shirai, K.; Okamoto, M.; Kaminuma, T.; Yoshida, Y.; Shirao, T.; Nakano, T. Comparison of the radiosensitivities of neurons and glial cells derived from the same rat brain. *Exp. Ther. Med.* 2014, 8, 754–758, doi:10.3892/etm.2014.1802.
 19. Shirai, K.; Mizui, T.; Suzuki, Y.; Okamoto, M.; Hanamura, K.; Yoshida, Y.; Hino, M.; Noda, S.E.; Al-jahdari, W.S.; Chakravar-ti, A.; et al. X irradiation changes dendritic spine morphology and density through reduction of cytoskeletal proteins in mature neurons. *Radiat. Res.* 2013, 179, 630–636, doi:10.1667/RR3098.1.
 20. Okamoto, M.; Suzuki, Y.; Shirai, K.; Mizui, T.; Yoshida, Y.; Noda, S.E.; Al-Jahdari, W.S.; Shirao, T.; Nakano, T. Effect of radiation on the development of immature hippocampal neurons in vitro. *Radiat. Res.* 2009, 172, 718–724, doi:10.1667/RR1741.1.
 21. Puspitasari, A.; Yamazaki, H.; Kawamura, H.; Nakano, T.; Takahashi, A.; Shirao, T.; Held, K.D. X-irradiation of developing hippocampal neurons causes changes in neuron population phenotypes, dendritic morphology and synaptic protein expression in surviving neurons at maturity. *Neurosci. Res.* 2019, 160, 11–24, doi:10.1016/j.neures.2019.11.002.
 22. Puspitasari, A.; Koganezawa, N.; Ishizuka, Y.; Kojima, N.; Tanaka, N.; Nakano, T.; Shirao, T. X irradiation induces acute cognitive decline via transient synaptic dysfunction. *Radiat. Res.* 2016, 185, 423–430, doi:10.1667/RR14236.1.
 23. Miao, S.; Koganezawa, N.; Hanamura, K.; Puspitasari, A.; Shirao, T. N-methyl-D-aspartate receptor mediates X-irradiation-induced drebrin decrease in hippocampus. *Kitakanto Med. J.* 2018, 68, 111–115.
 24. Mikhailova, A. Recovery of syrian hamster hippocampal signaling following its depression during oxygen-glucose deprivation is enhanced by cold temperatures and by hibernation. *Neurosci. Lett.* 2016, 621, 98–103, doi:10.1016/j.neulet.2016.04.011.
 25. Chatfield, P.O.; Lyman, C.P.; Purpura, D.P. The effects of temperature on the spontaneous and induced electrical activity in the cerebral cortex of the golden hamster. *Electroencephalogr. Clin. Neurophysiol.* 1951, 3, 225–230, doi:10.1016/0013-4694(51)90015-6.

26. Krilowicz, B.L.; Glotzbach, S.F.; Heller, H.C. Neuronal activity during sleep and complete bouts of hibernation. *Am. J. Physiol.* 1988, 255, R1008–R1019, doi:10.1152/ajpregu.1988.255.6.R1008.
27. Krilowicz, B.L.; Edgar, D.M.; Heller, H.C. Reticular formation activity during wakefulness and sleep in a hibernator (*Spermophilus lateralis*). *Brain Res.* 1991, 540, 266–272, doi:10.1016/0006-8993(91)90516-x.
28. Shirao, T.; Hanamura, K.; Koganezawa, N.; Ishizuka, Y.; Yamazaki, H.; Sekino, Y. The role of drebrin in neurons. *J. Neurochem.* 2017, 141, 819–834, doi:10.1111/jnc.13988.
29. Duman, J.G.; Dinh, J.; Zhou, W.; Cham, H.; Mavratsas, V.C.; Paveškov, M.; Mulherkar, S.; McGovern, S.L.; Tolias, K.F.; Grosshans, D.R. Memantine prevents acute radiation-induced toxicities at hippocampal excitatory synapses. *Neuro Oncol.* 2018, 20, 655–665, doi:10.1093/neuonc/nox203.
30. Ross, A.P.; Christian, S.L.; Zhao, H.W.; Drew, K.L. Persistent tolerance to oxygen and nutrient deprivation and N-methyl-D-aspartate in cultured hippocampal slices from hibernating arctic ground squirrel. *J. Cereb. Blood Flow Metab.* 2006, 26, 1148–1156, doi:10.1038/sj.jcbfm.9600271.
31. Hughson, R.L.; Shoemaker, J.K.; Blaber, A.P.; Arbeille, P.; Greaves, D.K.; Pereira-Junior, P.P.; Xu, D. Cardiovascular regulation during long-duration spaceflights to the International Space Station. *J. Appl. Physiol.* (1985) 2012, 112, 719–727, doi:10.1152/jappphysiol.01196.2011.
32. Hada, M.; Ikeda, H.; Rhone, J.R.; Beitman, A.J.; Plante, I.; Souda, H.; Yoshida, Y.; Held, K.D.; Fujiwara, K.; Saganti, P.B.; et al. Increased chromosome aberrations in cells exposed simultaneously to simulated microgravity and radiation. *Int. J. Mol. Sci.* 2019, 20, 43, doi:10.3390/ijms20010043.
33. Leach, C.S.; Alfrey, C.P.; Suki, W.N.; Leonard, J.I.; Rambaut, P.C.; Inners, L.D.; Smith, S.M.; Lane, H.W.; Krauhs, J.M. Regulation of body fluid compartments during short-term spaceflight. *J. Appl. Physiol.* (1985) 1996, 81, 105–116, doi:10.1152/jappl.1996.81.1.105.
34. Nelson, O.L.; McEwen, M.M.; Robbins, C.T.; Felicetti, L.; Christensen, W.F. Evaluation of cardiac function in active and hibernating grizzly bears. *J. Am. Vet. Med. Assoc.* 2003, 223, 1170–1175, doi:10.2460/javma.2003.223.1170.
35. Childers, C.L.; Tessier, S.N.; Storey, K.B. The heart of a hibernator: EGFR and MAPK signaling in cardiac muscle during the hibernation of thirteen-lined ground squirrels, *Ictidomys tridecemlineatus*. *PeerJ* 2019, 7, e7587, doi:10.7717/peerj.7587.
36. Akiyama, T.; Horie, K.; Hinoi, E.; Hiraiwa, M.; Kato, A.; Maekawa, Y.; Takahashi, A.; Furukawa, S. How does spaceflight affect the acquired immune system? *npj Microgravity* 2020, 6, 14, doi:10.1038/s41526-020-0104-1.

37. Novoselova, E.G.; Lunin, S.M.; Khrenov, M.O.; Parfenyuk, S.B.; Novoselova, T.V.; Shenkman, B.S.; Fesenko, E.E. Changes in immune cell signalling, apoptosis and stress response functions in mice returned from the BION-M1 mission in space. *Im-munobiology* 2015, 220, 500–509, doi:10.1016/j.imbio.2014.10.021.
38. Tajima, G.; Delisle, A.J.; Hoang, K.; O'Leary, F.M.; Ikeda, K.; Hanschen, M.; Stoecklein, V.M.; Lederer, J.A. Immune system phenotyping of radiation and radiation combined injury in outbred mice. *Radiat. Res.* 2013, 179, 101–112, doi:10.1667/RR3120.1.
39. Rooney, B.V.; Crucian, B.E.; Pierson, D.L.; Laudenslager, M.L.; Mehta, S.K. Herpes virus reactivation in astronauts during spaceflight and its application on earth. *Front. Microbiol.* 2019, 10, 16, doi:10.3389/fmicb.2019.00016.
40. Mehta, S.K.; Laudenslager, M.L.; Stowe, R.P.; Crucian, B.E.; Feiveson, A.H.; Sams, C.F.; Pierson, D.L. Latent virus reactivation in astronauts on the International Space Station. *npj Microgravity* 2017, 3, 11, doi:10.1038/s41526-017-0015-y.
41. Sahdo, B.; Evans, A.L.; Arnemo, J.M.; Fröbert, O.; Särndahl, E.; Blanc, S. Body temperature during hibernation is highly cor-related with a decrease in circulating innate immune cells in the brown bear (*Ursus arctos*): A common feature among hi-bernators? *Int. J. Med. Sci.* 2013, 10, 508–514, doi:10.7150/ijms.4476.
42. Shivatcheva, T.M. Survival of skin allografts in European ground squirrels, *Spermophilus citellus* L., during hibernation. *Folia Biol. (Krakow)* 1988, 36, 213–221.
43. Suomalainen, P.; Rosokivi, V. Studies on the physiology of the hibernating hedgehog. 17. The blood cell count of the hedgehog at different times of the year and in different phases of the hibernating cycle. *Ann. Acad. Sci. Fenn. Biol.* 1973, 198, 1–8.
44. Laurens, C.; Simon, C.; Vernikos, J.; Gauquelin-Koch, G.; Blanc, S.; Bergouignan, A. Revisiting the role of exercise counter-measure on the regulation of energy balance during space flight. *Front. Physiol.* 2019, 10, 321, doi:10.3389/fphys.2019.00321.
45. Kempner, E.S.; Haigler, H.T. The influence of low temperature on the radiation sensitivity of enzymes. *J. Biol. Chem.* 1982, 257, 13297–13299.
46. Fluke, D.J. Temperature dependence of the direct action of ionizing radiation on beef heart lactate dehydrogenase: Enzyme activity, substrate and coenzyme affinities. *Radiat. Res.* 1972, 51, 56–71.
47. Stahn, A.C.; Werner, A.; Opatz, O.; Maggioni, M.A.; Steinach, M.; von Ahlefeld, V.W.; Moore, A.; Crucian, B.E.; Smith, S.M.; Zwart, S.R.; et al. Increased core body temperature in astronauts during long-duration space missions. *Sci. Rep.* 2017, 7, 16180, doi:10.1038/s41598-017-15560-w.
48. Goropashnaya, A.V.; Barnes, B.M.; Fedorov, V.B. Transcriptional changes in muscle of hibernating arctic ground squirrels (*Urocitellus parryi*): Implications for attenuation of disuse

muscle atrophy. *Sci. Rep.* 2020, 10, 9010, doi:10.1038/s41598-020-66030-9.

49. Stenvinkel, P.; Jani, A.H.; Johnson, R.J. Hibernating bears (Ursidae): Metabolic magicians of definite interest for the nephrologist. *Kidney Int.* 2013, 83, 207–212, doi:10.1038/ki.2012.396.
50. Di Prampero, P.E.; Narici, M.V. Muscles in microgravity: From fibres to human motion. *J. Biomech.* 2003, 36, 403–412, doi:10.1016/s0021-9290(02)00418-9.
51. Fitts, R.H.; Trappe, S.W.; Costill, D.L.; Gallagher, P.M.; Creer, A.C.; Colloton, P.A.; Peters, J.R.; Romatowski, J.G.; Bain, J.L.; Riley, D.A. Prolonged space flight-induced alterations in the structure and function of human skeletal muscle fibres. *J. Physiol.* 2010, 588, 3567–3592, doi:10.1113/jphysiol.2010.188508.

Retrieved from <https://encyclopedia.pub/entry/history/show/17693>