

# Microbiota, Diet and Mucus in Inflammatory Bowel Disease

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The gastrointestinal tract is optimized to efficiently absorb nutrients and provide a competent barrier against a variety of lumen environmental compounds. Different regulatory mechanisms jointly collaborate to maintain intestinal homeostasis, but alterations in these mechanisms lead to a dysfunctional gastrointestinal barrier and are associated to several inflammatory conditions usually found in chronic pathologies such as inflammatory bowel disease (IBD). The gastrointestinal mucus, mostly composed of mucin glycoproteins, covers the epithelium and plays an essential role in digestive and barrier functions. However, its regulation is very dynamic and is still poorly understood. This review presents some aspects concerning the role of mucus in gut health and its alterations in IBD. In addition, the impact of gut microbiota and dietary compounds as environmental factors modulating the mucus layer is addressed. To date, studies have evidenced the impact of the three-way interplay between the microbiome, diet and the mucus layer on the gut barrier, host immune system and IBD. This review emphasizes the need to address current limitations on this topic, especially regarding the design of robust human trials and highlights the potential interest of improving our understanding of the regulation of the intestinal mucus barrier in IBD.

Keywords: dietary compounds ; gastrointestinal barrier ; gut microbiota ; inflammatory bowel disease ; mucus layer

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## 1. Introduction to Inflammatory Bowel Disease

Inflammatory bowel disease (IBD) is a global disease associated to Western and recently westernized countries <sup>[1]</sup>. The emergence of this disease was parallel to the industrial revolution in the 1800s <sup>[2]</sup>. Being a chronic disease diagnosed early in life, the prevalence of this pathology is high and is increasing over time. Prevalence of IBD was 84 per 100,000 population in 2017 <sup>[3]</sup> and it has been estimated that it will continue increasing in the next generation, affecting tens of millions of people all over the world <sup>[4]</sup>. Therefore, the cost of this disease for health care systems is considerable and will increase steadily in the future <sup>[4][5]</sup>.

The origin and causes of IBD remain unknown. It is an immune-mediated inflammatory disease and its major causative factors could be genetic, immune and environmental such as the gut microbiome and diet. Genome wide-association studies identified approximately 200 gene loci in IBD, of which more than 50% are also associated with other inflammatory and autoimmune diseases <sup>[6]</sup>. The exposure to environmental conditions influence the microbiome composition and the consequent dysbiosis (changes in the healthy microbiota) in the gastrointestinal tract can trigger inflammatory responses <sup>[7][8]</sup>.

IBD is a general term encompassing ulcerative colitis (UC) and Crohn's disease (CD). UC is limited to the colon and presents superficial mucosal inflammation that can lead to ulcerations and bleeding. CD can affect any part of the digestive tract and presents transmural inflammation and complications such as fistulas or abscesses <sup>[9]</sup>; furthermore, IBD is associated to other extra-intestinal pathologies such as arthritis and skin diseases that aggravate the quality of life of these patients. Both IBD subtypes present periods of inflammation and quiescence <sup>[10]</sup>. Regarding IBD therapeutic approaches, several drugs have been developed over the last years, including biologics that target different molecules involved in IBD pathogenesis <sup>[11][12]</sup>. However, response to treatment is highly variable <sup>[13][14]</sup> and, since there is no cure for this disease, the therapeutic goal is to maintain patients' remission. Accordingly, a deeper understanding of the disease is needed to improve treatment of these patients.

In this review, we will focus on the gastrointestinal barrier in IBD with a particular emphasis on the role of the mucus layer in gut health and its alterations in this disease. In addition, the impact of the gut microbiota and dietary compounds as mucus modulatory factors and their complex interaction with the mucosal barrier in IBD is summarized. Data were obtained from articles published in English in journals indexed in PubMed and Web of Science from inception to August 2021 and retrieved using search terms related to (i) gastrointestinal barrier and gut homeostasis; (ii) mucus layer, mucins

and IBD; (iii) modulation of immune system and mucosal inflammation; (iv) gut microbiota, probiotics and IBD; (v) dietary compounds, food bioactives and IBD.

## **2. Gastrointestinal Barrier**

The intestinal mucosal barrier provides adequate containment of microorganisms and molecules, preserving the capacity to absorb nutrients [15]. Intestinal mucosa is covered with a monolayer of intestinal epithelial cells (IECs) that separate the external environment and sub-epithelium [16]. Alterations in this mucosal barrier may result in IBD, stressing its essential role to maintain a healthy gut environment [17]. A key regulator balancing this relationship is the gastrointestinal mucus layer, composed of a secreted mucus gel, which covers the surface of epithelium and the underlying mucosal immune system. Hence, the gut mucosa is protected by two barrier types: chemical and physical. Chemical barriers participate in the segregation of IECs and gut microbiota [18]. IECs are derived from stem cells within intestinal crypts that replicate and migrate towards villi to replenish the active turnover of epithelium [15]. Functionally, secretory IECs, as goblet and Paneth cells, are specialized in maintaining the epithelial barrier function [19]. Paneth cells are involved in the production of chemical barriers such as antimicrobial peptides in the small intestine [20], while goblet cells secrete mucins. Mucins and antimicrobial peptides are important for both physical and biochemical barriers. The different functions of IECs lead to a dynamic barrier, which protects the host from infection and inflammatory stimuli [19]. IECs act as sensors for microbial elements and can integrate signals from commensal bacteria into antimicrobial and immune regulatory responses [21]. These functions are enabled by the expression of pattern-recognition receptors that act as sensors of the microbial environment and are key regulatory elements in mucosal immune responses [19].

Mucosal homeostasis is a vital feature of the gut immune system [22]. One of the critical factors for developing IBD is the failure to maintain an adequate balance between response to pathogens and tolerance to commensal microorganisms and luminal beneficial antigens [23][24]. Under the conditions of gut barrier dysfunction, as it occurs in IBD, the homeostatic equilibrium is lost [25][26]. IBD is related with increased permeability in the gut and the associated disbalance in the immune response that leads to increased recruitment of circulating cells and secretion of pro-inflammatory mediators [15][27]. Therefore, factors as immune system, genetics and environmental ones influence the gastrointestinal barrier function and are, thus, involved in the “IBD integrome” [28].

### **2.1. Mucus Layer**

The small intestine has a single mucus layer that facilitates the pass of nutrients, while the colon is covered by a thicker barrier. However, in the colon, the mucus layer acts as a physical barrier maintaining bacteria in symbiosis with the host and preventing bacterial infiltration into the epithelium [16][18]. The large intestine epithelium is, thus, covered by two mucus layers: an outer loose layer and an inner firm mucus attached to the epithelia [29][30]. The principal components of the gastrointestinal mucus barrier are O-linked glycoproteins called mucins. They present densely packed oligosaccharides that bind to their terminal region sialic acid and sulfate residues protecting mucins from proteases and glycosidases [31]. Mucins are produced by goblet cells present within the intestinal epithelium [32]. Mucus exocytosis from goblet cells depends on several cellular processes that modulate mucin secretion, including endocytosis and autophagy [32].

There are 18 mucin members in humans classified in two types: transmembrane and secreted mucins. Mucin central domains are composed of proline, threonine and serine (PTS) residues working as attachment sites for O-linked glycans through covalent binding of N-acetylgalactosamine to serine or threonine residues [16]. The secreted mucin MUC2 is the main glycoprotein in the intestinal mucus. MUC2 has an N-terminal domain, two PTS domains and a C-terminal domain. MUC2 N-terminal domain comprises 3 complete von Willebrand factor domains (D1-3) and the C-terminal region of D4 domain. Cysteine residues in N- and C- terminal domains facilitate inter- and intramolecular disulfide bond formation responsible for mucin polymerization [33].

MUC2 polypeptide is synthesized and dimerized in the endoplasmic reticulum of intestinal cells. Then, threonine and serine residues are glycosylated in the cis-Golgi and the trimer formation takes place in the trans-Golgi before MUC2 is packaged into secretory granules. MUC2 is composed of heterogeneous glycan chains [16], which allow MUC2 trimers to form polymers creating mucus networks in the cell surface [31][34]. MUC2 polymers undergo rapid expansion on the intestinal epithelial surface to maintain the mucus barrier during homeostasis; this expansion depends on ionic composition and water availability. Polymers can expand their volume up to 1000 times to form the framework of the mucus gel [35].

On the other hand, intestinal transmembrane mucins (MUC1, MUC3, MUC4 and MUC13) are intercalated in the apical surface of the intestinal epithelium forming the glycocalyx layer [32]. In contrast to the sterile inner layer of mucus, the outer

mucus layer is rich in gut bacteria [29]. These bacteria use diet fiber as energy source; however, under a fiber-free diet they consume MUC2 polysaccharides, leading to a thinner inner mucus layer and dysbiosis [36], as well as bacteria penetration into the lamina propria contributing to IBD development [18].

## 2.2. Mucus Layer under Inflammatory Conditions

The stability of the mucus layer is crucial for intestinal homeostasis, in which MUC2 is secreted at a basal rate. This secretion can be influenced by mediators as cytokines, microbial products, autophagic proteins, reactive oxygen species and inflammasome components [37][38]. Commensal and pathogenic bacteria can regulate mucin production [28]. In the small intestine, a continuous basal secretion of mucus creates a flow towards the lumen that, together with antibacterial agents, keeps microorganisms away from the epithelial surface. Antibacterial agents are secreted by Paneth cells and enterocytes of the crypt bottom. On the other hand, in the colon, the inner mucus layer is the first line of defense against bacteria [39].

The mucus layer is a natural and selective habitat for the gut microbiota [40], which in turn influences mucus composition and may promote mucus secretion and increase mucus layer thickness [41]. Therefore, the gut microbiota affects mucus layer function, possibly through specific bacteria that shape the glycan profile of the mucus, although molecular details remain incompletely identified [42].

There is high number of enteropathogens that have evolved mechanisms to penetrate the mucus barrier. Most of them produce a kind of serine proteases that cleave glycoproteins such as mucins [43]. Moreover, cytokines are involved in the inflammatory response and regulate many cellular and molecular processes including mucus production. In this regard, TNF- $\alpha$  and IL-1 $\beta$ , which are implicated in inflammatory diseases, stimulate gel-forming mucins [43]. Th2 cytokines are implicated in mucin gene expression up-regulating MUC2 and MUC5AC by binding to IL-4 receptor. Endoplasmic reticulum stress in goblet cells produce immature mucins that trigger inflammation [44][45], whereas IL-10 has been found to inhibit endoplasmic reticulum stress and promote intestinal mucus production [43][46].

MUC2 knockout mice show colonization of gut epithelium by enteric pathogens [47][48]. These results suggest that the principal mucus function is to protect the gut against microbes. Binding to mucin oligosaccharide chains likely contributes to immobilize bacteria and prevents them from damaging the intestinal epithelium. MUC2 has also immune roles; small intestine goblet cells provide the passage of soluble luminal antigens by transcytosis. These low molecular weight antigens are delivered to underlying CD103<sup>+</sup> dendritic cells and may favor IgA production and expansion of regulatory T cells, thereby driving gut homeostasis and tolerance [49]. The commensal microbiota, through its relationship with mucus, prevents colonization by pathogens. In this regard, when antibiotics perturb the gut microbiota, niches are opened facilitating disease development. The gut microbiota also breaks down short-chain fatty acids (SCFA) including acetate, propionate and butyrate [50]. Since butyrate regulates MUC2 production, the microbiota is also involved in the homeostasis of the protective mucus layer [51].

Mucin composition is altered in IBD and mucin structural changes play an important role in IBD onset [52][53]. In fact, alterations of mucus barrier and mucins are observed at IBD onset; goblet cell pathology is a hallmark of UC and CD [43]. Recently, it has been observed that the reduced mucus layer in UC is due to a reduction in the number and secretory function of goblet cells because of an inflammatory environment and due to changes in mucin secretion that persist in the absence of inflammatory cells [54].

The mucus layer is thinner in UC than in the healthy colon, while goblet cell depletion and altered MUC2 glycosylation can be also observed; in addition, MUC2 is undersulfated, weakening mucin protective function [55][56][57]. Despite these results, the expression pattern of MUC2 in UC is not clear. Conversely, MUC5AC, is consistently increased during inflammation in UC [58][59] and its reduced expression is associated with endoscopic improvement in these patients [60]. In Muc5ac<sup>-/-</sup> mice with DSS colitis, there is an increase in bacterial-epithelial contact and neutrophil recruitment to the colon, therefore, the loss of Muc5ac may exacerbate injury and inflammation in experimental murine colitis [61]. This study also showed a significant increase in MUC5AC/Muc5ac expression during colonic inflammation in biopsies from UC patients and DSS-induced mice colitis [61].

In contrast, mucus thickness is normal or greater than normal in CD, maybe due to goblet cell hyperplasia or increased MUC2 expression, although with a 50% reduction in oligosaccharide chain length [62]. Hence, several changes in the mucosal barrier underlie the complex pathology of IBD.

### 3. Gut Microbiota and the Mucus Layer in IBD

The microbiome plays key roles in the development of mucosal immune responses, pathogen resistance and nutrient metabolism. This fact is in part due to the interaction of the microbiota with components of the mucus layer and the IECs underneath following mucus breakdown. Intestinal barrier, antimicrobial and immunomodulatory functions are influenced by several members of the gut microbiota, as reviewed in the present review.

### 4. Dietary Compounds and the Mucus Layer in IBD

Dietary factors need to be considered when evaluating the complex relationship between the host, microbiota and the mucus layer. Dietary patterns and specific foods or nutrients may affect the gut barrier directly or indirectly by shaping microbial species known to influence mucosal protection and inflammatory processes. The influence of the different food groups/compounds in the mucus barrier was summarised in the present review.

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#### References

1. Ng, S.C.; Shi, H.Y.; Hamidi, N.; Underwood, F.E.; Tang, W.; Benchimol, E.I.; Panaccione, R.; Ghosh, S.; Wu, J.C.Y.; Chan, F.K.L.; et al. Worldwide incidence and prevalence of inflammatory bowel disease in the 21st century: A systematic review of population-based studies. *Lancet* 2017, 390, 2769–2778.
2. Mulder, D.J.; Noble, A.J.; Justinich, C.J.; Duffin, J.M. A tale of two diseases: The history of inflammatory bowel disease. *J. Crohns Colitis* 2014, 8, 341–348.
3. Alatab, S.; Sepanlou, S.G.; Ikuta, K.; Vahedi, H.; Bisignano, C.; Safiri, S.; Sadeghi, A.; Nixon, M.R.; Abdoli, A.; Abolhassani, H.; et al. The global, regional, and national burden of inflammatory bowel disease in 195 countries and territories, 1990–2017: A systematic analysis for the Global Burden of Disease Study 2017. *Lancet Gastroenterol. Hepatol.* 2020, 5, 17–30.
4. Kaplan, G.G.; Ng, S.C. Understanding and preventing the global increase of inflammatory bowel disease. *Gastroenterology* 2017, 152, 313–321.
5. Molodecky, N.A.; Soon, I.S.; Rabi, D.M.; Ghali, W.A.; Ferris, M.; Chernoff, G.; Benchimol, E.I.; Panaccione, R.; Ghosh, S.; Barkema, H.W.; et al. Increasing incidence and prevalence of the inflammatory bowel diseases with time, based on systematic review. *Gastroenterology* 2012, 142, 46–54.e42.
6. Khor, B.; Gardet, A.; Ramnik, J.X. Genetics and Pathogenesis of Inflammatory Bowel Disease. *Nature* 2011, 474, 307–317.
7. Morgan, X.C.; Tickle, T.L.; Sokol, H.; Gevers, D.; Devaney, K.L.; Ward, D.V.; Reyes, J.A.; Shah, S.A.; LeLeiko, N.; Snapper, S.B.; et al. Dysfunction of the intestinal microbiome in inflammatory bowel disease and treatment. *Genome Biol.* 2012, 13, R79.
8. Aldars-García, L.; Chaparro, M.; Gisbert, J.P. Systematic review: The gut microbiome and its potential clinical application in inflammatory bowel disease. *Microorganisms* 2021, 9, 977.
9. Hoentjen, F.; Dieleman, L.A. Pathophysiology of inflammatory bowel diseases. *Handb. Prebiotics* 2008, 341–374.
10. Peyrin-Biroulet, L.; Chamaillard, M.; Gonzalez, F.; Beclin, E.; Decourcelle, C.; Antunes, L.; Gay, J.; Neut, C.; Colombel, J.F.; Desreumaux, P. Mesenteric fat in Crohn's disease: A pathogenetic hallmark or an innocent bystander? *Gut* 2007, 56, 577–583.
11. Bonovas, S.; Pantavou, K.; Evripidou, D.; Bastiampillai, A.J.; Nikolopoulos, G.K.; Peyrin-Biroulet, L.; Danese, S. Safety of biological therapies in ulcerative colitis: An umbrella review of meta-analyses. *Best Pract. Res. Clin. Gastroenterol.* 2018, 32–33, 43–47.
12. Weisshof, R.; ElJurdi, K.; Zmeter, N.; Rubin, D. Emerging therapies for inflammatory bowel diseases. *Dig. Dis.* 2016, 34, 67–73.
13. Gisbert, J.P.; Chaparro, M. Predictors of primary response to biologic treatment in patients with inflammatory bowel disease: From basic science to clinical practice. *J. Crohns Colitis* 2020, 14, 694–709.
14. Digby-Bell, J.L.; Atreya, R.; Monteleone, G.; Powell, N. Interrogating host immunity to predict treatment response in inflammatory bowel disease. *Nat. Rev. Gastroenterol. Hepatol.* 2020, 17, 9–20.
15. De Medina, F.S.; Romero-Calvo, I.; Mascaraque, C.; Martínez-Augustín, O. Intestinal inflammation and mucosal barrier function. *Inflamm. Bowel Dis.* 2014, 20, 2394–2404.
16. Sharpe, C.; Thornton, D.J.; Grecis, R.K. A sticky end for gastrointestinal helminths; the role of the mucus barrier. *Parasite Immunol.* 2018, 1–10.

17. König, J.; Wells, J.; Cani, P.D.; García-Ródenas, C.L.; MacDonald, T.; Mercenier, A.; Whyte, J.; Troost, F.; Brummer, R.J. Human intestinal barrier function in health and disease. *Clin. Transl. Gastroenterol.* 2016, 7, e196.
18. Okumura, R.; Takeda, K. Maintenance of intestinal homeostasis by mucosal barriers. *Inflamm. Regen.* 2018, 38, 5.
19. Peterson, L.W.; Artis, D. Intestinal epithelial cells: Regulators of barrier function and immune homeostasis. *Nat. Rev. Immunol.* 2014, 14, 141–153.
20. Salzman, N.H.; Underwood, M.A.; Bevins, C.L. Paneth cells, defensins, and the commensal microbiota: A hypothesis on intimate interplay at the intestinal mucosa. *Semin. Immunol.* 2007, 19, 70–83.
21. Aldars-García, L.; Marin, A.C.; Chaparro, M.; Gisbert, J.P. The interplay between immune system and microbiota in inflammatory bowel disease: A narrative review. *Int. J. Mol. Sci.* 2021, 22, 3706.
22. Sartor, R.B. Genetics and environmental interactions shape the intestinal microbiome to promote inflammatory bowel disease versus mucosal homeostasis. *Gastroenterology* 2010, 139, 1816–1819.
23. Blander, J.M.; Longman, R.S.; Iliev, I.D.; Sonnenberg, G.F.; Artis, D. Regulation of inflammation by microbiota interactions with the host. *Nat. Immunol.* 2017, 18, 851–860.
24. Fernández-Tomé, S.; Marin, A.C.; Moreno, L.O.; Baldan-Martin, M.; Mora-Gutiérrez, I.; Lanás-Gimeno, A.; Moreno-Monteagudo, J.A.; Santander, C.; Sánchez, B.; Chaparro, M.; et al. Immunomodulatory effect of gut microbiota-derived bioactive peptides on human immune system from healthy controls and patients with inflammatory bowel disease. *Nutrients* 2019, 11, 2605.
25. Maloy, K.J.; Powrie, F. Intestinal homeostasis and its breakdown in inflammatory bowel disease. *Nature* 2011, 474, 298–306.
26. Bernardo, D.; Marin, A.C.; Fernández-Tomé, S.; Montalban-Arques, A.; Carrasco, A.; Tristán, E.; Ortega-Moreno, L.; Mora-Gutiérrez, I.; Díaz-Guerra, A.; Caminero-Fernández, R.; et al. Human intestinal pro-inflammatory CD11c<sup>high</sup>CCR2<sup>+</sup>CX3CR1<sup>+</sup> macrophages, but not their tolerogenic CD11c-CCR2-CX3CR1- counterparts, are expanded in inflammatory bowel disease article. *Mucosal Immunol.* 2018, 11, 1114–1126.
27. Isidro, R.A.; Appleyard, C.B. Colonic macrophage polarization in homeostasis, inflammation, and cancer. *Am. J. Physiol. Gastrointest. Liver Physiol.* 2016, 311, G59–G73.
28. Dharmani, P.; Srivastava, V.; Kisson-Singh, V.; Chadee, K. Role of intestinal mucins in innate host defense mechanisms against pathogens. *J. Innate Immun.* 2009, 1, 123–135.
29. Johansson, M.E.V.; Phillipson, M.; Petersson, J.; Velcich, A.; Holm, L.; Hansson, G.C.; Petersson, J.; Velcich, A.; Holm, L.; Hansson, G.C.; et al. The inner of the two Muc2 mucin-dependent mucus layers in colon is devoid of bacteria. *Proc. Natl. Acad. Sci. USA* 2008, 105, 15064–15069.
30. Rodriguez-Pineiro, A.M.; Bergstrom, J.H.; Ermund, A.; Gustafsson, J.K.; Schutte, A.; Johansson, M.E.V.; Hansson, G.C. Studies of mucus in mouse stomach, small intestine, and colon. II. Gastrointestinal mucus proteome reveals Muc2 and Muc5ac accompanied by a set of core proteins. *AJP Gastrointest. Liver Physiol.* 2013, 305, G348–G356.
31. Ambort, D.; Johansson, M.E.V.; Gustafsson, J.K.; Ermund, A.; Hansson, G.C. Perspectives on mucus properties and formation-lessons from the biochemical world. *Cold Spring Harb. Perspect. Med.* 2012, 2, 1–9.
32. Linden, S.K.; Sutton, P.; Karlsson, N.G.; Korolik, V.; McGuckin, M.A. Mucins in the mucosal barrier to infection. *Mucosal Immunol.* 2008, 1, 183–197.
33. Klomp, L.W.J.; Rens, L.V.A.N.; Stroust, G.J. Cloning and analysis of human gastric mucin cDNA reveals two types of conserved cysteine-rich domains. *Biochem. J.* 1995, 311, 831–838.
34. Corfield, A.P. Mucins: A biologically relevant glycan barrier in mucosal protection. *Biochim. Biophys. Acta Gen. Subj.* 2015, 1850, 236–252.
35. Ambort, D.; Johansson, M.E.V.; Gustafsson, J.K.; Nilsson, H.E.; Ermund, A. Calcium and pH-dependent packing and release of the gel-forming MUC2 mucin. *Proc. Natl. Acad. Sci. USA* 2012, 109, 5645–5650.
36. Desai, M.S.; Seekatz, A.M.; Koropatkin, N.M.; Kamada, N.; Hickey, C.A.; Wolter, M.; Pudlo, N.A.; Kitamoto, S.; Muller, A.; Young, V.B.; et al. A Dietary Fiber-Deprived Gut Microbiota Degrades the Colonic Mucus Barrier and Enhances Pathogen Susceptibility. *Cell* 2017, 167, 1339–1353.
37. Patel, K.K.; Miyoshi, H.; Beatty, W.L.; Head, R.D.; Malvin, N.P.; Cadwell, K.; Guan, J.; Saitoh, T.; Akira, S.; Seglen, P.O.; et al. Autophagy proteins control goblet cell function by potentiating reactive oxygen species production. *EMBO J.* 2013, 32, 3130–3144.
38. Wlodarska, M.; Thaiss, C.A.; Nowarski, R.; Henao-Mejia, J.; Zhang, J.P.; Brown, E.M.; Frankel, G.; Levy, M.; Katz, M.N.; Philbrick, W.M.; et al. NLRP6 inflammasome orchestrates the colonic host-microbial interface by regulating goblet cell mucus secretion. *Cell* 2014, 156, 1045–1059.

39. Paone, P.; Cani, P.D. Mucus barrier, mucins and gut microbiota: The expected slimy partners? *Gut* 2020, 69, 2232–2243.
40. Bergstrom, K.S.B.; Xia, L. Mucin-type O-glycans and their roles in intestinal homeostasis. *Glycobiology* 2013, 23, 1026–1037.
41. Wells, J.M.; Brummer, R.J.; Derrien, M.; MacDonald, T.T.; Troost, F.; Cani, P.D.; Theodorou, V.; Dekker, J.; Méheust, A.; de Vos, W.M.; et al. Homeostasis of the gut barrier and potential biomarkers. *Am. J. Physiol. Gastrointest. Liver Physiol.* 2017, 312, G171–G193.
42. Schroeder, B.O. Fight them or feed them: How the intestinal mucus layer manages the gut microbiota. *Gastroenterol. Rep.* 2019, 7, 3–12.
43. Cornick, S.; Tawiah, A.; Chadee, K. Roles and regulation of the mucus barrier in the gut. *Tissue Barriers* 2015, 3, 1–2.
44. Heazlewood, C.K.; Cook, M.C.; Eri, R.; Price, G.R.; Tauro, S.B.; Taupin, D.; Thornton, D.J.; Chin, W.P.; Crockford, T.L.; Cornall, R.J.; et al. Aberrant mucin assembly in mice causes endoplasmic reticulum stress and spontaneous inflammation resembling ulcerative colitis. *PLoS Med.* 2008, 5, 0440–0460.
45. Shkoda, A.; Ruiz, P.A.; Daniel, H.; Kim, S.C.; Rogler, G.; Sartor, R.B.; Haller, D. Interleukin-10 Blocked endoplasmic reticulum stress in intestinal epithelial cells: Impact on chronic inflammation. *Gastroenterology* 2007, 132, 190–207.
46. Hasnain, S.Z.; Tauro, S.; Das, I.; Tong, H.; Chen, A.C.H.; Jeffery, P.L.; McDonald, V.; Florin, T.H.; McGuckin, M.A. IL-10 Promotes production of intestinal mucus by suppressing protein misfolding and endoplasmic reticulum stress in goblet cells. *Gastroenterology* 2013, 144, 357–368.e9.
47. Bergstrom, K.S.B.; Kissoon-Singh, V.; Gibson, D.L.; Ma, C.; Montero, M.; Sham, H.P.; Ryz, N.; Huang, T.; Velcich, A.; Finlay, B.B.; et al. Muc2 protects against lethal infectious colitis by disassociating pathogenic and commensal bacteria from the colonic mucosa. *PLoS Pathog.* 2010, 6.
48. Hasnain, S.Z.; Wang, H.; Ghia, J.E.; Haq, N.; Deng, Y.; Velcich, A.; Grecnis, R.K.; Thornton, D.J.; Khan, W.I. Mucin gene deficiency in mice impairs host resistance to an enteric parasitic infection. *Gastroenterology* 2010, 138, 1763–1771.e5.
49. McDole, J.R.; Wheeler, L.W.; McDonald, K.G.; Wang, B.; Konjufca, V.; Knoop, K.A.; Newberry, R.D.; Miller, M.J. Goblet cells deliver luminal antigen to CD103+ dendritic cells in the small intestine. *Nature* 2012, 483, 345–349.
50. Hooper, L.V.; Midtvedt, T.; Gordon, J.I. How host-microbial interactions shape the nutrient environment of the mammalian intestine. *Annu. Rev. Nutr.* 2002, 22, 283–307.
51. Finnie, I.A.; Dwarakanath, A.D.; Taylor, B.A.; Rhodes, J.M. Colonic mucin synthesis is increased by sodium butyrate. *Gut* 1995, 36, 93–99.
52. Morita, H.; Kettlewell, M.G.W.; Jewell, D.P.; Kent, P.W. Glycosylation and sulphation of colonic mucus glycoproteins in patients with ulcerative colitis and in healthy subjects. *Gut* 1993, 34, 926–932.
53. Einerhand, A.W.C.; Renes, I.B.; Makkink, M.K.; Van Der Sluis, M.; Büller, H.A.; Dekker, J. Role of mucins in inflammatory bowel disease: Important lessons from experimental models. *Eur. J. Gastroenterol. Hepatol.* 2002, 14, 757–765.
54. Varsha, S.; Kelli, J.; Jianyi, Y.; Sun, L.; Ruxian, L.; Huimin, Y.; Julie, I.; Jennifer, F.-A.; Nicholas, Z.C.; Mark, D.; et al. Chronic inflammation in ulcerative colitis causes long term changes in goblet cell function. *Cell. Mol. Gastroenterol. Hepatol.* 2021, 18, 1–14.
55. Pullan, R.D.; Thomas, G.A.O.; Rhodes, M.; Newcombe, R.G.; Williams, G.T.; Allen, A.; Rhodes, J. Thickness of adherent mucus gel on colonic mucosa in humans and its relevance to colitis. *Gut* 1994, 35, 353–359.
56. Van Klinken, B.J.W.; Van Der Wal, J.W.G.; Einerhand, A.; Büller, H.A.; Dekker, J. Sulphation and secretion of the predominant secretory human colonic mucin MUC2 in ulcerative colitis. *Gut* 1999, 44, 387–393.
57. Larsson, J.M.H.; Karlsson, H.; Crespo, J.G.; Johansson, M.E.V.; Eklund, L.; Sjövall, H.; Hansson, G.C. Altered O-glycosylation profile of MUC2 mucin occurs in active ulcerative colitis and is associated with increased inflammation. *Inflamm. Bowel Dis.* 2011, 17, 2299–2307.
58. Shaoul, R.; Okada, Y.; Cutz, E.; Marcon, M.A. Colonic Expression of MUC2, MUC5AC, and TFF1 in Inflammatory Bowel Disease in Children. *J. Pediatr. Gastroenterol. Nutr.* 2004, 38, 488–493.
59. Fogue-Lafitte, M.E.; Fabiani, B.; Levy, P.P.; Maurin, N.; Flejou, J.F.; Bara, J. Abnormal expression of M1/MUC5AC mucin in distal colon of patients with diverticulitis, ulcerative colitis and cancer. *Int. J. Cancer* 2007, 121, 1543–1549.
60. Borralho, P.; Vieira, A.; Freitas, J.; Chaves, P.; Soares, J. Aberrant gastric apomucin expression in ulcerative colitis and associated neoplasia. *J. Crohns Colitis* 2007, 1, 35–40.

61. Olli, K.E.; Rapp, C.; Connell, L.O.; Collins, C.B.; McNamee, E.N.; Jensen, O.; Jedlicka, P.; Allison, K.C.; Goldberg, M.S.; Gerich, M.E.; et al. Muc5ac expression protects the colonic barrier in experimental colitis. *Inflamm. Bowel Dis.* 2020, 26, 1353–1367.
  62. Derrien, M.; Van Passel, M.W.J.; Van De Bovenkamp, J.H.B.; Schipper, R.G.; De Vos, W.M.; Dekker, J. Mucin-bacterial interactions in the human oral cavity and digestive tract. *Gut Microbes* 2010, 1, 254–268.
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