Nymphaea hybrid

Subjects: Biochemical Research Methods Contributor: Sheng-Nan Lei

Nymphaea hybrid is a perennial aquatic herb of *Nymphaea* in *Nymphaeaceae*. It is fragrant, colourful and indispensable as the main plant in features of water purification. Here includes the extraction from *Nymphaea hybrid*, isolation, structural characterization and evaluation of antioxidant capacity in vitro.

Nymphaea hybrid ultrasound-assisted cellulase

1. Introduction

Nymphaea hybrid is a perennial aquatic herb of *Nymphaea* in *Nymphaeaceae* ^[1]. It is fragrant, colorful and indispensable as the main plant in features of water purification ^{[2][3]}. In-depth study of flavonoids and polyphenols of *Nymphaea* has widespread application prospects in functional food, medicine and cosmetics industries. However, the studies about *Nymphaea hybrid* have been focused on the aromatic components for fragrance application, while there are only a few studies about flavonoids and polyphenols and other active ingredients.

2. Extraction of active substances

Ultrasonic-assisted cellulase extraction (UCE) was applied to extract flavonoids and polyphenols from the *Nymphaea hybrid* flower. The extraction conditions were optimized using the response surface method (RSM) coupled with a Box-Behnken design. According to the response surface experiment, the optimal process conditions for extraction were as follows: the liquid-to-solid ratio was 40.45:1 (mL/g), cellulase amount was 5.23% (*w/w*), the ethanol concentration was 52% (*v/v*). Under such conditions, the predicted flavonoid of NHE was 26.92 mg/g. To verify these optimal conditions, the experiment was repeated five times and the average actual flavonoid content of NHE was 27.26 \pm 0.64 mg/g (*n* = 5), which was close to the predicted value and proved the validity of the models.

3. AB-8 Macroporous Resin

NHEP was prepared by the purification of NHE with AB-8 macroporous resin. As shown in Table 1, the content of flavonoids and polyphenols in the NHEP sample was higher than in the NHE sample with the same dry weight. After purification, the contents of flavonoids and polyphenols in NHEP increased from 55.60 mg/g to 157.39 mg/g and from 414.97 mg/g to 888.63 mg/g, respectively. This result showed that it was efficient for the enrichment of flavonoids and polyphenols of NHE by AB-8 macroporous resin.

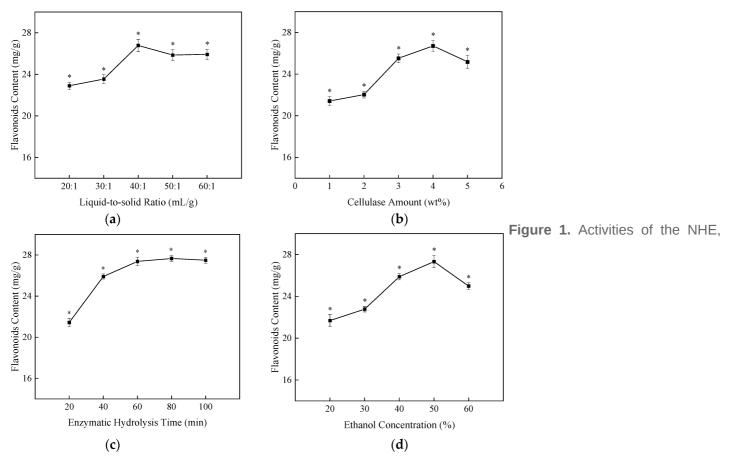
Table 1. The content of flavonoids and polyphenols in the NHE and NHEP with the same dry weight.

Sample	Compound			
	Flavonoids Content (mg/g)	Polyphenols Content (mg/g)		
NHE	55.60 ± 0.38	414.97 ± 2.95		
NHEP	157.39 ± 1.18 *	888.63 ± 4.41 *		

The data shows as the mean \pm S.D. (n = 3). *: p < 0.05 compared to NHE.

4. Activity Determination by Chemical Method

To explore in vitro antioxidant activity of NHE and NHEP, the free radical scavenging abilities against ABTS, DPPH, and reduction power, were detected compared with VC. As shown in Figure 1, the IC₅₀ values of the DPPH radical of NHE, NHEP and VC were 11.09 \pm 0.15 µg/mL, 3.28 \pm 0.03 µg/mL and 5.18 \pm 0.06 µg/mL, respectively. The IC₅₀ values of the ABTS radical of NHE, NHEP and VC were 18.21 \pm 0.05 µg/mL, 10.50 \pm 0.10 µg/mL and 7.77 \pm 0.04 µg/mL, respectively. The reduction power of NHE, NHEP, VC for Fe³⁺ were strong with high absorbances ^[4]. The absorbances were 2.20 \pm 0.04, 2.36 \pm 0.04 and 2.60 \pm 0.07 in the 1 mg/mL concentration, respectively. This result indicated that the scavenging free radical abilities and reduction power of NHEP were similar to VC and stronger than that of NHE. NHE and NHEP were also evaluated for the anti-inflammatory activity by using hyaluronidase inhibition assays. The IC₅₀ value of NHE and NHEP in hyaluronidase inhibition assay was 0.55 \pm 0.01 mg/mL and 0.32 \pm 0.01 mg/mL. The inhibition rate of 10 µg/mL dipotassium glycyrrhizinate (Dg) was 94.23 \pm 1.72%. This result showed that both NHE and NHEP display an anti-inflammatory effect, and NHEP has a stronger anti-inflammatory effect than NHE. Hence, in vitro bioactivity evaluation by chemical assays showed that both NHE and NHEP displayed strong free radical scavenging abilities against the DPPH and ABTS radicals, good reduction power, and anti-inflammatory effect.



NHEP and positive control by chemical method. (a) DPPH, (b) ABTS, (c) Reduction power, (d) Hyaluronidase inhibition assay. Dg stands for Dipotassium glycyrrhizinate, and the concentration is 10 μ g/mL. The date is shown as the mean ± S.D. (*n* = 3).

5. Characterization of FTIR and HPLC

Infrared spectroscopy analysis can be used as a powerful tool to characterize and identify functional groups present in the compounds. As shown in Figure 2, the absorption peak at the vicinity of 3424 cm⁻¹ is relatively wide and strong, which is attributed to the stretching vibration of O–H. The signal in wavelength of 2930 cm⁻¹ is assigned to the absorption peak of stretching vibration of the C–H bond of aliphatic hydrocarbon. The band at 3000-3100 cm⁻¹ is due to the stretching vibration of C–H of the aromatic ring. The aromatic C–H bond was out-of-plane bending vibration, resulting in absorption peaks at 834 and 760 cm⁻¹ wavelengths ^[5]. A characteristic absorption peak at 1718 cm⁻¹ may be assigned to the carboxylic acid of a sharp stretching band of the C=O. The peaks at 1613 and 1449 cm⁻¹ wavelength belong to the C=C bond of the aromatic ring vibrates ^[6]. The absorption peaks at 1512 cm⁻¹ may be assigned to the aromatic skeletal vibrations, ring breathing with C–O starching vibration ^[5]. The adsorption peaks at 1449 and 1349 cm⁻¹ are assigned to the bending vibrations of –CH₃ and –CH₂. The peaks at 1210 and 1038 cm⁻¹ wavelength belong to the stretching vibration of C–H and C–O bonds ^[6]. The characteristic peaks of the benzene ring, C–O stretching vibration and stretching vibration of O–H, indicate the presence of phenolic hydroxy groups ^[7].

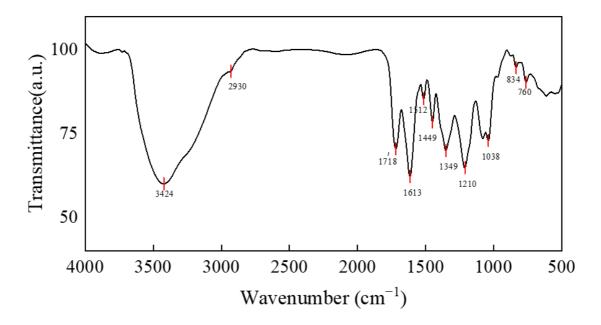


Figure 2. FTIR-NHEP illustrating structural features.

The total polyphenol and flavonoid compositions of NHEP were identified by the characterization of HPLC in Figure 3 and Table 2. This result showed some of the main NHEP bioactive phytochemicals, including gallic acid (1.60 \pm 0.36 mg/g), corilagin (65.60 \pm 3.36 mg/g), ellagic acid (97.72 \pm 0.92 mg/g), rutin (34.41 \pm 4.24 mg/g), myricetin (3.13 \pm 0.87 mg/g), quercetin (4.64 \pm 0.15 mg/g), naringin (4.03 \pm 0.24 mg/g) etc. The contents of corilagin, ellagic acid and rutin in NHEP were higher among these components. These standard substances have been reported to have excellent antioxidant activity, which further indicated that NHEP has great potential as a natural antioxidant. Uddin et al. identified 9 bioactive phytochemicals from methanolic extract of *Nymphaea nouchali* using an HPLC system ^[8]. Among these compounds, rutin (39.44 mg/g), myricetin (30.77 mg/g), ellagic acid (11.05 mg/g), gallic acid (5.33 mg/g) and quercetin (0.90 mg/g) were polyphenols or flavonoids, which were also identified as bioactive ingredients of NHEP in this entry. The researchers found the ellagic acid content (97.72 \pm 0.92 mg/g) of NHEP was much greater than that (11.05 mg/g) of *Nymphaea nouchali* and that (6.16 \pm 0.019 mg/g) of ethanolic extract of Water Lily (*Nymphaea Tetragona Georgi*) ^[9].

Standard	Retention Time (min)	Regression Equation	R ²	NHEP Compound (mg/g)
Gallic acid	5.33	y = 28.696 x - 6.5794	0.9997	1.60 ± 0.36

Table 2. Polyphenols and flavonoids compositions of NHEP by the characterization of HPLC.

Corilagin	16.21	y = 14.902 x - 40.435	0.9992	65.60 ± 3.36
Ellagic acid	30.58	y = 10.3 x - 11.978	0.9999	97.72 ± 0.92
Rutin	31.32	y = 10.71 x - 9.2865	0.9999	34.41 ± 4.24
Myricetin	40.86	y = 12.76 x + 4.3092	0.9996	3.13 ± 0.87
Quercetin	47.15	y = 11.001 x + 2.6758	0.9999	4.64 ± 0.15
Naringin	50.69	y = 34.715 x + 24.424	0.9999	4.03 ± 0.24

Y = The peak area (mAU*s); X = The concentration of standards (μ g/mL); R² = Coefficients of determination. The data shows as the mean ± S.D. (*n* = 3).

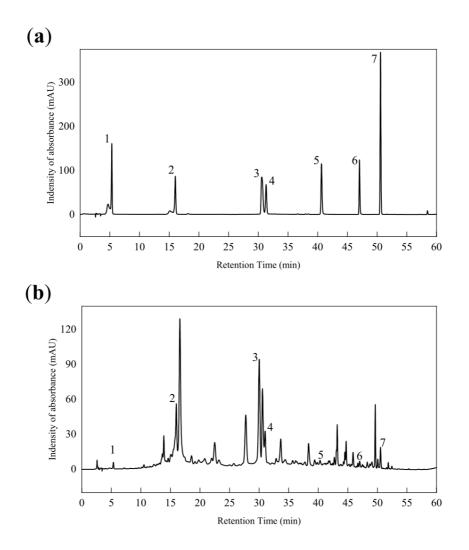


Figure 3. HPLC of mixed standards (**a**) and NHEP (**b**). 1-gallic acid, 2-corilagin, 3-ellagic acid, 4-rutin, 5-myricetin, 6-quercetin, 7-naringin.

6. HaCaT Cells and B16 Cells Experiments

As shown in Figure 4-7, the cell viability by CCK-8 assays showed that NHEP had no significant cytotoxicity for B16 and HaCaT cells when the concentration was below 100 μ g/mL and 120 μ g/mL, respectively. NHEP with a concentration of 20-160 μ g/mL can more effectively reduce the ROS level in H₂O₂ damaged HaCaT cells compared with 10 μ g/mL of VC. The 40 μ g/mL of NHEP had similar activity against intracellular melanin production in the B16 melanoma cells compared with 20 μ g/mL Kojic acid. Good activities of antioxidation, whitening and protective effect against H₂O₂-induced oxidative damage promote the potential for NHEP as a functional raw material in the field of cosmetics and medicine.

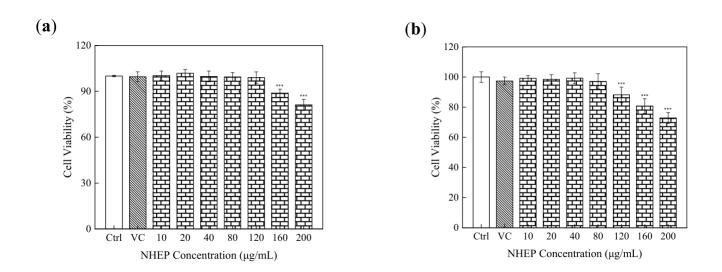
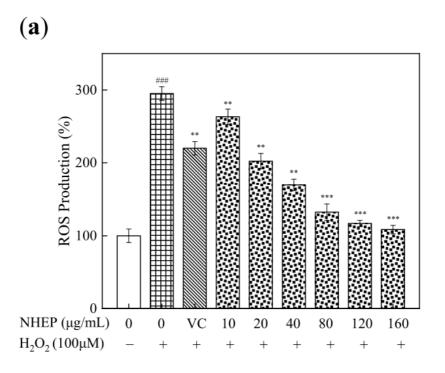


Figure 4. Cytotoxicity of NHEP on HaCaT cells. (**a**) cell viability of the HaCaT cells cultured with the NHEP for 24 h, (**b**) cell viability of the HaCaT cells cultured with the NHEP for 48 h. VC stands for Vitamin C, and the concentration is 10 μ g/mL. Significance: compared with the control group, *** *p* < 0.001. The data shows as the mean ± S.D. (*n* = 5).



(b)

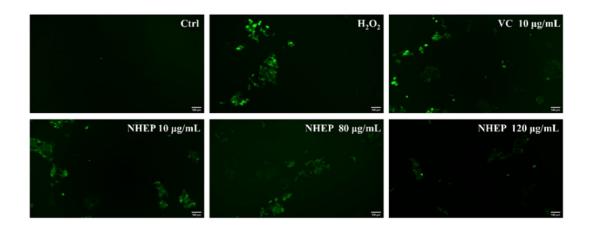


Figure 5. Effects of NHEP on the ROS level of HaCaT cells with stimulation of H_2O_2 : (a) ROS production, (b) fluorescence intensity by detected the DCFH-DA. VC stands for Vitamin C, and the concentration is 10 µg/mL (scale bar is 100 µm). Significance: ### p < 0.001 compared with the control group. ** p < 0.01, *** p < 0.001 compared with the H₂O₂ stimulation group. The data are shown as the mean ± S.D. (n = 5).

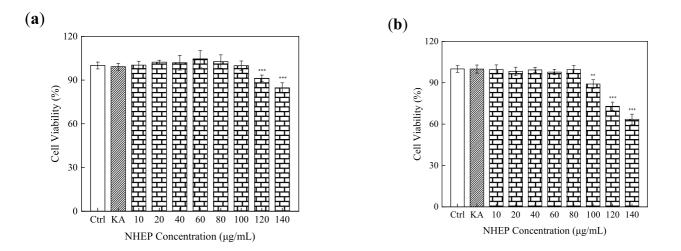


Figure 6. Cytotoxicity of NHEP on B16 cells: (**a**) viability of the B16 cells cultured with the NHEP for 24 h, (**b**) viability of the B16 cells cultured with the NHEP for 48 h. Significance: compared with the control group, ** p < 0.01, *** p < 0.0001. The data shows as the mean ± S.D. (n = 5). KA stands for Kojic acid, and the concentration is 20 µg/mL.

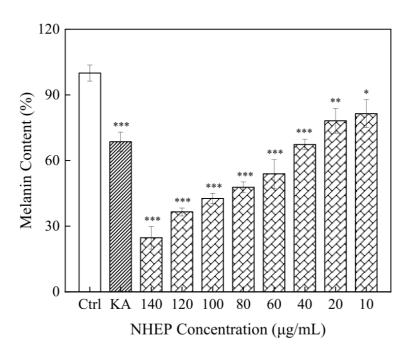


Figure 7. Effects of the NHEP on B16 cells melanin production. Significance: compared with the control group, * p < 0.05, ** p < 0.01, *** p < 0.001. The data shows as the mean ± S.D. (n = 5). KA stands for Kojic acid, and the concentration is 20 µg/mL.

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