

Microbe-Associated Bone Cell Differentiation

Subjects: Biochemistry & Molecular Biology

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Gut microbiota has emerged as an important regulator of bone homeostasis. In particular, the modulation of innate immunity and bone homeostasis is mediated through the interaction between microbe-associated molecular patterns (MAMPs) and the host pattern recognition receptors including Toll-like receptors and nucleotide-binding oligomerization domains. Pathogenic bacteria such as *Porphyromonas gingivalis* and *Staphylococcus aureus* tend to induce bone destruction and cause various inflammatory bone diseases including periodontal diseases, osteomyelitis, and septic arthritis. On the other hand, probiotic bacteria such as *Lactobacillus* and *Bifidobacterium* species can prevent bone loss. In addition, bacterial metabolites and various secretory molecules such as short chain fatty acids and cyclic nucleotides can also affect bone homeostasis.

Keywords: bone diseases ; bone homeostasis ; bacteria ; microbe-associated molecular patterns ; osteoblast ; osteoclast ; pattern-recognition receptors ; secretory microbial molecules

1. Introduction

The bone remodeling process is regulated by representative bone cells known as osteoclasts and osteoblasts [1]. The balance between bone-resorbing osteoclasts and bone-forming osteoblasts is essential for maintaining bone homeostasis [2]. However, imbalance between bone resorption and formation could lead to bone diseases [3]. Excessive osteoclast activity causes various bone diseases including osteoporosis, septic arthritis, osteomyelitis, and alveolar bone loss in periodontal diseases [4][5][6]. Especially, bacterial infections can directly affect bone homeostasis by increasing osteoclast differentiation and activation and/or decreasing osteoblast differentiation and activation [7]. For example, *Streptococcus pyogenes*, *Staphylococcus aureus*, and *Neisseria gonorrhoeae* are commonly found in patients with septic arthritis, resulting in cartilage and bone destruction within the joint [8]. *Staphylococcus* species such as *S. aureus* and *Staphylococcus epidermidis* are etiological agents of osteomyelitis [9]. Major oral pathogens, including *Porphyromonas gingivalis* and *Fusobacterium nucleatum*, are associated with periodontal diseases, manifesting alveolar bone loss [9]. However, unlike those pathogens, probiotics which are microorganisms that offer health benefits to the hosts are known to increase mineral density and volume of the bone [10]. For instance, *Lactobacillus reuteri* and *Lactobacillus rhamnosus* GG upregulate bone volume of mice [11][12]. In addition, other probiotics such as *Lactobacillus gasseri* and *Lactobacillus brevis* reduce bone loss and inflammation in mouse periodontitis model [13][14].

Bacteria have unique structural components called microbe-associated molecular patterns (MAMPs) including lipopolysaccharide (LPS), lipoteichoic acid (LTA), lipoprotein (LPP), and peptidoglycan (PGN) [15]. The recognition of MAMPs by pattern recognition receptors (PRRs) is crucial for inducing host immune responses [15]. In addition, secretory microbial molecules including short chain fatty acid (SCFA), extracellular vesicle (EV), extracellular polysaccharide, and cyclic dinucleotide (CDN) also modulate bone cells [16][17][18]. Therefore, for a clear understanding of the regulation of bone metabolism by bacteria, it is essential to understand the effects of MAMPs and secretory microbial molecules on bone cells and their regulatory mechanism.

2. Microbe-Associated Molecular Patterns

MAMPs are structural or secretory molecules that are highly conserved in most microbes [19]. Well-known MAMPs are bacterial polysaccharides (LPS and LTA), surface proteins (LPP and adhesin), PGNs, and secretory molecules (SCFA, EV, extracellular polysaccharide, and CDN) [20]. These MAMPs can be sensed by various host PRRs, such as Toll-like receptors (TLRs) and nucleotide-binding oligomerization domain (NOD)-like receptors (NLRs), or G-protein coupled receptors (GPCRs) [21][22]. Indeed, there are many host PRRs classified according to their location, function, and ligand specificity [23]. There are typically four types of PRRs: TLRs, NLRs, C-type lectin receptors, and RIG-1 like receptors [21]. Among these, TLRs localized at plasma membrane or in endosomes and NLRs localized in cytoplasm are the major PRRs in recognizing bacterial MAMPs [21]. For instance, TLR4 senses LPS, and TLR2 senses LPP and LTA [24]. On the

other hand, NOD1 and NOD2 recognize bacterial PGNs through their distinct structural moieties, d-glutamyl-meso-diaminopimelic acid (iE-DAP) and muramyl dipeptide (MDP), respectively [25]. Based on their displayed patterns, each host receptor responds to its specific bacterial ligand, subsequently producing anti- or pro-inflammatory cytokines and chemokines to counteract against invading microbes [26]. It has been reported that pathogens or probiotics and their MAMPs could also affect osteoimmunological responses (Table 1) [27]. Therefore, we will focus on MAMPs and their effects on bone homeostasis in this section.

Table 1. Effects of cell wall components on bone metabolism.

| MAMPs | Receptor | Effects | References |
|--------------------|----------|--|------------------------------|
| Lipopolysaccharide | TLR4 | Inducing bone loss | |
| | | Inhibiting osteoclastogenesis on macrophages | [28][29][30][31][32][33][34] |
| | | Facilitating osteoclast differentiation on committed osteoclasts | [35] |
| Lipoteichoic acid | TLR2 | Downregulating osteoblast differentiation | |
| | | Healing femoral fractures in mice | |
| | | Attenuating osteoclast differentiation and activating phagocytosis | [36][37][38][39][40] |
| Lipoprotein | TLR2 | Upregulating osteogenic markers and osteoblastogenesis | |
| | | Promoting bone resorption | |
| | | Upregulating osteoclast differentiation | [41][42][43] |
| Fimbria | TLR4 | Stimulating osteoblasts to elevate RANKL/OPG ratio | |
| | | Inducing osteoclastogenesis and bone resorption | [44][45][46][47][48][49] |
| | | Enhancing osteoclastogenesis and bone resorption | [50][51][52][53][54][55][56] |
| Peptidoglycan | NOD1 | Triggering osteoclast differentiation synergistically with LPS | |
| | | Upregulation of bone density | |
| | | Facilitating osteoblast differentiation | [54][57] |
| NOD2 | | Diminishing osteoclastogenesis by reducing RANKL/OPG ratio | |

3. Therapeutics

Microbes influence bone metabolism by constant interaction with host using their various MAMPs (Table 2) [2]. In infectious condition, MAMPs often trigger immoderate osteoclastogenesis or inhibit osteoblast differentiation through the activation of immune responses, causing bone diseases such as osteomyelitis, osteoporosis, and periodontitis [2]. Antibiotics are commonly used to treat MAMP-induced bone diseases in bacterial infection [58]. Nevertheless, the emergence of antibiotic-resistant bacteria and remaining MAMPs after treatment pose significant challenge for complete clearance [59]. Therefore, further studies are needed to understand the role of MAMPs in bone diseases and to control the immune responses induced by MAMPs.

Table 2. Effects of secretory microbial molecules on bone metabolism.

| MAMPs | Mechanism | Effects on Bone Metabolism | References |
|-------------------------------|---|---|------------------|
| Short chain fatty acids | Activation of GPCRs Inhibition of histone deacetylases | Inhibited osteoclast differentiation and function Upregulated osteogenic factors in low dose Attenuated osteoblast differentiation and mineralization | [60][61][62][63] |
| | | Prevented bone loss in various mouse models | |
| Extracellular vesicles | Activation of TLR2 Induction of pro-inflammatory cytokines | Downregulated osteoblast differentiation and activity Regulated RANKL and OPG expression in mesenchymal cells | [17] |
| Extracellular polysaccharides | Activation of TLR2 | Inhibited osteoclast differentiation from macrophages, but some EPS increased collagenolytic activity of osteoclasts Enhanced osteoblast differentiation, but oral pathogen-derived CPS decreased proliferation of osteoblasts | [64][65][66][67] |
| Cyclic dinucleotides | Induction of STING-mediated IFN-β | Inhibited differentiation of macrophages into mature osteoclasts Alleviated RANKL-induced bone destruction | [18] |

On the other hand, several studies investigated that some MAMPs, especially derived from probiotics, decrease bone resorption or enhance bone formation by controlling the differentiation of osteoclasts or osteoblasts, respectively, in both *in vitro* and *in vivo* studies [18][57][63]. Many therapeutic drugs, such as bisphosphonates, monoclonal antibodies, or hormone preparations, are traditionally developed to treat bone diseases by inhibiting bone resorption or inducing bone formation [68][69][70]. However, conventional drugs show unexpected side effects, such as nausea or osteonecrosis of jaw [70][71][72]. Therefore, we suggest that probiotic-derived MAMPs could alternatively be used in place of conventional therapies. To evaluate their therapeutic use, we have discussed below how to treat MAMP-induced bone diseases and how to exploit MAMPs in bone health.

References

- Kular, J.; Tickner, J.; Chim, S.M.; Xu, J. An overview of the regulation of bone remodelling at the cellular level. *Clin. Biochem.* 2012, 45, 863–873.
- Robling, A.G.; Castillo, A.B.; Turner, C.H. Biomechanical and molecular regulation of bone remodeling. *Annu. Rev. Bio med. Eng.* 2006, 8, 455–498.
- Feng, X.; McDonald, J.M. Disorders of bone remodeling. *Annu. Rev. Pathol.* 2011, 6, 121–145.
- Krauss, J.L.; Roper, P.M.; Ballard, A.; Shih, C.C.; Fitzpatrick, J.A.J.; Cassat, J.E.; Ng, P.Y.; Pavlos, N.J.; Veis, D.J. *Staphylococcus aureus* Infects Osteoclasts and Replicates Intracellularly. *mBio* 2019, 10.
- Wright, J.A.; Nair, S.P. Interaction of staphylococci with bone. *Int. J. Med. Microbiol.* 2010, 300, 193–204.
- Martin, T.R.; Mathison, J.C.; Tobias, P.S.; Leturcq, D.J.; Moriarty, A.M.; Maunder, R.J.; Ulevitch, R.J. Lipopolysaccharide binding protein enhances the responsiveness of alveolar macrophages to bacterial lipopolysaccharide. Implications for cytokine production in normal and injured lungs. *J. Clin. Investig.* 1992, 90, 2209–2219.
- Charles, J.F.; Nakamura, M.C. Bone and the innate immune system. *Curr. Osteoporos. Rep.* 2014, 12, 1–8.
- Sakurai, A.; Okahashi, N.; Nakagawa, I.; Kawabata, S.; Amano, A.; Ooshima, T.; Hamada, S. *Streptococcus pyogenes* infection induces septic arthritis with increased production of the receptor activator of the NF-κappaB ligand. *Infect. Imm*

9. Binder Gallimidi, A.; Fischman, S.; Revach, B.; Bulvik, R.; Maliutina, A.; Rubinstein, A.M.; Nussbaum, G.; Elkin, M. Peri odontal pathogens *Porphyromonas gingivalis* and *Fusobacterium nucleatum* promote tumor progression in an oral-spe cific chemical carcinogenesis model. *Oncotarget* 2015, 6, 22613–22623.
10. Parvaneh, K.; Jamaluddin, R.; Karimi, G.; Erfani, R. Effect of probiotics supplementation on bone mineral content and b one mass density. *Sci. World J.* 2014, 2014, 595962.
11. Li, J.Y.; Chassaing, B.; Tyagi, A.M.; Vaccaro, C.; Luo, T.; Adams, J.; Darby, T.M.; Weitzmann, M.N.; Mulle, J.G.; Gewirtz, A.T.; et al. Sex steroid deficiency-associated bone loss is microbiota dependent and prevented by probiotics. *J. Clin. In vestig.* 2016, 126, 2049–2063.
12. McCabe, L.R.; Irwin, R.; Schaefer, L.; Britton, R.A. Probiotic use decreases intestinal inflammation and increases bone density in healthy male but not female mice. *J. Cell. Physiol.* 2013, 228, 1793–1798.
13. Kobayashi, R.; Kobayashi, T.; Sakai, F.; Hosoya, T.; Yamamoto, M.; Kurita-Ochiai, T. Oral administration of *Lactobacillus gasseri* SBT2055 is effective in preventing *Porphyromonas gingivalis*-accelerated periodontal disease. *Sci. Rep.* 201 7, 7, 545.
14. Maekawa, T.; Hajishengallis, G. Topical treatment with probiotic *Lactobacillus brevis* CD2 inhibits experimental periodo ntal inflammation and bone loss. *J. Periodontal Res.* 2014, 49, 785–791.
15. Chu, H.; Mazmanian, S.K. Innate immune recognition of the microbiota promotes host-microbial symbiosis. *Nat. Immun ol.* 2013, 14, 668–675.
16. Iwami, K.; Moriyama, T. Effects of short chain fatty acid, sodium butyrate, on osteoblastic cells and osteoclastic cells. *In t. J. Biochem.* 1993, 25, 1631–1635.
17. Song, M.K.; Kim, H.Y.; Choi, B.K.; Kim, H.H. Filifactor alocis-derived extracellular vesicles inhibit osteogenesis through TLR2 signaling. *Mol. Oral Microbiol.* 2020, 35, 202–210.
18. Kwon, Y.; Park, O.J.; Kim, J.; Cho, J.H.; Yun, C.H.; Han, S.H. Cyclic Dinucleotides Inhibit Osteoclast Differentiation Thr ough STING-Mediated Interferon-beta Signaling. *J. Bone Miner. Res.* 2019, 34, 1366–1375.
19. Boller, T.; Felix, G. A renaissance of elicitors: Perception of microbe-associated molecular patterns and danger signals by pattern-recognition receptors. *Annu. Rev. Plant Biol.* 2009, 60, 379–406.
20. Choi, H.W.; Klessig, D.F. DAMPs, MAMPs, and NAMPs in plant innate immunity. *BMC Plant Biol.* 2016, 16, 232.
21. Akira, S.; Uematsu, S.; Takeuchi, O. Pathogen recognition and innate immunity. *Cell* 2006, 124, 783–801.
22. Sun, M.; Wu, W.; Liu, Z.; Cong, Y. Microbiota metabolite short chain fatty acids, GPCR, and inflammatory bowel diseas es. *J. Gastroenterol.* 2017, 52, 1–8.
23. Brubaker, S.W.; Bonham, K.S.; Zanoni, I.; Kagan, J.C. Innate immune pattern recognition: A cell biological perspective. *Annu. Rev. Immunol.* 2015, 33, 257–290.
24. Kawai, T.; Akira, S. TLR signaling. *Semin. Immunol.* 2007, 19, 24–32.
25. Platnich, J.M.; Muruve, D.A. NOD-like receptors and inflammasomes: A review of their canonical and non-canonical sig naling pathways. *Arch. Biochem. Biophys.* 2019, 670, 4–14.
26. Chatterjee, S.; Jungraithmayr, W.; Bagchi, D. Immunity and Inflammation in Health and Disease: Emerging Roles of Nu traceuticals and Functional Foods in Immune Support; Academic Press: London, UK, 2018; pp. 175–187.
27. D'Amelio, P.; Sassi, F. Gut Microbiota, Immune System, and Bone. *Calcif. Tissue Int.* 2018, 102, 415–425.
28. Chen, M.F.; Chang, C.H.; Hu, C.C.; Wu, Y.Y.; Chang, Y.; Ueng, S.W.N. Periprosthetic Joint Infection Caused by Gram-P ositive Versus Gram-Negative Bacteria: Lipopolysaccharide, but not Lipoteichoic Acid, Exerts Adverse Osteoclast-Medi ated Effects on the Bone. *J. Clin. Med.* 2019, 8, 1289.
29. Ridwan, R.D.; Sidarningsih, T.K.; Salim, S. Effect of lipopolysaccharide derived from surabaya isolates of *Actinobacillus actinomycetemcomitans* on alveolar bone destruction. *Vet. World* 2018, 11, 161–166.
30. Nishihara, T.; Ishihara, Y.; Koseki, T.; Boutsi, E.A.; Senpuku, H.; Hanada, N. Membrane-associated interleukin-1 on ma crophages stimulated with *Actinobacillus actinomycetemcomitans* lipopolysaccharide induces osteoclastic bone resorpti on in vivo. *Cytobios* 1995, 81, 229–237.
31. Zou, W.; Bar-Shavit, Z. Dual modulation of osteoclast differentiation by lipopolysaccharide. *J. Bone Miner. Res.* 2002, 1 7, 1211–1218.
32. Liu, J.; Wang, S.; Zhang, P.; Said-Al-Naief, N.; Michalek, S.M.; Feng, X. Molecular mechanism of the bifunctional role of lipopolysaccharide in osteoclastogenesis. *J. Biol. Chem.* 2009, 284, 12512–12523.

33. Kadono, H.; Kido, J.; Kataoka, M.; Yamauchi, N.; Nagata, T. Inhibition of osteoblastic cell differentiation by lipopolysaccharide extract from *Porphyromonas gingivalis*. *Infect. Immun.* 1999, 67, 2841–2846.
34. Tomomatsu, N.; Aoki, K.; Alles, N.; Soysa, N.S.; Hussain, A.; Nakachi, H.; Kita, S.; Shimokawa, H.; Ohya, K.; Amagasa, T. LPS-induced inhibition of osteogenesis is TNF-alpha dependent in a murine tooth extraction model. *J. Bone Miner. Res.* 2009, 24, 1770–1781.
35. Bandow, K.; Maeda, A.; Kakimoto, K.; Kusuyama, J.; Shamoto, M.; Ohnishi, T.; Matsuguchi, T. Molecular mechanisms of the inhibitory effect of lipopolysaccharide (LPS) on osteoblast differentiation. *Biochem. Biophys. Res. Commun.* 2010, 402, 755–761.
36. Yang, J.; Park, O.J.; Kim, J.; Baik, J.E.; Yun, C.H.; Han, S.H. Lipoteichoic Acid of *Enterococcus faecalis* Inhibits the Differentiation of Macrophages into Osteoclasts. *J. Endod.* 2016, 42, 570–574.
37. Wang, S.; Heng, B.C.; Qiu, S.; Deng, J.; Shun Pan Cheung, G.; Jin, L.; Zhao, B.; Zhang, C. Lipoteichoic acid of *Enterococcus faecalis* inhibits osteoclastogenesis via transcription factor RBP-J. *Innate Immun.* 2019, 25, 13–21.
38. Yang, J.; Ryu, Y.H.; Yun, C.H.; Han, S.H. Impaired osteoclastogenesis by staphylococcal lipoteichoic acid through Toll-like receptor 2 with partial involvement of MyD88. *J. Leukoc. Biol.* 2009, 86, 823–831.
39. Liu, X.; Wang, Y.; Cao, Z.; Dou, C.; Bai, Y.; Liu, C.; Dong, S.; Fei, J. Staphylococcal lipoteichoic acid promotes osteogenic differentiation of mouse mesenchymal stem cells by increasing autophagic activity. *Biochem. Biophys. Res. Commun.* 2017, 485, 421–426.
40. Hu, C.C.; Chang, C.H.; Hsiao, Y.M.; Chang, Y.; Wu, Y.Y.; Ueng, S.W.N.; Chen, M.F. Lipoteichoic Acid Accelerates Bone Healing by Enhancing Osteoblast Differentiation and Inhibiting Osteoclast Activation in a Mouse Model of Femoral Defects. *Int. J. Mol. Sci.* 2020, 21, 5550.
41. Kim, J.; Yang, J.; Park, O.J.; Kang, S.S.; Kim, W.S.; Kurokawa, K.; Yun, C.H.; Kim, H.H.; Lee, B.L.; Han, S.H. Lipoproteins are an important bacterial component responsible for bone destruction through the induction of osteoclast differentiation and activation. *J. Bone Miner. Res.* 2013, 28, 2381–2391.
42. Sato, N.; Takahashi, N.; Suda, K.; Nakamura, M.; Yamaki, M.; Ninomiya, T.; Kobayashi, Y.; Takada, H.; Shibata, K.; Yamamoto, M.; et al. MyD88 but not TRIF is essential for osteoclastogenesis induced by lipopolysaccharide, diacyl lipopeptide, and IL-1alpha. *J. Exp. Med.* 2004, 200, 601–611.
43. Souza, J.A.C.; Magalhaes, F.A.C.; Oliveira, G.; RS, D.E.M.; Zuanon, J.A.; Souza, P.P.C. Pam2CSK4 (TLR2 agonist) induces periodontal destruction in mice. *Braz. Oral Res.* 2020, 34, e012.
44. Sasaki, H.; Watanabe, K.; Toyama, T.; Koyata, Y.; Hamada, N. *Porphyromonas gingivalis* 41-kDa fimbriae induced osteoclast differentiation and cytokine production. *J. Vet. Med. Sci.* 2015, 77, 265–271.
45. Hiramine, H.; Watanabe, K.; Hamada, N.; Umemoto, T. *Porphyromonas gingivalis* 67-kDa fimbriae induced cytokine production and osteoclast differentiation utilizing TLR2. *FEMS Microbiol. Lett.* 2003, 229, 49–55.
46. Kawata, Y.; Hanazawa, S.; Amano, S.; Murakami, Y.; Matsumoto, T.; Nishida, K.; Kitano, S. *Porphyromonas gingivalis* fimbriae stimulate bone resorption in vitro. *Infect. Immun.* 1994, 62, 3012–3016.
47. Hanazawa, S.; Kawata, Y.; Murakami, Y.; Naganuma, K.; Amano, S.; Miyata, Y.; Kitano, S. *Porphyromonas gingivalis* fimbria-stimulated bone resorption in vitro is inhibited by a tyrosine kinase inhibitor. *Infect. Immun.* 1995, 63, 2374–2377.
48. Zhang, W.; Ju, J.; Rigney, T.; Tribble, G.D. Fimbriae of *Porphyromonas gingivalis* are important for initial invasion of osteoblasts, but not for inhibition of their differentiation and mineralization. *J. Periodontol.* 2011, 82, 909–916.
49. Zhang, W.; Ju, J.; Rigney, T.; Tribble, G. Integrin alpha5beta1-fimbriae binding and actin rearrangement are essential for *Porphyromonas gingivalis* invasion of osteoblasts and subsequent activation of the JNK pathway. *BMC Microbiol.* 2013, 13, 5.
50. Kishimoto, T.; Kaneko, T.; Ukai, T.; Yokoyama, M.; Ayon Haro, R.; Yoshinaga, Y.; Yoshimura, A.; Hara, Y. Peptidoglycan and lipopolysaccharide synergistically enhance bone resorption and osteoclastogenesis. *J. Periodontal Res.* 2012, 47, 446–454.
51. Ozaki, Y.; Kishimoto, T.; Yamashita, Y.; Kaneko, T.; Higuchi, K.; Mae, M.; Oohira, M.; Mohammad, A.I.; Yanagiguchi, K.; Yoshimura, A. Expression of osteoclastogenic and anti-osteoclastogenic cytokines differs in mouse gingiva injected with lipopolysaccharide, peptidoglycan, or both. *Arch. Oral Biol.* 2021, 122, 104990.
52. Ishida, M.; Kitaura, H.; Kimura, K.; Sugisawa, H.; Aonuma, T.; Takada, H.; Takano-Yamamoto, T. Muramyl dipeptide enhances lipopolysaccharide-induced osteoclast formation and bone resorption through increased RANKL expression in stromal cells. *J. Immunol. Res.* 2015, 2015, 132765.
53. Sato, T.; Watanabe, K.; Kumada, H.; Toyama, T.; Tani-Ishii, N.; Hamada, N. Peptidoglycan of *Actinomyces naeslundii* induces inflammatory cytokine production and stimulates osteoclastogenesis in alveolar bone resorption. *Arch. Oral Biol.*

54. Chaves de Souza, J.A.; Frasnelli, S.C.; Curylofo-Zotti, F.A.; Avila-Campos, M.J.; Spolidorio, L.C.; Zamboni, D.S.; Graves, D.T.; Rossa, C., Jr. NOD1 in the modulation of host-microbe interactions and inflammatory bone resorption in the peri-odontal disease model. *Immunology* 2016, 149, 374–385.
55. Kitaura, H.; Ishida, M.; Kimura, K.; Sugisawa, H.; Kishikawa, A.; Shima, K.; Ogawa, S.; Qi, J.; Shen, W.R. Role of Muramyl Dipeptide in Lipopolysaccharide-Mediated Biological Activity and Osteoclast Activity. *Anal. Cell. Pathol.* 2018, 2018, 8047610.
56. Jiao, Y.; Darzi, Y.; Tawaratsumida, K.; Marchesan, J.T.; Hasegawa, M.; Moon, H.; Chen, G.Y.; Nunez, G.; Giannobile, W.V.; Raes, J.; et al. Induction of bone loss by pathobiont-mediated Nod1 signaling in the oral cavity. *Cell Host Microbe* 2013, 13, 595–601.
57. Park, O.J.; Kim, J.; Yang, J.; Yun, C.H.; Han, S.H. Muramyl Dipeptide, a Shared Structural Motif of Peptidoglycans, Is a Novel Inducer of Bone Formation through Induction of Runx2. *J. Bone Miner. Res.* 2017, 32, 1455–1468.
58. Cortes-Penfield, N.W.; Kulkarni, P.A. The History of Antibiotic Treatment of Osteomyelitis. *Open Forum Infect. Dis.* 2019, 9, ofz181.
59. Handel, A.; Margolis, E.; Levin, B.R. Exploring the role of the immune response in preventing antibiotic resistance. *J. Theor. Biol.* 2009, 256, 655–662.
60. Chang, M.C.; Chen, Y.J.; Lian, Y.C.; Chang, B.E.; Huang, C.C.; Huang, W.L.; Pan, Y.H.; Jeng, J.H. Butyrate Stimulates Histone H3 Acetylation, 8-Isoprostanate Production, RANKL Expression, and Regulated Osteoprotegerin Expression/Secretion in MG-63 Osteoblastic Cells. *Int. J. Mol. Sci.* 2018, 19, 4071.
61. Montalvany-Antonucci, C.C.; Duffles, L.F.; de Arruda, J.A.A.; Zicker, M.C.; de Oliveira, S.; Macari, S.; Garlet, G.P.; Madeira, M.F.M.; Fukada, S.Y.; Andrade, I., Jr.; et al. Short-chain fatty acids and FFAR2 as suppressors of bone resorption. *Bone* 2019, 125, 112–121.
62. Morozumi, A. High concentration of sodium butyrate suppresses osteoblastic differentiation and mineralized nodule formation in ROS17/2.8 cells. *J. Oral Sci.* 2011, 53, 509–516.
63. Lucas, S.; Omata, Y.; Hofmann, J.; Bottcher, M.; Iljazovic, A.; Sarter, K.; Albrecht, O.; Schulz, O.; Krishnacoumar, B.; Krönke, G.; et al. Short-chain fatty acids regulate systemic bone mass and protect from pathological bone loss. *Nat. Commun.* 2018, 9, 55.
64. Wallimann, A.; Hildebrand, M.; Groeger, D.; Stanic, B.; Akdis, C.A.; Zeiter, S.; Richards, R.G.; Moriarty, T.F.; O'Mahony, L.; Thompson, K. An Exopolysaccharide Produced by *Bifidobacterium longum* 35624(R) Inhibits Osteoclast Formation via a TLR2-Dependent Mechanism. *Calcif. Tissue Int.* 2021, 108, 654–666.
65. Zanchetta, P.; Lagarde, N.; Guezennec, J. A new bone-healing material: A hyaluronic acid-like bacterial exopolysaccharide. *Calcif. Tissue Int.* 2003, 72, 74–79.
66. Velasco, C.R.; Baud'huin, M.; Sinquin, C.; Maillasson, M.; Heymann, D.; Collicet-Jouault, S.; Padrines, M. Effects of a sulfated exopolysaccharide produced by *Alteromonas infernus* on bone biology. *Glycobiology* 2011, 21, 781–795.
67. Yamamoto, S.; Mogi, M.; Kinpara, K.; Ishihara, Y.; Ueda, N.; Amano, K.; Nishihara, T.; Noguchi, T.; Togari, A. Anti-proliferative capsular-like polysaccharide antigen from *Actinobacillus actinomycetemcomitans* induces apoptotic cell death in mouse osteoblastic MC3T3-E1 cells. *J. Dent. Res.* 1999, 78, 1230–1237.
68. Drake, M.T.; Clarke, B.L.; Khosla, S. Bisphosphonates: Mechanism of action and role in clinical practice. *Mayo Clin. Proc.* 2008, 83, 1032–1045.
69. Hanley, D.A.; Adachi, J.D.; Bell, A.; Brown, V. Denosumab: Mechanism of action and clinical outcomes. *Int. J. Clin. Pract.* 2012, 66, 1139–1146.
70. Nikitovic, D.; Kavasi, R.M.; Berdiaki, A.; Papachristou, D.J.; Tsiaouassis, J.; Spandidos, D.A.; Tsatsakis, A.M.; Tzanakakis, G.N. Parathyroid hormone/parathyroid hormone-related peptide regulate osteosarcoma cell functions: Focus on the extracellular matrix (Review). *Oncol. Rep.* 2016, 36, 1787–1792.
71. Kyrgidis, A.; Toulis, K.A. Denosumab-related osteonecrosis of the jaws. *Osteoporos. Int.* 2011, 22, 369–370.
72. Woo, T.; Adachi, J.D. Role of bisphosphonates and calcitonin in the prevention and treatment of osteoporosis. *Best Pract. Res. Clin. Rheumatol.* 2001, 15, 469–481.